

Articles

Neutron Spin–Echo Study of Dynamics of Hydrophobically Modified Polymer-Doped Surfactant Bilayers

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We characterize the effect of adsorbed hydrophobically modified polymers (hm-polymers) on the dynamics of surfactant bilayers by small-angle neutron scattering and neutron spin–echo spectroscopy. Two kinds of hm-polymers, (a) hydrophobically modified poly(acrylate) with tetradecyl (C₁₄) sidegroups randomly grafted to the poly(acrylate) backbone and (b) poly(PEG_{6k}-lysine-stearylamine) with equally spaced hydrophobes, are added to bilayers of penta(ethylene glycol) dodecyl ether (C₁₂E₅) and hexanol. Both bare and polymer-doped membranes exhibit a stretched exponential relaxation in the form of $S(\vec{q}, t) \approx S(\vec{q}) \exp[-(\Gamma_q t)^{2/3}]$, where $S(\vec{q})$ is the static structure factor and Γ_q is the relaxation rate. The relaxation rate depends subtly on the surface coverage of polymer. At high surface coverage, polymer slows down the relaxation by 20%, whereas at low surface coverage, polymer enhances the rate relative to that of the bare membranes. Hindered flow of solvent through the adsorbed polymer layer at high coverage appears to explain the retardation. The faster dynamics at low polymer coverage may be due to stiffening of the membrane or lateral diffusion of the dilute adsorbed polymer chains.

Introduction

Polymers confined in lyotropic lamellar phases have received theoretical^{1,2} and experimental^{3–7} attention. Polymers change intermembrane (e.g., osmotic compressibility and phase separation) and intramembrane interactions (e.g., membrane flexibility and the mean curvature). Accordingly, new phase behavior is expected.^{8,9} The effect of polymers on the bilayer properties depends on both the molecular structure of the polymer and the polymer concentration. Therefore, it is possible to tune or to stabilize a particular state of surfactant organization by manipulating the polymer structure and concentration.

We would expect that the polymer would affect membrane dynamics by (1) increasing bilayer mean bending modulus κ , which would speed up the relaxation, (2)

increasing the repulsive bilayer interaction, which would speed up the relaxation, and (3) enhancing the dissipation in the adsorbed polymer brush layer, which would slow down the relaxation. Theories regarding the effect of polymer on the bending modulus κ are contradictory. Hiergeist and Lipowsky¹⁰ suggested that polymers “anchored” to the membrane would increase κ . In contrast, Brooks et al. claimed that polymer “adsorption” to the membrane would decrease κ .³ Zilman and Granek¹¹ found that the relaxation rate of the surfactant bilayer is inversely proportional to $\kappa^{1/2}$, unlike the prediction of hydrodynamic theories which indeed predict the direct proportionality of the relaxation to κ . A possible softening of the membrane due to adsorbing polymer may be considered. Hence, the observation of an increased relaxation rate of the membrane in the presence of adsorbed polymers, compared to that of the bare membrane, would confirm the result of Zilman and Granek. However, we have seen that the Zilman–Granek approximation works relatively well only for $\kappa \gg 1 k_B T$. Some researchers relate κ to the degree of homogeneity of the spontaneous curvature, that is, the polymer density on the surface.^{12,13} Our previous study on bicontinuous microemulsion phases with amphiphilic block copolymers anchored to the surfactant monolayer has shown that the most significant contribution on the local (high \vec{q}) dynamics, observed by neutron spin–echo, comes from the friction (viscosity) and the surfactant concentration (membrane area per unit

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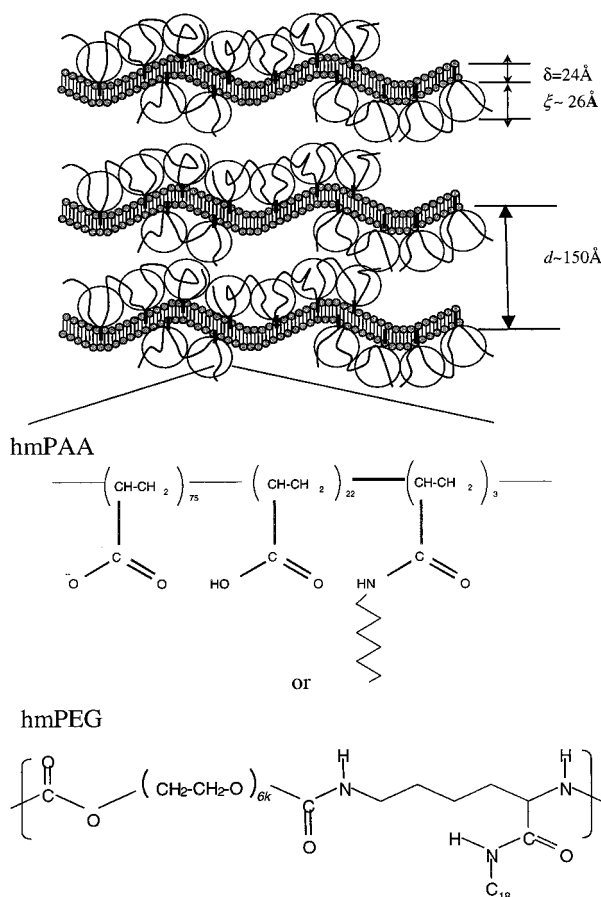


Figure 1. Sketch of the anticipated structure of the mixture of hydrophobically modified polymer and surfactant lamellar phase. The length scales are for the system of membrane volume fraction $\phi = 0.3$ doped with hmPAA with 3% hydrophobe substitution. The enlarged pictures are the chemical composition of hmPAA and hmPEG.

volume), and no polymer effect is observed in this \bar{q} range. An effect of the anchored polymer density is only observed in the hydrodynamic, low \bar{q} , region via dynamic light scattering (DLS) measurements. There a speeding up of the effective diffusion with the polymer density (at constant structure size) suggests increased repulsive interaction between layers.¹⁴

We have previously characterized the changes induced by hydrophobically modified polymers (hm-polymers) on the static elastic properties (bilayer mean bending modulus κ and layer compression modulus \bar{B}) of a nonionic membrane system using small-angle neutron scattering (SANS).^{9,15} Two hm-polymers have been investigated: (a) hydrophobically modified poly(acrylate) (hmPAA) with tetradecyl (C_{14}) sidegroups randomly grafted to the poly(acrylate) backbone and (b) poly(PEG_{6k}-lysine-stearyl-amide) (hmPEG) with equally spaced hydrophobes.^{9,15–18} Figure 1 sketches the structure of the interacting hydrophobically modified polymer and surfactant lamellar bilayer. The lamellar phase conformation has been confirmed by our previous SANS study.^{9,15} The hm-

polymers are associated with the two-dimensional membrane through their hydrophobic anchoring sites. The system can be thought of as the two-dimensional surfactant membrane doped with the strings of blobs formed by the hydrophilic segments ("loops") between hydrophobes. The interlamellar spacing d is obtained from the Bragg peak position and structure factor of the scattering spectrum. The membrane thickness δ is calculated from the form factor of the scattering spectrum. The size of a blob is determined by the molecular weight of the loop. Our previous studies found that the "rigidity" (the product of κ and \bar{B}) and layer compression modulus \bar{B} of the membranes are independent of hydrophobe length, molecular weight, and polydispersity of the hm-polymers but increase with polymer concentration and hydrophobe substitution level.^{9,15,19} Nevertheless, the dynamics and exact value of bending modulus κ of surfactant bilayers, with and without polymer doping, cannot be measured through the static scattering. In our previous studies, we observed contradictory results when membrane rigidity was calculated from SANS data using the peak divergence analysis^{20,21} and the excess area method.²² Measuring the membrane dynamics directly should help clarify this discrepancy.

The aim of this paper is to systematically study the effect of membrane volume fraction and addition of hm-polymers on the dynamic behavior of the surfactant bilayer using SANS and neutron spin-echo (NSE) spectroscopy, respectively. The so-called excess area method of Roux et al.²² is used to extract the bending modulus κ from SANS data. The dynamic structure factors of a surfactant bilayer, with or without hm-polymer doping, are given by the NSE spectroscopy. The static and dynamic measurements were compared for both bare membrane and polymer-doped membrane systems. We had anticipated a correspondence between increased membrane stiffness (from SANS) and increased relaxation rates of thermal fluctuations (from NSE). However, the response is more complicated. The limitations of models relating measured SANS and NSE quantities to membrane properties are discussed. The first section of this paper describes the materials and the experimental techniques applied in this study. The second section describes the static and dynamic data on bare and polymer-doped membranes. The final section summarizes the highlights of this study.

Experiments

Systems and Materials. The surfactant lamellar phase consists of the nonionic surfactant penta(ethylene glycol) dodecyl ether (C_{12}E_5 ; >99%, Nikko Chemical Co. Ltd., Tokyo) and 1-hexanol (>99%, Fluka), used as received. Pure C_{12}E_5 /water mixtures have a wide L_α phase region at 60 °C, spanning a membrane volume fraction ϕ from as low as 0.005 to order unity.²³ Adding hexanol to the membrane reduces the rigidity²⁴ and extends the wide lamellar range of the phase diagram to room temperature. The C_{12}E_5 /hexanol molar ratio in all of our samples is fixed at 1:1.43, which reduces the membrane rigidity to $k_B T$.²⁴ The solvent phase is 0.1 M NaCl(D_2O), which effectively screens the electrostatic interactions of the added anionic polymers; the Debye length l_D is 10 Å for the NaCl solution at 0.1 M. In the

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Table 1. Summary of Parameters for Bare Membrane and Polymer Doped Membrane Systems Studied.

	bare membrane		polymer-doped membrane			
			hmPAA		hmPEG	
	$\phi = 0.2$	$\phi = 0.3$	$\phi = 0.2$ $C_p = 2 \text{ wt } \%$	$\phi = 0.3$ $C_p = 2 \text{ wt } \%$	$\phi = 0.3$ $C_p = 0.5 \text{ wt } \%$	$\phi = 0.3$ $C_p = 2 \text{ wt } \%$
N_b			208		24	
ξ (Å)			26		43	
d (Å)	144	99	154	103	103	104
δ (Å)	24	24	24	24	24	24
σ	0	0	0.82	0.54	0.07	0.30

brine solution, the hmPAA can be viewed as an almost neutral chain with an average number of 3500 (for $M_w = 250 \text{ kg/mol}$) backbone monomers. When the electrostatic repulsion is screened with brine, the undulation force is the dominant long-range repulsion. HmPEG is nonionic.

HmPAA is synthesized by the graft reaction of precursor poly-(acrylic acid) with *n*-tetradecylamine (C_{14}) in the presence of 1,3-dicyclohexylcarbodiimide (DCC), using *n*-methylpyrrolidinone (NMP) as the solvent.²⁵ The precursor PAA (Aldrich; $M_w = 250 \text{ kg/mol}$, $M_w/M_n \sim 2$) in a 35 wt % aqueous solution was freeze-dried before use. Modified polymers were precipitated and recovered from the solution by adding concentrated NaOH (40 wt %) aqueous solution. The modified polymers have the same polymerization degree as the precursor with a typical structure illustrated in the enlarged picture in Figure 1. The degree of neutralization of the hmPAAs is $75 \pm 1.5 \text{ mol } \%$, determined by elemental analysis, and the hydrophobe substitution level is 3 mol %, determined by ^1H NMR. The distribution of the hydrophobic side chains (C_{14}) along the PAA backbone is random.²⁶

The details of synthesis and characterization of hmPEG are reported elsewhere.¹⁷ This involves synthesis of (a) lysine stearylamine dihydrochloride, (b) bis(succinimidylloxycarbonyl PEG) (BSC-PEG),^{27,28} and (c) poly(PEG_{6k}-lysine-stearylamine), sequentially. Lysine stearylamine dihydrochloride was polymerized interfacially with BSC-PEG in the presence of potassium carbonate according to a published procedure.²⁷ The structure of the synthesized polymers was characterized by ^1H NMR. Molecular weight is determined by GPC and calculated relative to poly(ethylene oxide) standards. The weight-averaged molecular weight of PEG spacers (m_b) is 6 kg/mol, and the hydrophobe is $C_{18}H_{37}$. The total molecular weights M_w and M_n of hmPEG are 112 and 78 kg/mol, respectively. That corresponds to average of 13 hydrophilic loops per polymer chain, calculated by M_w/m_b .

The interlamellar spacing d and membrane thickness δ were determined by small-angle neutron scattering. To calculate the surface coverage of hm-polymers on the surfactant membrane, we regard the anchored hm-polymer as a swollen planar coil of blobs composed of N_b loops with average blob size ξ . The details for the calculation of N_b and ξ were reported elsewhere.^{9,15} The total membrane area (per unit volume) is

$$A_{\text{membrane}} \approx \frac{2}{d} \approx \frac{2\phi}{\delta} \quad (1)$$

The blobs on the surfactant membranes cover an area (per unit volume) of

$$A_{\text{polymer}} \approx \frac{\text{no. of chains}}{\text{volume}} \frac{\text{blobs area}}{\text{chain blob}} \approx \left(\frac{C_p N_A}{M_n} \right) N_b \xi^2 \quad (2)$$

where C_p is polymer concentration and N_A is Avogadro's number.^{9,15,19} The surface coverage of polymer on the membrane is calculated by

$$\sigma = \frac{A_{\text{polymer}}}{A_{\text{membrane}}} \quad (3)$$

Table 1 summarizes the geometry and surface coverage of the systems applied in this study.

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Sample Preparation. The samples were prepared by mixing surfactant ($C_{12}E_5$), alcohol ($C_6\text{OH}$), and stock solutions of the hm-polymers by weight in 0.1 M NaCl(D_2O). In the following, we define the "membrane volume fraction" ϕ as the ratio of the volume of the $C_{12}E_5$ plus $C_6\text{OH}$ divided by the total sample volume. For conversion from weight to volume fractions, the following densities (g/cm^3) have been used: 0.996 ($C_{12}E_5$), 0.820 ($C_6\text{OH}$), 1.105 (D_2O), and 0.998 (H_2O), neglecting the effect of the added polymers on the solvent density.

Small-Angle Neutron Scattering. The small-angle neutron scattering experiments were performed at Argonne National Laboratory at Argonne, IL, with the time-of-flight small-angle diffractometer (SAD). The scattering vector (q) varied from 0.005 to 0.35 \AA^{-1} . Samples were held in 1 mm path length quartz cells. The data were processed according to the standard procedures, including the subtraction of scattering from the solvent as background and empty cell contributions.⁹ Particular attention has been paid to maintaining the lamellar samples in a polycrystalline state to obtain a powder-average. Prior to every neutron run, samples were quenched to produce small liquid crystal domain size by first raising the temperature into the isotropic sponge phase range and then quickly immersing in a water bath at $T = 25^\circ\text{C}$. Isotropic two-dimensional spectra are then obtained, and duplicate runs indicated no sensitivity of the d -spacing to quenching details.

Neutron Spin-Echo. The NSE experiments were performed at Institut für Festkörperforschung—Forschungszentrum in Jülich, Germany. The neutron spin-echo spectrometer is a high-resolution instrument suited for the study of relaxation-type motion induced by thermally excited density fluctuations in the system. It may also be viewed as an augmentation of small-angle scattering with inelastic, dynamical information. The Jülich NSE spectrometer uses cold neutrons issued from the H_2 cold source in the D_2O -moderated, 20 MW reactor. A Dornier velocity selector 10% fwhm (full width at half-maximum) in wavelength spread and a selective polarizing multilayer mirror provide polarized neutrons with a wavelength of 8 \AA . Two water-cooled copper solenoids with a maximum field integral of 0.5 T m (Tesla times meter) produce the precession fields for the neutrons. A cobalt-titanium supermirror assembly performs the final polarization analysis. A larger area converging collimator in front of the analyzer is used to reduce the background. The neutrons are detected by a Helium-3 array detector made of 32×32 cells with a total area of 1 cm^2 . The details of the NSE principles and instrument were described in detail elsewhere.^{29,30}

The individual neutron-spin precession in a magnetic field before and after scattering is used as timekeeper to measure the velocity change of the neutron during the scattering. This allows for a broad incoming neutron velocity band (10–20%) as usual in SANS, yielding reasonable intensity simultaneously with sensitivity to relative velocity changes in the 10^{-4} – 10^{-6} range. The spin-echo method directly measures the normalized intermediate dynamic structure factor $S(q, t)/S(q, 0)$, which is proportional to the cosine Fourier transform of the scattering

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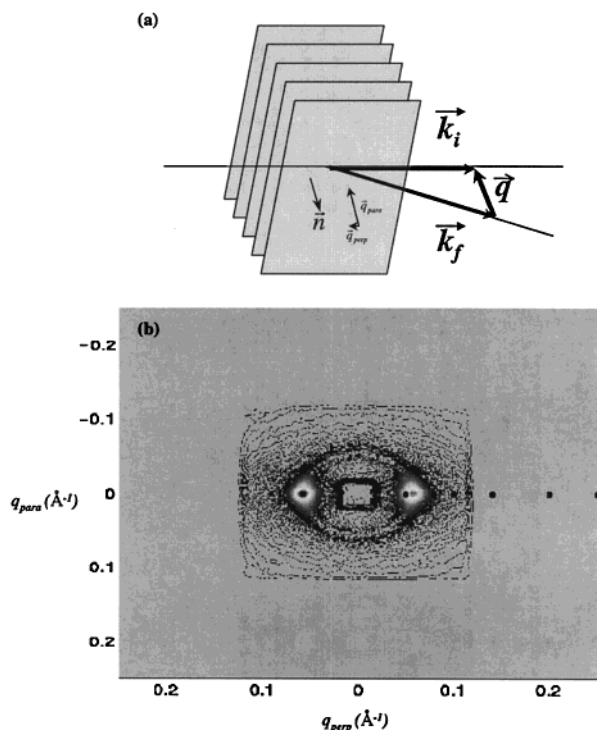


Figure 2. (a) Orientation of the ordered lamellae, incident neutron beam \vec{k}_i , scattered beam \vec{k}_f , and scattering vector \vec{q} . (b) Two-dimensional SANS intensity map from an oriented lamellar sample ($\phi = 0.3$ with 2 wt % hmPAA) taken at the small-angle neutron scattering instrument (KWS-2) at IFF, Jülich, Germany. Note that q_{para} is the component of \vec{q} parallel to the normal \vec{n} (often called q_z); q_{perp} is the component perpendicular to the normal, i.e. parallel to the membrane.

function $S(q, \omega)$. The information on the investigated system is gained through the scattering length density correlator $\langle \delta\rho(\vec{r}, t) \delta\rho(\vec{r}', 0) \rangle$ by

$$S(\vec{q}, t) \sim \int_{\vec{r}, \vec{r}'} \langle \delta\rho(\vec{r}, t) \delta\rho(\vec{r}', 0) \rangle \exp[i\vec{q}(\vec{r} - \vec{r}')] d^3\vec{r} d^3\vec{r}' \quad (4)$$

where $\delta\rho(\vec{r}, t)$ describes the density fluctuations, at position \vec{r} and time t , from the average scattering length density $\rho(\vec{r}, t)$.^{29,30}

The lamellar phase samples were oriented by injecting the mixtures into a specially designed sample cell, which consists of a stack of very thin lamellar quartz plates ($0.2 \times 5 \times 45$ mm) in a stainless steel frame (Figure 2a). The spacing between quartz plates is 0.1 mm. The orientation was achieved through the proximity of the neighboring quartz plates and by shear during injection. All the experiments were performed at $T = 25$ °C. The quality of the orientation was checked by a preliminary measurement with the SANS instrument (KWS-2). The dynamic experiments were performed immediately after on the NSE spectrometer. The orientation of the lamellar stack relative to the scattering plane was chosen so that the momentum transfer vector \vec{q} is perpendicular to the lamellar membrane (Figure 2a). In Figure 2b, the two-dimensional SANS intensity map is shown for the sample of $\phi = 0.2$ doped with 2 wt % hmPAA. From the two-dimensional SANS intensity map (Figure 2b), adequate intensities for NSE were obtained for $q < 0.2$ \AA^{-1} . NSE experiments have therefore been performed for $q = 0.05, 0.08, 0.1, 0.14, 0.2$, and 0.24 \AA^{-1} to cover dynamics from just below the Bragg peak to well beyond it.

Instrumental resolution effects were corrected by the division

$$\frac{S(\vec{q}, t)/S(\vec{q}, 0)_{\text{sample}}}{S(\vec{q}, t)/S(\vec{q}, 0)_{\text{reference}}} \quad (5)$$

where a sample holder (without quartz plates) filled with carbon powder served as a reference. Due to the high scattering intensity of the sample, background subtraction was not necessary and

not performed. The evaluation of $S(\vec{q}, t)/S(\vec{q}, 0)$ was performed for each 4×4 cell group of the detector, individually. The results were then summed into several \vec{q} groups (rings) for each scattering arm angle corresponding to the q values mentioned above.

Results and Discussion

Bare Membrane. To interpret the dynamics of the membrane systems Zilman et al. considered an ensemble of membrane plaquettes at random orientations and calculated the effect from the undulations bending energy and standard hydrodynamic dispersion law.¹¹ They predict a stretched exponential relaxation in the form of

$$S(\vec{q}, t) \cong S(\vec{q}) e^{-(\Gamma_q t)^{2/3}}$$

where $S(q)$ is the static structure factor and Γ_q is the relaxation rate.

Figure 3 shows the normalized dynamic structure factor $S(q, t)/S(q, 0)$ for the bare membrane samples ($\phi = 0.2$ and $\phi = 0.3$). Both of our bare membrane systems exhibit a relaxation behavior that cannot be fitted by a single exponential but is well fitted by this stretched exponential. Results based on eq 6 are shown as dashed lines ($\phi = 0.2$) and full lines ($\phi = 0.3$). The $\phi = 0.2$ sample relaxes faster than $\phi = 0.3$ for the whole q ($= 0.05$ \AA^{-1}). The length scale l in the system probed by scattering vector q of NSE follows $q \sim 2\pi/l$. The dynamics of the interacting membranes is probed at $q \sim 0.04$ and 0.06 \AA^{-1} for $\phi = 0.2$ ($d \sim 150$ \AA) and $\phi = 0.3$ ($d \sim 100$ \AA), respectively. The local fluctuations of the single bilayer (membrane thickness $\delta \sim 24$ \AA) are probed at $q > 0.1$ \AA^{-1} , and in this range of q the difference in relaxation behavior becomes larger. This means that the dynamics of the single bilayer at different membrane volume fractions ($\phi = 0.2$ and $\phi = 0.3$) are different (by about 50%); membrane interactions are significant. This will be discussed in more detail below.

Polymer-Doped Membrane. The dynamics of the polymer-doped membrane systems were also investigated by neutron spin-echo spectroscopy with the same q range and orientation as those of the bare membrane experiments. Figure 4 shows the dynamic structure factor $S(q, t)/S(q, 0)$ for the samples with and without hmPAA polymer for $\phi = 0.2$. The polymer-doped system also exhibits a stretched exponential relaxation behavior, similar to that observed for the bare membrane systems. Fitted results based on eq 6 are shown as dashed lines (bare membrane) and full lines (polymer-doped membrane). At $\phi = 0.2$, the polymer-doped membrane relaxes more slowly than the bare membrane for the full range of q investigated. The difference becomes much more prominent for $q > 0.1$ \AA^{-1} where the length scales are comparable to the single bilayer thickness of 24 \AA . For the system with the same amount of hmPAA polymer ($C_p = 2$ wt %) but with more surfactant membrane area available ($\phi = 0.3$ vs 0.2), the addition of hmPAA also slows down the membrane relaxation but the effect is less significant (Figure 5), than at the higher surface coverage (Figure 4).

The role of surface coverage over the range of $\sigma = 0.07$ – 0.30 (Table 1) was studied for the hmPEG-doped membrane systems at $\phi = 0.3$ with two different polymer concentrations of 0.5 wt % (Figure 6a) and 2 wt % (Figure 6b). In both figures, the relaxation of the bare membrane system ($\phi = 0.3$) at corresponding q values is shown for comparison. At the higher surface coverage of $\sigma = 0.30$ (Figure 6b), the polymer-doped membrane relaxes more slowly than the bare membrane, which is consistent with what is observed in the hmPAA system for similar high surface coverages of $\sigma = 0.54$ – 0.82 (Figure 4 and Figure 5). Figure 6a shows the relaxation of the hmPEG system

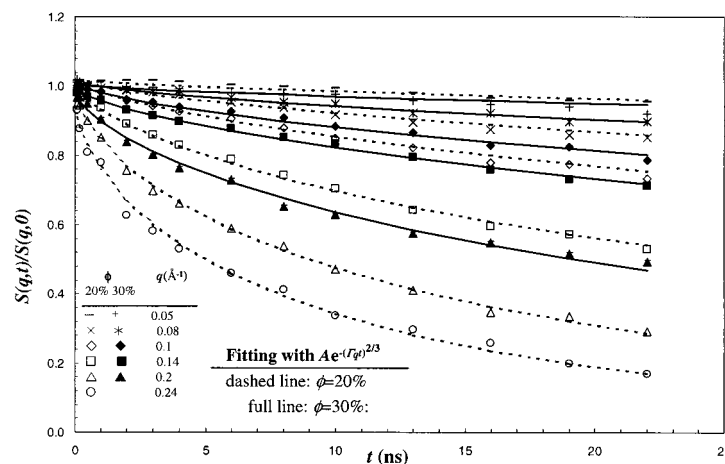


Figure 3. Normalized dynamic structure factor $S(\vec{q}, t)/S(\vec{q}, 0)$ from the NSE measurement of the bare membrane systems ($\phi = 0.2$ and $\phi = 0.3$). The lines are fitting curves to eq 6.

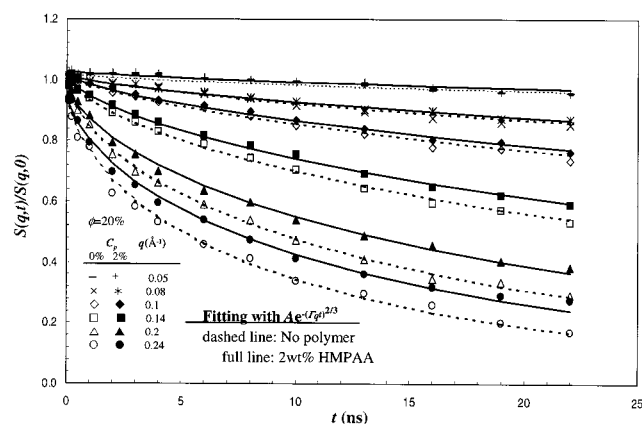


Figure 4. Normalized dynamic structure factor $S(\vec{q}, t)/S(\vec{q}, 0)$ from the NSE measurement of the bare and HMPAA-doped membrane systems at $\phi = 0.2$. The lines are fitting curves to eq 6.

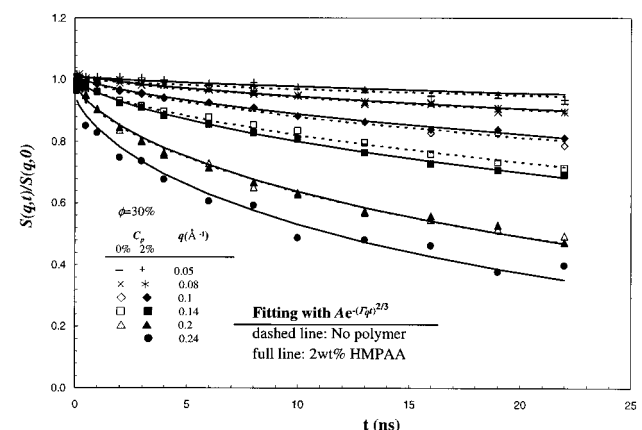


Figure 5. Normalized dynamic structure factor $S(\vec{q}, t)/S(\vec{q}, 0)$ from the NSE measurement of the bare and HMPAA-doped membrane systems ($\phi = 0.3$). The lines are fitting curves to eq 6.

at the same membrane volume fraction ($\phi = 0.3$) at low surface coverage when the adsorbed chains should be “dilute” on the surface ($\sigma = 0.07$). At $\vec{q} > 0.1 \text{ \AA}^{-1}$, where \vec{q} is probing the length scale comparable to the thickness of a single bilayer or the adsorbed chain, the polymer-doped membrane relaxes *faster* than the bare membrane.

The observation that adsorbed hm-polymers speed up membrane relaxations at low coverage and slow down relaxations at high coverage is new. At this point, we

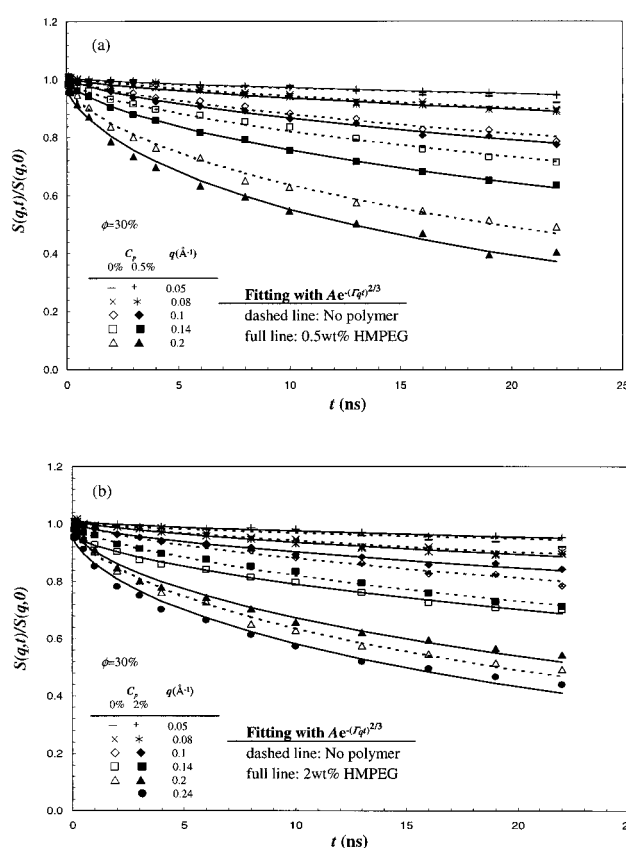


Figure 6. Normalized dynamic structure factor $S(\vec{q}, t)/S(\vec{q}, 0)$ from the NSE measurement of the bare and HMPAA-doped membrane systems at $\phi = 0.3$. The lines are fitting curves to eq 6. (a) $C_p = 0.5 \text{ wt } \%$; (b) $C_p = 2 \text{ wt } \%$.

propose the following physical model. There are two possibilities for the “speeding up”: (1) Polymers make the membrane stiffer,^{9,15,31} and therefore the relaxations are faster (as predicted from the dispersion relation for membrane undulations, $\Gamma_q \propto \kappa^{32,33}$) (Figure 7a). (2) There is a coupling between lateral polymer diffusion on the membrane surface and the relaxation of undulations in

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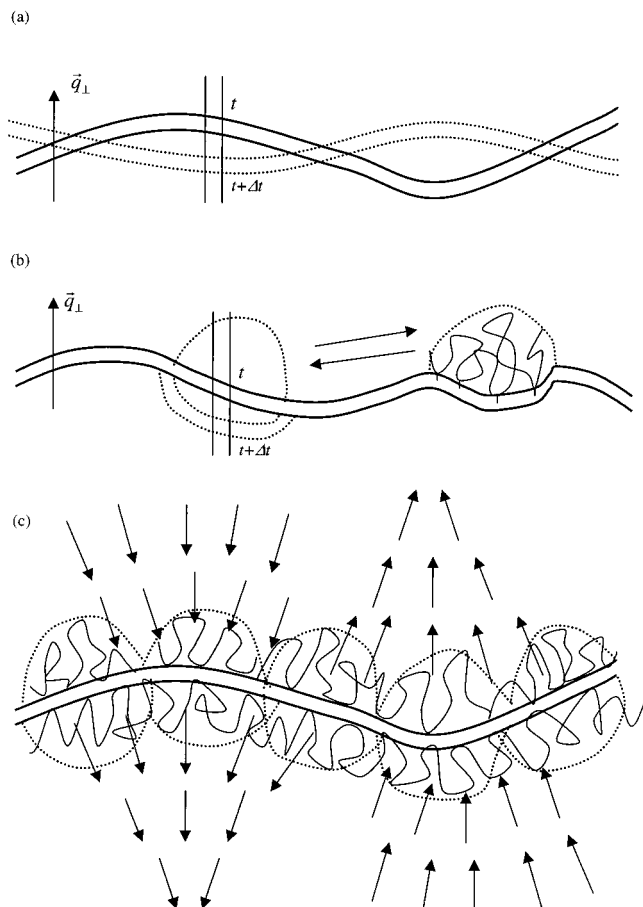


Figure 7. Sketch of the proposed polymer surface coverage effect on the membrane relaxation.

the low surface coverage case. The polymer diffusing across the membrane surface would “iron out” fluctuations causing a decrease in correlations (Figure 7b). This mechanism would apply only if the two-dimensional diffusion time is comparable to the observed \bar{q}_\perp fluctuation time scales. The lateral diffusion coefficient of polymer (anchor) on the two-dimensional membrane is $D_{\text{lateral}} = 10^{-7} - 10^{-6} \text{ cm}^2/\text{s}$.³⁴ Therefore, the relaxation time of polymer lateral diffusion $\tau_{\text{L,diff}} \sim \delta^2/D_{\text{lateral}} \sim 60 - 600 \text{ ns}$, which is comparable to the membrane relaxations probed by NSE (τ_{overall} : 10–1500 ns).

The slowing down of the membrane relaxation at high polymer coverage may arise from increased dissipation due to flow of solvent through the polymer layer (Figure 7c). Bending the polymer brush attached to the bilayer necessitates a flux of solvent into the polymer layer in regions where it is locally stretched and out of regions where it is compressed (Figure 7c). The problem has been addressed in the context of the deflection of sol–gel layers by Scherer.³⁵ The permeability of the polymer brush determines the resistance to flow, and the elasticity of the bilayer with polymer determines the elastic restoring force. This hydrodynamic relaxation time is given by³⁵

$$\tau_{\text{hyd}} = \frac{1}{2}(1 - \nu) \frac{\eta_L \xi^2}{DH} \quad (7)$$

where η_L is the viscosity of the liquid, ν (~ 0 for the polymer brush) is the Poisson ratio, D is the permeability, and H

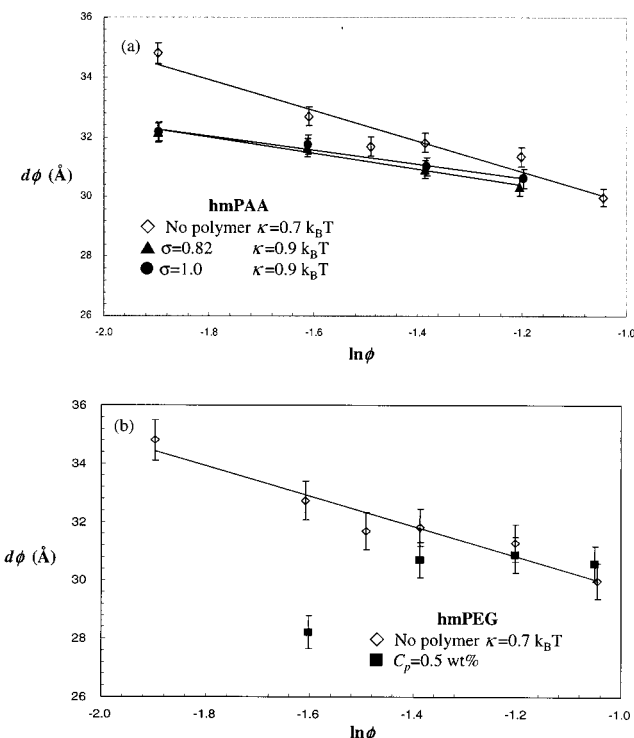


Figure 8. The measured values of $d\phi$ vs $\ln \phi$ over the whole dilution range for (a) the hmPAA systems with different surface coverages and (b) the hmPEG system.

$= (1 - \nu)E/[(1 + \nu)(1 - 2\nu)]$ is the longitudinal modulus of the polymer layer, with E being the Young's modulus (assumed equal to that of the surfactant membrane). The permeability of the PAA layer can be estimated by $D \sim \eta_L/(2.1 \times 10^{12}\omega^{3/2})$, with ω being the mass fraction of polymer in the brush layer.³⁶ The Young's modulus E is related to the bending modulus κ by $\kappa = E\delta^3/12(1 - \nu_m^2)$, where ν_m is the Poisson ratio of the surfactant membrane, which is ~ 0.5 assuming the membrane is incompressible.^{37,38} For the hmPEG-doped membrane system at $\phi = 0.3$ and $C_p = 2 \text{ wt } \%$, the estimated hydrodynamic relaxation time τ_{hyd} for solvent to flow through the polymer layer is $\sim 900 \text{ ns}$, assuming the bending modulus $\kappa \sim 1 k_B T$.^{9,15,31} This time scale is again comparable to that of the relaxation of polymer-doped membranes, which supports the hypothesis of slowed dynamics due to viscous dissipation in the adsorbed polymer layer.

Excess Area Method. The bending modulus κ can be extracted from SANS data using the excess area method.²² The relationship between the interlamellar distance d , membrane volume fraction ϕ , and membrane thickness δ depends on the bilayer mean bending modulus κ according to^{19,22}

$$\frac{\phi d}{\delta} = 1 + \frac{1}{4\pi(\kappa/k_B T)} \ln \left[\frac{\delta}{\phi b} \sqrt{\frac{32(\kappa/k_B T)}{3\pi}} \right] \quad (8)$$

where b is a microscopic cutoff length taken to be 7 \AA .^{19,24} The second term in the right-hand side of eq 8 represents the “excess area” associated with undulations in the membrane bilayer.

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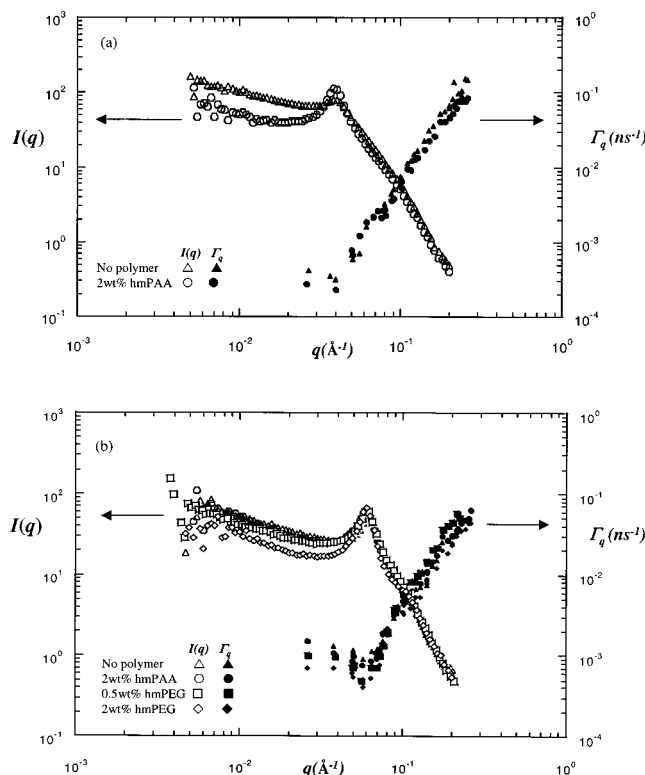


Figure 9. The \bar{q} dependence of relaxation rate Γ_q obtained from the fitting to eq 6 and corresponding SANS spectrum for the bare and polymer-doped membrane systems: (a) $\phi = 0.2$; (b) $\phi = 0.3$.

The interlamellar distance d as a function of the membrane volume fraction ϕ was determined from the first-order Bragg peak position q_0 of the small-angle neutron scattering spectra; $d = 2\pi/q_0$ at constant hexanol/ $C_{12}E_5$ mole ratio ($=1.43$) and temperature (25°C). For these conditions, the membrane volume fraction was varied ($0.15 \leq \phi \leq 0.35$) within the single lamellar phase region. The best fit of the data with eq 8 gives the membrane thickness δ and the bending modulus κ . Increasing the polymer concentration at constant membrane volume fraction increases the polymer surface coverage. Figure 8a shows data for hmPAA-doped membrane samples with changing membrane volume fraction at constant surface coverage σ . Samples with two different surface coverages ($\sigma = 0.82$ and 1.0) within the homogeneous lamellar phase region yield bending constants κ of $0.9 \pm 0.2 k_B T$. However, for the hmPEG-doped membrane at low surface coverage ($\sigma < 0.06$) shown in Figure 8b, the best fit line of $d\phi$ versus $\ln \phi$ gives a positive slope, which would be interpreted as decreasing the membrane elasticity upon the addition of polymer. Our previous study using the peak divergence method to measure membrane stiffness showed that the hmPEG polymers increased the rigidity, that is κB , of the membrane phase.^{9,15} We believe that the increase in area for the polymer-doped membrane at low polymer coverage is caused by a local bending of the membrane bilayer under the tension from the multiply attached polymer chain (Figure 7b). This would account for excess area that is not necessarily related to membrane flexibility or undulations.²²

The basis of the excess area method depends on the bending modulus κ of the membrane being independent of the membrane volume fraction, so that κ can be obtained from a dilution series at several ϕ values. However, the NSE results show that membrane dynamics depends on membrane separation (i.e., ϕ) for bare membranes or for

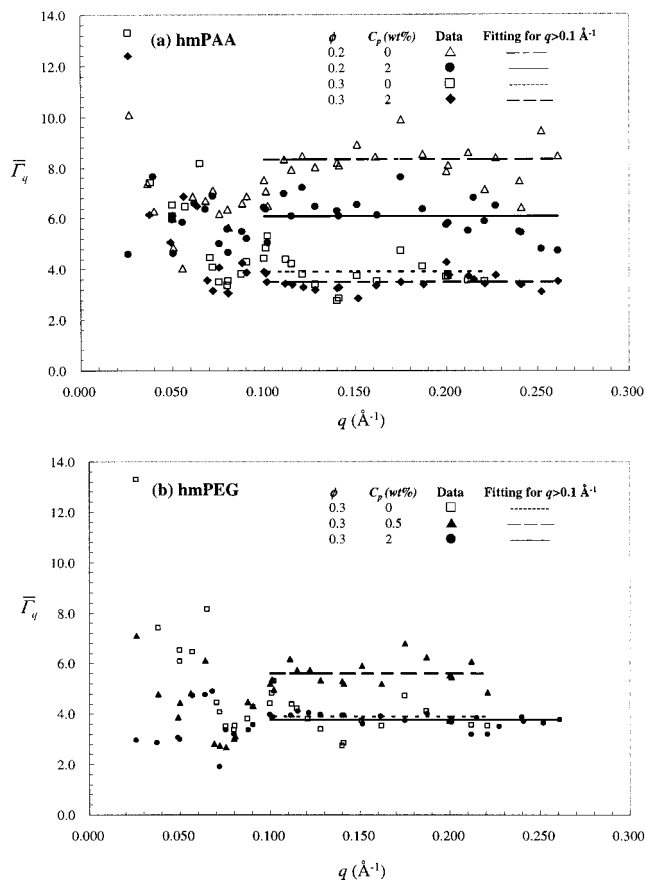


Figure 10. The q dependence of the normalized relaxation rate $\bar{\Gamma}_q = \Gamma_q S(q)/q^3$: (a) hmPAA-doped membranes; (b) hmPEG-doped membranes.

polymer-doped membranes. The compression modulus \bar{B} is significant. Therefore, for this surfactant system the elastic constant cannot be obtained from the excess area method.

Comparison between SANS and NSE Data. The relaxation rate Γ_q obtained from the fitting to eq 6 over the range of \bar{q} from 0.03 to 0.26 \AA^{-1} is shown in Figure 9. Parts a and b of Figure 9 show the data for bare and polymer-doped membranes at $\phi = 0.2$ and $\phi = 0.3$, respectively. The corresponding small-angle scattering spectra are also plotted for comparison. For q at which the interlamellar interaction is probed, the location of the minimum of Γ_q is comparable to the peak position of the static SANS spectra ($q_0 \sim 0.04 \text{ \AA}^{-1}$ for $\phi = 0.2$; $q_0 \sim 0.06 \text{ \AA}^{-1}$ for $\phi = 0.3$). This observation is consistent with the behavior of $S(q) \sim 1/\Gamma_q$.³⁹ At high q values ($q \geq 0.1 \text{ \AA}^{-1}$), which reflect smaller length scales with $S(q) \approx 1$, Γ_q scales as q^3 . The universal scaling rule of $\Gamma_q \sim q^3$ has proven accurate in both lamellar and sponge systems.^{11,32,33,40}

Parts a and b of Figure 10 show the q dependence of the normalized relaxation rate $\bar{\Gamma}_q = S(q)\Gamma_q/q^3$ for the hmPAA- and hmPEG-doped membranes and bare membranes, respectively. The normalized relaxation rate $\bar{\Gamma}_q$ is almost independent of q beyond the Bragg peak, with average values shown in lines. However, at q in the vicinity of or below the peak the normalized relaxation rates are not distinguishable. In Figure 10a, the relaxation rate with 2 wt % ($\sigma = 0.82$, eq 3 and Table 1) hmPAA is significantly slower than for the bare membrane at $\phi = 0.2$ but only

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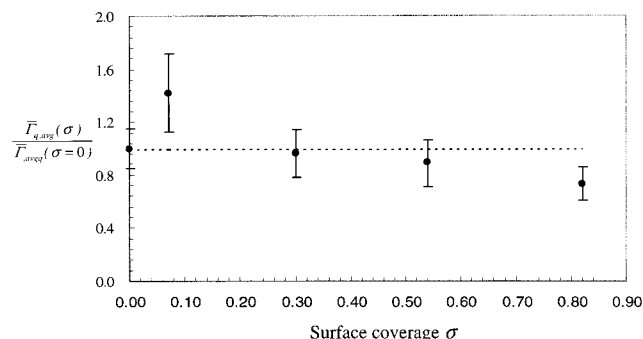


Figure 11. Dependence on the surface coverage σ of the ratio of the average normalized relaxation rate to that of the corresponding bare membrane, for both hmPAA- and hmPEG-doped membranes.

slightly slower at $\phi = 0.3$ ($\sigma = 0.54$). Figure 10b shows that membranes at $\phi = 0.3$ containing hmPAA at $C_p = 2$ wt % relax a bit slower than the bare membrane, while at $C_p = 0.5$ wt % polymer the relaxation is accelerated relative to the bare membrane. In Figure 11, we illustrate the dependence on the surface coverage σ of the ratio of the average normalized relaxation rate $\bar{\Gamma}_{q,avg}$ above $q = 0.1 \text{ \AA}^{-1}$ to that of the corresponding bare membrane for both hmPAA- and hmPEG-doped membranes. The relaxation rate depends subtly on the surface coverage of polymer, slowing down by 20% at high surface coverage and increasing slightly at low coverage, relative to the bare membranes.

Summary

This work is a small-angle neutron scattering and neutron spin-echo study of the static and dynamic behavior of surfactant bilayer membranes comprising $C_{12}E_5$ and hexanol. The effects of membrane volume fraction and the addition of hydrophobically modified polymers were investigated. We observed a difference in the relaxation behavior for two bare membrane systems ($\phi = 0.2$ and $\phi = 0.3$). This intermembrane dependence violates the assumptions inherent in “single-membrane theories” (e.g., the excess area method and the Zilman and Granek model¹¹). We are currently working on

identifying the hydrodynamic modes probed by \bar{q} in our oriented lamellar phase and developing a quantitative model of membrane dynamics that includes interlamellar interactions and single-membrane fluctuations.

Both the bare and polymer-doped membranes exhibit the relaxation behavior that is well fitted by a stretched exponential of $S(\bar{q}, t) \cong S(\bar{q})e^{-(\Gamma_{\bar{q}}t)^{2/3}}$. At high surface coverage, polymer doping slows the membrane relaxation and the effect is more prominent for large \bar{q} in which the dynamics of the single bilayer is probed. A calculation of the slowing of dynamics due to limited permeability in the adsorbed polymer brush layer shows the time scales are comparable to those observed by NSE. In contrast, polymers speed up the relaxation at low surface coverage. The increased relaxation rates at low polymer coverage are consistent with either stiffening of the membrane or lateral diffusion of the adsorbed polymers on the surface. The \bar{q} ranges covered by the static and dynamic measurements coincide. The slowest relaxation rate occurs at length scales (i.e., \bar{q} vectors) of the diffraction peak corresponding to interlamellar spacings. Fluctuations at shorter length scales (i.e., higher q values) relax at faster rates. While the theory of Zilman and Granek¹¹ predicts the $\Gamma_q \propto q^3$ relaxation rate that we observe, the theory is a single-membrane theory that assumes dynamics to be independent of interlamellar spacing. This is contradicted by our data. A dynamic theory of multilamellar membrane stacks along the lines of the Nallet et al. theory²¹ for membrane average static properties is required. Progress along those lines has been made by Ning Lei;⁴¹ however, the results are not in a tractable form at this time. The quality of the NSE data available invites the development of new theories of the dynamics of lamellar phase liquid crystalline fluids.

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