The role of abiotic processes in the formation and degradation of gaseous nitrogen compounds in the soil

Jannis Heil
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Abstract

Soils are a major source of nitrogen (N) trace gases, especially of nitrous oxide (N\textsubscript{2}O) and nitric oxide (NO). The two microbial processes nitrification and denitrification are considered the major contributors to these emissions. While microbial denitrification has long been identified as a source of N trace gases under reducing conditions, N trace gas formation under aerobic conditions is far from being completely understood. Several abiotic reactions involving the nitrification intermediates hydroxylamine (NH\textsubscript{2}OH) and nitrite (NO\textsubscript{2}\textsuperscript{−}) have been identified leading to N\textsubscript{2}O and NO emissions, but are neglected in most current studies. Further, there is a potential abiotic sink function of soils for N\textsubscript{2}O via photochemical destruction. For better N trace gas mitigation strategies, the identification of the major source and sink processes and their role in the global N cycle is vital.

Prior to the experimental work, this thesis reviews information about the role of abiotic processes in the formation of N trace gases from the few available studies reporting on abiotic emissions. It merges the gained information into a new conceptual model explaining the formation of the N trace gases N\textsubscript{2}O, NO, as well as gaseous nitrous acid (HONO) by coupled biotic–abiotic reaction mechanisms. The relevant reactions are: the self-decomposition of NO\textsubscript{2}\textsuperscript{−}, reactions of NO\textsubscript{2}\textsuperscript{−} with reduced metal cations, the nitrosation of soil organic matter (SOM) by NO\textsubscript{2}\textsuperscript{−}, the comproportionation of NO\textsubscript{2}\textsuperscript{−} and NH\textsubscript{2}OH, and the oxidation of NH\textsubscript{2}OH by manganese or iron. While reactions involving NO\textsubscript{2}\textsuperscript{−} have been shown to produce primarily NO, reactions of NH\textsubscript{2}OH are known to lead to N\textsubscript{2}O as their main product.

In soils it is difficult to discriminate between biological and abiotic processes. Here, stable isotope techniques are a promising tool to give more insight into the production processes. Especially the site preference (SP) of 15N in N\textsubscript{2}O can help to source partition between processes. Experiments have been designed to study the abiotic formation of N\textsubscript{2}O from NH\textsubscript{2}OH in solutions and in different non-sterile and sterile soils from forest, grassland, and cropland. While organic forest soils showed hardly any N\textsubscript{2}O formation upon NH\textsubscript{2}OH addition, an immediate and strong formation of N\textsubscript{2}O was observed in cropland soil, also in sterilized samples. A correlation analysis revealed a potential positive relationship of the NH\textsubscript{2}OH-induced N\textsubscript{2}O formation with soil pH and manganese content, construing an effect of pH on NH\textsubscript{2}OH stability and of manganese acting as an oxidation agent for NH\textsubscript{2}OH. A negative correlation between abiotic N\textsubscript{2}O formation and C/N ratio was found that could indicate a possible competitive reaction of NH\textsubscript{2}OH with functional groups of SOM. All abiotic N\textsubscript{2}O production pathways showed a characteristic, high SP unaffected by reaction conditions.

For studying a photochemical decomposition mechanism of N\textsubscript{2}O that could potentially act as a sink for N\textsubscript{2}O in hot desert regions of the world, experiments simulating such conditions have been conducted using a laser absorption spectrometer coupled to a flow-through reaction chamber in a
closed loop mode. However, N$_2$O decomposition could not be observed, at least not within the short timeframe and the conditions of the experiments, and thus photochemical destruction on hot siliceous surfaces could not be verified.

This thesis suggests a coupled biotic–abiotic production of N$_2$O during nitrification, which could be initialized by a leakage of the nitrification intermediate NH$_2$OH from nitrifying microorganisms with subsequent reaction in the soil matrix. This mechanism could be significant in agroecosystems showing high nitrification rates upon fertilizer application and commonly having a low organic matter content and a near-neutral pH, but further research is needed to quantify the contribution of abiotic processes to total N$_2$O emissions.
Zusammenfassung


Vor der experimentellen Arbeit werden in dieser Dissertation die verfügbaren Informationen über die Rolle abiotischer Prozesse bei der Bildung von N-Spurengasen aus den wenigen Studien zu dieser Thematik zusammengefasst und zu einem neuen konzeptionellen Modell zusammengeführt, welches die Bildung der N-Spurengase N₂O und NO sowie gasförmiger salpetriger Säure (HONO) durch gekoppelte biologisch-chemische Reaktionsmechanismen erklärt. Relevante Prozesse sind: die Selbstzersetzung von NO₂⁻, Reaktionen von NO₂⁻ mit reduzierten Metallkationen, die Nitrosierung von organischer Bodensubstanz (SOM) durch NO₂⁻, die Komproportionierung zwischen NO₂⁻ und NH₂OH und die Oxidation von NH₂OH durch Mangan oder Eisen. Während Reaktionen, an denen nur NO₂⁻ beteiligt ist, primär NO produzieren, ist N₂O das Hauptprodukt von Reaktionen mit NH₂OH.

Verhältnis gefunden, die auf eine mögliche Konkurrenzkaktion von NH₂OH mit funktionellen Gruppen der SOM hindeutet. Alle abiotischen N₂O-Bildungsprozesse zeigten eine von den Reaktionsbedingungen unabhängige, hohe positive Positionsabhängigkeit von ¹⁵N innerhalb der gebildeten N₂O-Moleküle.

Um einen photochemischen Abbaumechanismus für N₂O, der in heißen Wüstenregionen der Erde als potenzielle Senke für N₂O dienen könnte, zu untersuchen, wurden Experimente, die solche Bedingungen simulierten, mit Hilfe eines Laserabsorptionsspektrometers, welches in einem geschlossenen Kreislauf mit einer Durchflussreaktionskammer gekoppelt war, durchgeführt. Allerdings konnte im zeitlich begrenzten Rahmen und unter den gewählten Versuchsbedingungen kein signifikanter Abbau von N₂O beobachtet werden.

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<th>Description</th>
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<tbody>
<tr>
<td>Ah</td>
<td>humic mineral topsoil horizon</td>
</tr>
<tr>
<td>anammox</td>
<td>anaerobic ammonium oxidation</td>
</tr>
<tr>
<td>AOA</td>
<td>ammonia-oxidizing archaea</td>
</tr>
<tr>
<td>AOB</td>
<td>ammonia-oxidizing bacteria</td>
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<tr>
<td>C/N</td>
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</tr>
<tr>
<td>cw-QCL</td>
<td>continuous wave quantum cascade laser</td>
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<td>δ</td>
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<td>δ¹⁵N of the terminal position of N₂O</td>
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<td>δ¹⁸O</td>
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<td>DNRA</td>
<td>dissimilatory nitrate reduction to ammonium</td>
</tr>
<tr>
<td>e⁻</td>
<td>electron</td>
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<td>isotope ratio mass spectrometer</td>
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<td>quantum cascade laser absorption spectrometer</td>
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Chapter 1

Introduction
1.1. Rationale

Nitrogen (N) is a key component of all living organisms, but the vast majority of the Earth’s N, bound as diatomic nitrogen (N₂) and making up approximately 78% of the Earth’s atmosphere, is unavailable to most organisms (Galloway et al., 2004). Only few species can use N₂ and transform it into reactive N that is available to plants and animals, thus most natural ecosystems are N-limited, although evolution developed adaptive mechanisms for an efficient N use. Before the industrial revolution, the conversion of N₂ into reactive N was in equilibrium with losses of reactive N back to N₂, until the invention of the Haber-Bosch process led to an uncoupling of this equilibrium (Ciais et al., 2013). Since then, the amount of anthropogenically produced reactive N has been much higher than the amount returned back to the atmosphere as N₂, and is still increasing due to increasing use of artificial fertilizer, enabling humankind to greatly increase food production to feed the growing global population (Gruber and Galloway, 2008). However, this has led to a lot of environmental problems from eutrophication of ecosystems to acidification, as well as to the production of environmental and climate-relevant N trace gases.

Soils are a major source of nitrous oxide (N₂O), nitric oxide (NO), and N₂, with increasing tendency due to anthropogenic activities (Ciais et al., 2013). While N₂ is an inert gas and the major component of the Earth’s atmosphere, NO is a highly reactive trace gas with great environmental impact. NO and its oxidation product NO₂, subsumed as NOₓ, catalyze the formation of tropospheric ozone in the presence of volatile organic compounds (Crutzen, 1979). NO has negative impacts on human health as well as plant productivity. Plant damage from tropospheric ozone is believed to be responsible for more than $2 billion per year in crop losses in the USA alone (Delucchi et al., 1996). Natural soil and agricultural NOₓ emissions are estimated at 7.3 and 3.7 Tg N yr⁻¹, respectively, combining to 23% of total global emissions (Ciais et al., 2013). N₂O, on the other hand, is contributing significantly to stratospheric ozone destruction (Ravishankara et al., 2009) and is the fourth-most important anthropogenic greenhouse gas (Davidson, 2009), almost 300 times more potent than carbon dioxide (CO₂) and still about 12 times more potent than methane (CH₄) in a time frame of 100 years (Ciais et al., 2013). The atmospheric concentration of N₂O increased from a pre-industrial value of 270 ppb to 325 ppb in 2012 at a rate of about 0.80 ppb yr⁻¹ over the last decade (WMO, 2013). The global N₂O source strength is still highly uncertain, which is reflected in the high uncertainty of the estimate given by the IPCC in 2013, ranging from 8.1–30.7 Tg N yr⁻¹ (Ciais et al., 2013). With an estimated 50–60% of global N₂O emissions, soils – especially agricultural soils – have been identified as the major source of this potent greenhouse gas (USEPA, 2010). A globally growing demand for food and increasing use of N fertilizer will further increase emissions (Wuebbles, 2009), although new approaches for better agricultural efficiency and mitigation are being developed (Smith et al., 2007).
Microbial nitrification and denitrification are widely accepted as the major sources of these N gas emissions from soils (Ciais et al., 2013). NO and N$_2$O release during both processes has been described by Firestone and Davidson (1989) in their conceptual ‘hole-in-the-pipe’ model, but N trace gas production in soils, especially during nitrification, is far from being completely understood. The model attributes NO and N$_2$O emissions from soils during nitrification and denitrification to leaks in the N transformation from ammonium ($\text{NH}_4^+$) via hydroxylamine ($\text{NH}_2\text{OH}$) and nitrite ($\text{NO}_2^-$) to nitrate ($\text{NO}_3^-$), and to the incomplete sequential reduction of NO$_3^-$ via NO$_2^-$, NO, and N$_2$O to N$_2$. However, this model is oversimplified, as it is known that there are a variety of processes and metabolic pathways involved in soil N trace gas production. Because denitrification can both produce and consume NO and N$_2$O, an imbalance between NO or N$_2$O formation and reduction, depending on enzyme regulation, can make denitrifying bacteria net N trace gas producers or consumers. The fact that soils can, at least temporarily, function as significant N$_2$O sinks has been reported recently (Chapuis-Lardy et al., 2007; Goldberg and Gebauer, 2009).

Apart from soil bacteria, fungi can also denitrify, but largely lack N$_2$O reductase and therefore produce N$_2$O (Laughlin and Stevens, 2002). Fungi are also involved in a hybrid reaction, called codenitrification, in which inorganic and organic N precursors lead to NO or N$_2$O formation (Spott et al., 2011). Nitrifying bacteria produce N$_2$O as a side product during the oxidation of NH$_2$OH, but can also reduce nitrite under oxygen-limiting conditions or at elevated nitrite concentrations in a process similar to denitrification known as nitrifier denitrification (Poth and Focht, 1985; Wrage et al., 2001). There are more alternative processes potentially involved in N trace gas formation in soils, such as heterotrophic nitrification, dissimilatory nitrate reduction to ammonium (DNRA), and nitrification by archaea, but besides these various, widely unexplored and partially not very well understood microbial processes there are also several abiotic pathways that are known for years but are widely neglected in most current studies (Bremner, 1997; Butterbach-Bahl et al., 2013; Santoro et al., 2011; Stevens et al., 1998; Yamulki et al., 1997). Those abiotic N trace gas formation pathways include (i) chemodenitrification, i.e., the decomposition of soil NO$_2^-$ with NO as main product, but N$_2$O as minor product (van Cleemput and Samater, 1996), (ii) the abiotic decomposition of ammonium nitrate on reactive surfaces in the presence of light (Rubasinghege et al., 2011), and (iii) the oxidation of the nitrification intermediate NH$_2$OH that can be oxidized by several soil constituents to form N$_2$O (Bremner, 1997).

The occurrence of non-enzymatic NO$_2^-$ decomposition associated with gaseous N losses was proposed by Clark (1962) and referred to as chemodenitrification. Since then, the role of NO$_2^-$ decomposition in N trace gas formation has been reviewed occasionally (Chalk and Smith, 1983; Nelson, 1982; van Cleemput and Samater, 1996). However, those reviews did not try to link abiotic mechanisms to other known biotic soil processes that could potentially deliver the substrate for the abiotic N trace gas formation. Until now, it has never been attempted to merge the various biotic and abiot-
ic N transformation processes into a conceptual model explaining N trace gas formation in soils. A potential mechanism for these abiotic reactions could be a leakage of biologically produced NH$_2$OH and NO$_2^-$ out of the respective microorganisms into the soil matrix, where they could react with oxidizing (in case of NH$_2$OH) or reducing (in case of NO$_2^-$) compounds. Alternatively, both substrates could react with each other to yield N$_2$O, a reaction which has been shown to lead to N$_2$O formation (van Cleemput and Samater, 1996). These reactions could easily be overlooked because of the simultaneous activity of biological and abiotic N$_2$O source processes in close vicinity in soils. While reactions involving NO$_2^-$ are more commonly associated with NO formation, another nitrification intermediate, NH$_2$OH, is linked with the formation of N$_2$O (Bremner, 1997), as it is highly reactive and can undergo reactions with several soil constituents to form N$_2$O.

NO$_2^-$ occurs in soil as an intermediate product of microbial nitrification and denitrification, and early studies used the chemical decomposition of NO$_2^-$ to explain gaseous losses of fertilizer N from agricultural systems (Nelson and Bremner, 1970a). In their review on chemodenitrification, Chalk and Smith (1983) presented abiotic mechanisms leading to gaseous N losses, such as the self-decomposition of nitrous acid (HNO$_2$; the protonated form of NO$_2^-$), the reaction of HNO$_2$ with amino compounds and NH$_4^+$, the reaction of HNO$_2$ with soil organic matter (SOM) and the reduction of NO$_2^-$ by metal ions. Although NO$_2^-$ generally does not accumulate in soils under natural conditions due to its reactive character (Robertson and Groffman, 2007), there are situations, especially after fertilizer application, in which NO$_2^-$ can accumulate to a greater or lesser extent (Gelfand and Yakir, 2008). Therefore, N trace gas formation from abiotic reactions of NO$_2^-$ have been considered in studies occasionally (e.g., Cheng et al., 2004; Ding et al., 2010; Kesik et al., 2006; Li et al., 2000; Yamulki et al., 1997). NO$_2^-$ has long been considered to be the key intermediate for abiotic N trace gas formation (Venterea, 2007), although NH$_2$OH plays a fundamental role in the formation particularly of N$_2$O (Bremner et al., 1980). The fact that NH$_2$OH is even more short-lived than NO$_2^-$ and was for a long time non-detectable in soils, has led an omission of these NH$_2$OH-induced N trace gas formation in favor of microbial pathways. Non-detection is usually explained by the highly reactive character of NH$_2$OH (Moews and Audrieth, 1959), but another factor that led to this omission was that NH$_2$OH is generally not a free intermediate of nitrification as NO$_2^-$, i.e., NH$_2$OH is generally not assumed to be released by nitrifying microorganisms (De Boer and Kowalchuk, 2001), although a release has been reported (Schmidt et al., 2004b; Stüven et al., 1992). Since NH$_2$OH can be oxidized by several soil constituents to form N$_2$O (Bremner et al., 1980), it can be hypothesized that an underestimation of the importance of NH$_2$OH led consequently to an underestimation of abiotic N trace gas formation, especially of N$_2$O. Additionally, other novel microbial soil processes as DNRA or the anaerobic ammonium oxidation (anammox) could be potential sources of NH$_2$OH besides nitrification.
Despite the general knowledge about abiotic N trace gas formation, little is known about the magnitude of these chemical processes in the global N cycle. Especially under field conditions it is difficult to identify the processes responsible for the formation of the respective N gases, as diverse biotic and abiotic processes act simultaneously (Venterea, 2007). A better understanding of these abiotic processes and their contributions to NO, N₂O, and N₂ fluxes is needed (Gärdenäs et al., 2011), as especially changing climatic conditions, such as increasing frequency and/or intensity of drying/rewetting or freeze/thaw cycles, might increase the importance of abiotic reactions for soil N trace gas formation, e.g., NO₂⁻ accumulation with subsequent decomposition. This knowledge could improve modeling of ecosystem N cycling and constraining atmospheric greenhouse gas budgets, and will help to quantify the feedback to global climate change and other environmental problems, such as the destruction of stratospheric ozone by N₂O or tropospheric ozone formation or acid deposition by NO (Ciais et al., 2013; Crutzen, 1979; Ravishankara et al., 2009).

1.2. Objectives and outline of the thesis

The overall aim of this dissertation was to characterize abiotic N trace gas formation, processes that have been known for years but which are widely neglected in current studies. This could be due to the factors discussed above, so that it can be hypothesized that abiotic N trace gas formation is overlooked in favor of other unclear production mechanisms. This thesis aimed to clarify the role of these abiotic processes in soils. To achieve this, the existing knowledge on abiotic N trace gas formation from soils was gathered in the first instance. This review about the chemical N trace gas formation can be found in chapter two. It summarizes the over the last 50 to 60 years infrequently occurring and, in a lot of cases, overlooked studies reporting on abiotic N trace gas formation in soils and merges them into one conceptual model, revisiting the “hole-in-the-pipe” model and explaining abiotic formation of N₂O, NO and gaseous nitrous acid (HONO). Further, the second chapter emphasizes the coupling between biotic nitrification and abiotic mechanisms leading to N trace gas formation. Based on this hypothesis of a coupled biotic–abiotic N trace gas formation during nitrification, experiments were developed to demonstrate the relevance of these abiotic reactions in soils.

Because of the simultaneous occurrence of biotic and abiotic trace gas formation processes in soils, experiments were created helping to improve source partitioning the different N₂O formation processes. Stable isotopes, and especially the site-specific position of ¹⁵N inside the N₂O molecule (site preference, SP), are considered promising tools allowing to differentiate between the different N₂O production and consumption processes. The SP is defined as the difference in ¹⁵N isotope signatures (δ¹⁵N) between the central (Nα) and the terminal (Nβ) positions of the asymmetric linear N₂O molecule (Toyoda and Yoshida, 1999). While the δ¹⁵N in N₂O produced in soils is supposed
to be controlled by the isotopic signature of the substrate, the SP is assumed to represent the production process (Sutka et al., 2006). Lately, the site-specific isotopic signature of N\textsubscript{2}O from distinct microbial pathways has been studied in pure microbial populations as well as in mixed culture systems (Bol et al., 2003; Frame and Casciotti, 2010; Opdyke et al., 2009; Sutka et al., 2006; Sutka et al., 2003; Toyoda et al., 2005; Well et al., 2006; Wunderlin et al., 2013). It has been shown, that the \textsuperscript{15}N SP in N\textsubscript{2}O can be used to differentiate between the microbial formation processes nitrification and denitrification, albeit with a considerable uncertainty (Ostrom and Ostrom, 2011). However, isotopic signatures are not known for all processes, and for some known processes signatures vary over large ranges, making source partitioning using stable isotopes challenging. The study of the site-specific signature of abiotically produced N\textsubscript{2}O could be another piece in the puzzle of source partitioning. As the SP is supposed to reflect the N\textsubscript{2}O production mechanism, it can be hypothesized that if a distinct SP for abiotic N\textsubscript{2}O production was found, it could be used to distinguish between microbial and abiotic N\textsubscript{2}O production in soils and by this serve as a proof of abiotic N\textsubscript{2}O formation in soils. However, if previously observed N\textsubscript{2}O formation during nitrification had been wrongly assigned to microbial production, but is actually of abiotic origin, it could be hypothesized that the SP for abiotic N\textsubscript{2}O production could be the same as observed for microbial processes. To answer these questions, first experiments on possible abiotic reactions being discussed in literature to contribute to N trace gas formation from soils involving the nitrification intermediates NH\textsubscript{2}OH and/or NO\textsubscript{2}\textsuperscript{–} in combination with other soils constituents, were tested in aqueous solution for their production of NO and N\textsubscript{2}O using chemiluminescence detection and laser adsorption spectrometry, respectively. Based on these results, experiments have been developed to determine the \textsuperscript{15}N site-specific isotopic signature of abiotically produced N\textsubscript{2}O from identified reaction mechanisms in solutions. For this, the latest laser absorption spectroscopy was used to achieve high precision data in virtually real-time at a temporal resolution of 1 Hz. This not only allowed studying the site-specific isotopic signature of abiotically produced N\textsubscript{2}O for different abiotic reactions under various conditions, but also gave insight into isotopic fractionation over time and the kinetics of the reactions. These results are presented in chapter three. The isotopic signatures have been discussed with regard to a possible source determination and have been compared in this context with isotopic signatures of N\textsubscript{2}O produced by microbial soil processes.

As the first experiments in this thesis were designed to validate potential N\textsubscript{2}O formation processes from nitrification intermediates, to quantify N\textsubscript{2}O formation under different conditions, and to find a distinct SP of abiotically produced N\textsubscript{2}O in solutions, experiments presented in chapter four were designed to provide evidence that these abiotic processes could also occur in soils. To find evidence for abiotic N\textsubscript{2}O production from the nitrification intermediate NH\textsubscript{2}OH in soils, incubation experiments with live and sterilized soils were conducted. These experiments were designed to show: (i) the potential of soils to oxidize NH\textsubscript{2}OH abiotically, (ii) the influence of soil parameters
Introduction

on NH₂OH oxidation potential, (iii) the kinetics of NH₂OH oxidation reactions, and (iv) the isotopic signature of abiotically produced N₂O. It is known for years that transition metals, primarily iron and manganese, can oxidize NH₂OH, leading to the formation of N₂O (Bremner et al., 1980). As iron and manganese are to a lesser or greater extent constituents of most soils, it can be hypothesized that soils have a potential to oxidize NH₂OH to N₂O, depending on their content of transition metals and/or maybe other unknown constituents. Other potential influential soil parameters can be hypothesized to be soil pH, as it controls the stability of NH₂OH and kinetics of the reactions, and also SOM content, as incorporation of NH₂OH into SOM has been observed (Bremner et al., 1980; Porter, 1969) that could act as a reaction competing with the oxidation of NH₂OH. Thus, it could be assumed that high SOM contents counteract NH₂OH oxidation to N₂O. Compared to microbial reactions, abiotic N₂O production is supposedly very fast and proceeds immediately upon the availability of the substrate. After the addition of NH₂OH, a peak-like formation of gaseous products could be expected.

The isotopic signatures of N₂O from different soil processes have been shown to be dependent on the substrate, but the SP for N₂O produced during nitrification was found to be in a high positive range, independent of the substrate (Decock and Six, 2013; Sutka et al., 2006). As it was presumed that the SP depends on the production process, particularly on the last intermediate step in the formation of N₂O (Toyoda et al., 2002), it can be expected that the SP of abiotically produced N₂O from the oxidation of NH₂OH will be in a similar range. To validate the hypotheses above, the laboratory incubation experiment in this study used soils covering a wide range of land use types (cropland, grassland, and forest). Soil incubations were conducted at conditions favorable for nitrification. Incubations with and without the addition of NH₂OH solution, as well as with non-sterile and sterile soils have been conducted using gas chromatography for N₂O quantification. For the analysis of the kinetics of NH₂OH-induced N₂O formation N₂O mixing ratios where quantified online at a high temporal resolution using quantum cascade laser absorption spectroscopy. Furthermore, isotope ratio mass spectrometry was used to analyze the isotopic signatures (i.e., δ¹⁵N, δ¹⁸O, and SP) of abiotically formed N₂O. All results will be discussed regarding a potential abiotic formation of N₂O in soils and soil parameters that could have an influence on the production. This knowledge can help to verify the hypothesized abiotic N₂O production mechanisms and help to understand the influencing soil parameters.

The very long estimated atmospheric lifetime of N₂O of about 120 years is mainly due to the lack of significant N₂O sinks in the troposphere (Ciais et al., 2013). Although it has been shown lately that soils, at least temporarily, can act as N₂O sinks (Berger et al., 2013; Chapuis-Lardy et al., 2007; Goldberg and Gebauer, 2009), these sinks could be of importance on a local scale, but at a global scale soils remain net producers of N₂O at a still increasing rate due to agricultural intensification. The only known significant sink for N₂O is the photochemical or oxidative destruction in
the stratosphere, by which only less than 1% of the atmospheric N\textsubscript{2}O is annually removed (Montzka et al., 2011). The knowledge of additional sink processes in the troposphere would therefore greatly enhance our ability to constrain the global N\textsubscript{2}O budget.

Photolysis could also play a role in N\textsubscript{2}O production and destruction in the troposphere. Recently, a photochemical pathway for the production of N\textsubscript{2}O has been proposed by Rubasinghege et al. (2011). It describes the abiotic N\textsubscript{2}O production from ammonium nitrate fertilizer via photolysis at the surface in the presence of light at 298 K and some air humidity, a mechanism that has not been considered in previous N\textsubscript{2}O budgets. Lately, this process has also been used to explain the abiotic N\textsubscript{2}O formation from a hypersaline pond in Antarctica (Peters et al., 2014). Besides these photochemical processes leading to N\textsubscript{2}O production, the photochemical destruction of N\textsubscript{2}O had also been proposed in the past. The possibility of large desert areas as a possible tropospheric sink for N\textsubscript{2}O was first suggested by Junge et al. (1971), who found lower concentrations of N\textsubscript{2}O during a cruise in the Atlantic Ocean associated with air masses of West African origin. The findings of Schütz et al. (1970) at a mountain observatory on the island of Tenerife, that is almost permanently in the Saharan air layer, gave further evidence for this process. The authors found that the concentration of N\textsubscript{2}O at the mountain observatory (260 ppb) was significantly lower than compared to 320 ppb N\textsubscript{2}O at the sea level station on the island that was not in the Saharan air layer. There was no explanation for this, until Rebbert and Ausloos (1978) suggested the photochemical destruction of N\textsubscript{2}O as a possible mechanism. The authors showed that N\textsubscript{2}O can be adsorbed on very dry sand surfaces and subsequently be decomposed by photons of sunlight that N\textsubscript{2}O cannot absorb in the gas phase. Although it is known that N\textsubscript{2}O can be photochemically decomposed at elevated temperatures, Rebbert and Ausloos (1978) showed that dry particulate matter in the troposphere may be responsible for the decomposition of N\textsubscript{2}O. The reaction was shown to proceed at 23 °C with light at wavelengths greater than 280 nm, with decreasing rate of photolysis at increasing wavelength. The addition of water vapor to the system resulted in a dramatic decrease of photolysis. It was predicted, that dry desert sands may act as sinks for atmospheric N\textsubscript{2}O and that they might be able to significantly reduce the effect on the stratospheric ozone layer as has been predicted on the basis of very long tropospheric lifetimes (Pierotti et al., 1978). Therefore, final experiments were conducted to study a possible photochemical N\textsubscript{2}O decomposition mechanism over hot and dry surfaces, conditions as found in hot desert areas of the world. For this, laboratory experiments using a flow-through reaction chamber in a closed loop connected to a laser absorption spectrometer were set up. This allowed for an online monitoring of N\textsubscript{2}O mixing ratios over quartz sand as a reactive surface at hot and dry conditions and at high ultraviolet (UV) light intensity. If it was found to be relevant, this potential photochemical destruction of N\textsubscript{2}O could be a mechanism, which is currently not considered in global N\textsubscript{2}O budget calculations.
Chapter 2

A review of chemical nitrogen trace gas formation reactions of nitrification intermediates in soils

Based on:
2.1. The role of nitrification intermediates in abiotic nitrogen trace gas formation

NH$_2$OH and NO$_2^-$ are intermediates of nitrification, the biological oxidation of ammonia (NH$_3$) to NO$_3^-$. The process is carried out by two groups of microorganisms: the first step, the conversion from NH$_3$ to NO$_2^-$ by ammonia-oxidizing bacteria (AOB), the second step, the oxidation of NO$_2^-$ to NO$_3^-$, by nitrite-oxidizing bacteria (NOB) (Bock et al., 1986). Taxonomically the two groups belong to one family, the *Nitrobacteraceae* (Robertson and Groffman, 2007). *Nitrosomonas europaea* is the best studied but not necessarily most common AOB (Chain et al., 2003). *Nitrobacter winogradskyi* is the best studied representative of NOB (Bock et al., 1986). NH$_2$OH is the first of several intermediates that are formed during nitrification. The enzyme ammonia monooxygenase catalyzes the oxidation of NH$_3$ to NH$_2$OH; two electrons are required for the reduction of one atom of O$_2$ to water (Robertson and Groffman, 2007):

\[ \text{NH}_3 + 2 \text{H}^+ + \text{O}_2 + 2 \text{e}^- \rightarrow \text{NH}_2\text{OH} + \text{H}_2\text{O} \quad (1) \]

Successively, NH$_2$OH is oxidized to NO$_2^-$, catalyzed by the enzyme hydroxylamine oxidoreductase:

\[ \text{NH}_2\text{OH} + \text{H}_2\text{O} \rightarrow \text{NO}_2^- + 5 \text{H}^+ + 4 \text{e}^- \quad (2) \]

Two of the four electrons from Equation (2) are returned to the ammonia monooxygenase reaction (1) (Chain et al., 2003). Both enzymes utilized for the oxidation of NH$_3$ to NO$_2^-$ are found in bacteria of the genus *Nitrosomonas* (Parkes et al., 2007).

NO$_2^-$ is further oxidized by NOB, such as *Nitrobacter*, catalyzed by nitrite oxidoreductase in a one-step reaction to NO$_3^-$(Bock et al., 1986):

\[ \text{NO}_2^- + \text{H}_2\text{O} \rightarrow \text{NO}_3^- + 2 \text{H}^+ + 2 \text{e}^- \quad (3) \]

The nitrifying bacteria are chemolithoautotrophic, i.e., they use the chemical energy from nitrification for the fixation of CO$_2$ as their carbon (C) source. The *Nitrobacteraceae* are aerobic microorganisms, and thus require O$_2$ (Robertson and Groffman, 2007).

Besides AOB, there are ammonia-oxidizing archaea (AOA) involved in the oxidization of NH$_3$. Leininger et al. (2006) found that AOA outnumber AOB in most soils, but their contribution to nitrification cannot be inferred from plain abundance and therefore needs to be assessed. Di et al. (2009), however, assumed that AOA, although being present in great quantity, contributed little to nitrification in N-rich grassland soils. Archaea are better adapted to resource limitations than bacte-
ria, which could give archaea competitive advantages over bacteria in unfavorable conditions, such as low nutrient availability and/or low or high soil pH (Di et al., 2009; Valentine, 2007).

Additionally, a variety of heterotrophic bacteria and fungi can oxidize NH$_3$ (De Boer and Kowalchuk, 2001). There are two pathways proposed for heterotrophic NH$_3$ oxidation (Kuenen and Robertson, 1994). In the first, heterotrophic bacteria use similar enzymes as their autotrophic counterparts (Moir et al., 1996). The second pathway seems to be limited to fungi and involves a reaction of N compounds with hydroxyl radicals formed in the presence of hydrogen peroxide and superoxide, which can occur during cell lysis and lignin degradation by fungi (De Boer and Kowalchuk, 2001).

Even though NO$_2^-$ does usually not accumulate in soils under natural conditions (Robertson and Groffman, 2007), it plays a key role in the global N cycle, as it is formed as a free intermediate in both aerobic nitrification and anaerobic denitrification processes (van Cleemput and Baert, 1984). As an anion, NO$_2^-$ is very mobile and constitutes a fundamental link between different N transformation processes in different soils under diverse environmental conditions (Kappelmeyer et al., 2003). It has been expected for a long time that the NO$_2^-$ anion is the key intermediate between the solid or solute and gaseous states of N (Wullstein and Gilmour, 1966). The first nitrification intermediate, NH$_2$OH, also plays a fundamental role, although it is usually not released by the bacteria into the soil matrix (Bremner et al., 1980). AOB transform NH$_3$ into NH$_2$OH, which is then released into the periplasm, where it is oxidized to NO$_2^-$ (De Boer and Kowalchuk, 2001). However, the release of NH$_2$OH into the surrounding medium had also been reported (Schmidt et al., 2004b; Stüven et al., 1992), and non-detection in soils was explained by the highly reactive character (Moews and Audrieth, 1959). However, only recently a novel, highly sensitive method for the detection of NH$_2$OH in soils was introduced, with which the authors were able to detect NH$_2$OH in an acidic forest soil (Liu et al., 2014).

NO$_2^-$ (or in its protonated form HNO$_2$) and NH$_2$OH are N compounds potentially involved in abiotic N trace gas emissions from soils (see below). As they are microbially derived, the two nitrification intermediates are only available as reagents for abiotic N trace gas formation with soil microbial activity. This already suggests a tight coupling between biotic and abiotic N transformation processes, with abiotic N trace gas production being initiated by the microbiological generation of NO$_2^-$ and NH$_2$OH via nitrification and/or denitrification. Venterea (2007) argues that all known biochemical and chemical processes of soil N$_2$O production involve the reduction of NO$_2^-$, but studies have shown that the nitrification intermediate NH$_2$OH in conjunction with – but also in the absence of NO$_2^-$ – is the precursor of abiotic N$_2$O production, while reactions involving NO$_2^-$ in the absence of NH$_2$OH tend to produce mostly NO (Bremner et al., 1980; Butler and Gordon, 1986; Nelson, 1978; Spott and Stange, 2011). In a laboratory study, Venterea and Rolston (2000a) showed that gross NO production rates in agricultural soils were highly correlated with HNO$_2$ con-
A review of chemical nitrogen trace gas formation reactions in soils

centration and not with nitrification rates or N substrate availability, indicating that HNO$_2$ protonation might be more important than NO$_2^-$ concentration alone.

2.2. Accumulation of NO$_2^-$

Due to its highly reactive nature, the nitrification intermediate NH$_2$OH will usually not accumulate in soils (Moews and Audrieth, 1959), but under certain conditions NO$_2^-$ can accumulate in agroecosystems as well as in arid or seasonally arid unmanaged ecosystems (Gelfand and Yakir, 2008). NO$_2^-$ will only accumulate in the soil, when NO$_2^-$ consumption is lower than NO$_2^-$ production (Burns et al., 1996). In most non-agricultural soils, the oxidation of NO$_2^-$ to NO$_3^-$ proceeds faster than the conversion of NH$_3$ to NO$_2^-$, therefore NO$_2^-$ is normally not present in concentrations greater than 1 mg N kg$^{-1}$ soil (Chalk and Smith, 1983). Its accumulation in soils is depending on a set of soil characteristics such as soil pH, C and O$_2$ availability, and even more on agricultural practices, especially when alkaline conditions are promoted (Bezdicek et al., 1971; Burns et al., 1995; Chalk et al., 1975; Shen et al., 2003; Smith et al., 1997).

2.2.1 Role of soil properties

Under alkaline conditions both AOB and NOB are suppressed, but NOB are usually more sensitive to high pH and salt concentrations than AOB (Shen et al., 2003). The optimum soil pH for nitrification is between 7 and 9, but since the activity of NOB is more strongly inhibited at high soil pH than that of AOB, NO$_2^-$ may accumulate (Chalk et al., 1975). With the discovery of AOA, this classic neutral pH optimum for soil NH$_3$ oxidation was challenged, as different archaeal as well as bacterial subgroups have been found to fill diverse ecological niches broadening the pH spectrum of nitrification (Nicol et al., 2008).

However, NO$_2^-$ accumulation in soil is not only the resultant of NH$_3$ oxidation (NO$_2^-$ formation) and NO$_2^-$ oxidation (NO$_2^-$ consumption) during nitrification, but also of NO$_3^-$ reduction (NO$_2^-$ formation) and NO$_2^-$ reduction (NO$_2^-$ consumption) during denitrification. Burns et al. (1996) showed in an experiment with $^{15}$N-enriched N pools that both nitrification and denitrification produce NO$_2^-$ simultaneously in the same soil, though nitrification was the more important process for NO$_2^-$ accumulation in that experiment.

Smith et al. (1997) developed a model explaining NO$_2^-$ accumulation in soil. The model describes NO$_2^-$ accumulation as a result of the inhibitory effect of free NH$_3$ on NOB. The simulation showed that NH$_4^+$ oxidation was only required to be slightly in excess of NO$_2^-$ oxidation to cause NO$_2^-$ accumulation. Smith et al. (1997) concluded that the main process causing NO$_2^-$ accumulation is nitrification, while denitrification is only of minor importance. Shen et al. (2003) showed the effect
of pH on NO$_2^-$ accumulation in an incubation experiment. They were able to maintain high NO$_2^-$ concentrations in slurries of pH > 8, although NH$_4^+$ was not detected after several days of incubation any longer. This suggests a low NO$_2^-$ oxidizer activity plus relatively high NO$_2^-$ stability at high pH levels (Shen et al., 2003). Besides high soil pH, low temperatures, low C availability, low soil moisture content, and high levels of NH$_4^+$ can also be a cause of NO$_2^-$ accumulation (Burns et al., 1996; Christianson and Cho, 1983; van Cleemput and Baert, 1983).

The accumulation of NO$_2^-$ during denitrification is even more complex. The reduction of NO$_3^-$ to NO$_2^-$ is the first step in the multi-stage denitrification process. Again, the initial step has to be faster than the subsequent steps to cause NO$_2^-$ to accumulate. Since denitrification is an anaerobic process, O$_2$ is the factor most likely determining the equilibrium between NO$_2^-$ formation and consumption besides NO$_3^-$ availability (Burns et al., 1996). NO$_2^-$ reductase is the denitrification enzyme most sensitive to O$_2$, but in some bacteria it can also be inhibited at very high NO$_3^-$ concentrations (Cole, 1994). Another factor for NO$_2^-$ accumulation during denitrification can be C availability. If available C in soil is low, the competition for electrons is increased, with NO$_3^-$ and NO being stronger electron acceptors than NO$_2^-$, leading to NO$_2^-$ accumulation (Burns et al., 1996).

### 2.2.2 Agricultural practices

Agricultural practices are more responsible for NO$_2^-$ accumulation than natural conditions. High NO$_2^-$ concentrations may be found in soil after application of N fertilizers that tend to form alkaline solutions upon hydrolysis, such as urea and anhydrous NH$_3$ (Chalk et al., 1975; Chapman and Liebig, 1952). The initially high NH$_4^+$ concentrations in conjunction with an increased soil pH may have an inhibitory effect on NO$_2^-$ oxidizers (Burns et al., 1996). Alkaline conditions promote dissociation of NH$_4^+$ to NH$_3$ (pK$_a$ = 9.3), and so the accumulation of NO$_2^-$ is often linked to the toxicity of free NH$_3$ for NOB (Venterea and Rolston, 2000b). The sensitivity of NOB to free NH$_3$ is about two orders of magnitude as much as than for AOB (Smith et al., 1997). Yet the increase in soil pH is only temporarily. With ongoing nitrification, pH decreases again because the oxidation of NH$_4^+$ to NO$_2^-$ by AOB is associated with the release of protons, lowering the pH back to its original value or even below it (Chalk and Smith, 1983). In experiments with urea-treated soil, Burns et al. (1996) showed how NH$_4^+$ accumulated due to urea hydrolysis, accompanied by an increase in soil pH several days after application. This caused an accumulation of NO$_2^-$ due to the inhibition of NO$_2^-$ oxidation by free NH$_3$. When soil pH and, as a consequence, free NH$_3$ concentrations decreased, nitrifiers became active again consuming NO$_2^-$ (Burns et al., 1996).

The granule size and mode of application of alkaline hydrolyzing fertilizer also influence NO$_2^-$ accumulation (Hauck and Stephenson, 1965). Soil chemistry in the proximity around a large water-soluble fertilizer granule will be affected more by the fertilizer than the same amount of N distrib-
uted more widely and uniformly in form of smaller granules, or fertilizer applied as solution. This indicates that rate and method of fertilizer application can reduce NO$_2^-$ accumulation.

Urine patches are another relevant site for NO$_2^-$ accumulation in soils. At these sites, spots of elevated pH and high NH$_3$ concentrations via urea hydrolysis can occur (Monaghan and Barraclough, 1992). By this, NO$_2^-$ accumulation is a common phenomenon in grassland soils receiving high N inputs not only in the form of artificial fertilizer, but also as animal excrements (Burns et al., 1995).

An accumulation of more than 100 mg NO$_2^-$-N kg$^{-1}$ has been found after urea application (van Cleemput and Samater, 1996). Bezdick et al. (1971) found NO$_2^-$ accumulation of up to 140–265 mg NO$_2^-$-N kg$^{-1}$ after the addition of granular NH$_4^+$ sulfate and urea prills to an alkaline soil. However, these are extreme values, and in most field studies NO$_2^-$ concentrations were generally much lower (Burns et al., 1995; Davidson et al., 1991; Gelfand and Yakir, 2008; Venterea et al., 2003).

In a recent study, the contribution of urea and anhydrous NH$_3$ to elevated N$_2$O emissions was tested in a field-scale experiment over different growing seasons (Venterea et al., 2010). It was found that anhydrous NH$_3$ led to higher NO$_2^-$ levels than urea and, therefore, to higher gaseous losses of N. NO$_2^-$ accumulation following urea application can also occur, especially if applied as bands or as liquid urea-NH$_4$NO$_3$ (Mulvaney et al., 1997). Recently observed N$_2$O emissions from corn fertilized with anhydrous NH$_3$ were even twice as high as with urea (Venterea et al., 2010).

### 2.2.3 Drying/rewetting in seasonally dry ecosystems

In seasonally dry ecosystems the biological activity is limited by the temporally low amounts of water. This results in an inactivity of the N-cycling and other microbial-driven soil processes during drought. When water becomes available again, these ecosystems usually show high rates of biological activity and high N trace gas emission rates (Davidson et al., 1991). It was observed in an incubation experiment that NH$_4^+$ oxidation was activated immediately after rewetting of a dry soil, and NO$_2^-$ accumulated at the same rate (Gelfand and Yakir, 2008). This NO$_2^-$ accumulation was interpreted as a time delay between the transformations of NH$_4^+$ and NO$_2^-$ as a consequence of different drought stress tolerance levels and recovery rates of AOB and NOB (Gelfand and Yakir, 2008). These findings are in accordance with the results of Tappe et al. (1999) who found that AOB were able to use substrate directly after starvation, while NOB needed a three-fold longer period before it was able to use NO$_2^-$. Another factor favoring NO$_2^-$ accumulation under field conditions in dry environments are high salt concentrations in soil, which can inhibit NO$_2^-$ oxidation to NO$_3^-$ (Nejidat, 2005). These observations suggest a seasonal pattern of NO$_2^-$ accumulation in ecosystems with highly periodic precipitation, with accumulation of NO$_2^-$ being highest at the beginning of the rainy season, leading to pulses of N trace gas emissions at the onset of the first rains.
2.3. Abiotic N trace gas production mechanisms

As mentioned above, there are purely chemical reactions known to produce N trace gases. These reactions involve the nitrification intermediates (NO$_2^-$ and NH$_2$OH) as N sources. These reactions can all occur simultaneously, and are all competing for N substrate between each other, and also with biological N transformation processes. The following sections give an overview over the main abiotic production mechanisms, their controls and kinetics.

2.3.1 Self-decomposition of NO$_2^-$

The chemical stability of NO$_2^-$ in soil solution is associated with the equilibrium between HNO$_2$ and NO$_2^-$ (van Cleemput and Samater, 1996). Therefore, it is highly dependent on pH. Due to the low pK$_a$ (3.3 at 25 °C) for the equilibrium reaction (Pires et al., 1994), notable concentrations of HNO$_2$ do not occur in most soils. NO$_2^-$ is stable above pH 5.5, so that HNO$_2$ should only form in acidic soils (van Cleemput and Baert, 1978). However, soil pH is not uniform and can vary largely at the microscale. The surface of a clay mineral is about 100 times more acidic than the surrounding soil solution (Harter and Ahlrichs, 1967), and the same applies to the vicinity of plant roots (Marschner et al., 1986), which could lead to shifts in the equilibrium.

The classic equation for HNO$_2$ decomposition is described as

\[ 3 \text{HNO}_2 \rightarrow \text{HNO}_3 + 2 \text{NO} + \text{H}_2\text{O} \]  

(Nelson and Bremner, 1970a) found that the following equation represents HNO$_2$ self-decomposition in soil solution better:

\[ 2 \text{HNO}_2 \rightarrow \text{NO} + \text{NO}_2 + \text{H}_2\text{O} \]  

HNO$_2$ decomposition produces NO$_3^-$ only in closed systems with available O$_2$, when there is no reagent in the system that sorbs NO and NO$_2$ (Nelson and Bremner, 1970a). In open systems, the produced NO may escape to the atmosphere, be absorbed by soil, or react under aerobic conditions with O$_2$ to form NO$_2$ (van Cleemput and Samater, 1996). Nelson and Bremner (1970a) found NO$_2$ as the main product of NO$_2^-$ self-decomposition; van Cleemput and Baert (1984), however, measured only up to 1.4% of added NO$_2^-$ being decomposed to NO$_2$, with NO being the main gaseous product. This suggests that most of the produced NO$_2$ dissolves in soil as formulated by Smith and Chalk (1980):

\[ 2 \text{NO}_2 + \text{H}_2\text{O} \rightarrow \text{NO}_2^- + \text{NO}_3^- + 2 \text{H}^+ \]
Thus, NO$_3^-$ is not the direct product of NO$_2^-$ decomposition, but a product of the disproportionation of NO$_2$, resulting from NO$_2^-$ decomposition, as given in Equation (6) (Kappelmeyer et al., 2003). The amount of NO$_3^-$ formed in this way is dependent on the rate of diffusion of NO$_2$ through the soil (Nelson, 1982).

In a recent study, Mørkved et al. (2007) found an immediate burst of NO production shortly after the addition of NO$_2^-$ to sterile slurries of acidic soils (pH 4.1). This instantaneous NO peak was proportional to the amount of added NO$_2^-$. It was suggested that the rapid NO production was due to chemical decomposition of NO$_2^-$. The same sudden release of NO after the addition of NO$_2^-$ to an acidic central African rain forest soil was found by Serça et al. (1994). The onset of the reaction only seconds after the addition of NO$_2^-$ indicates a purely chemical reaction, which could be corroborated by the modification of the pH (Serça et al., 1994). Yamulki et al. (1997) made the observation in a comparison of sterile and non-sterile grassland soil cores that an important fraction (29% at pH 3.9) of NO was produced via chemical decomposition of NO$_2^-$. These results are in agreement with a previous study by Nägele and Conrad (1990), who observed a 71% conversion rate of consumed NO$_2^-$ to NO in autoclaved soil slurries at pH 4.

It is accepted that the self-decomposition of NO$_2^-$ follows first-order or pseudo-first-order kinetics (van Cleemput and Baert, 1984), as a significant linear relationship between NO$_2^-$ concentration and incubation time in different treatments was found (Laudelout et al., 1977). The first-order rate constant was discovered to be influenced by NO$_2^-$ concentration, temperature, and pH (Laudelout et al., 1977). NO$_2^-$ decomposition is stimulated by decreasing pH and by addition of NO$_2^-$. Subsequently, the dilution of the soil solution leads to a decrease of the reaction. Venterea and Rolston (2000a) noticed that the emission of NO correlated rather with HNO$_2$ than with NO$_2^-$ alone, which also emphasizes the stimulating effect of NO$_2^-$ concentration and pH on NO$_2^-$ decomposition because both account for the formation of HNO$_2$ as mentioned above. As a purely chemical process, the decomposition of NO$_2^-$ has no temperature optimum and will increase with increasing temperature (Kesik et al., 2006). Venterea et al. (2005) found a negative correlation between abiotic NO emissions and soil water content, and emphasized the occurrence of abiotic NO production primarily at the interface of soil surface and soil solution. They explained the relation between NO emissions and soil water content by an increasing ratio of interfacial area between soil matrix and solution, and additionally the importance of surficial acidity of soil mineral and organic particles that in turn is also related to cation exchange capacity of a soil. The hypothesis of a surface-mediated reaction also implies the effect of soil clay and SOM content and surface charge density on abiotic NO formation (Venterea et al., 2005).

A recently found source of gaseous N loss from acidic soils that can be closely linked to nitrite self-decomposition is the release of nitrous acid in gaseous form (HONO) (Su et al., 2011). Besides the equilibrium between NO$_2^-$ and HNO$_2$ in the aqueous phase, aqueous HNO$_2$ is also in equilibri-
um with gaseous HONO, which is also controlled by acidity and the $\text{NO}_2^-$ concentration. When the HONO equilibrium gas phase concentration is higher than the actual HONO gas phase concentration, HONO is released from the aqueous phase (Su et al., 2011). Su et al. (2011) as well as Oswald et al. (2013) showed that soil $\text{NO}_2^-$ can act as a strong source of HONO. This makes $\text{NO}_2^-$ in acidic soils not only a potential source of elevated abiotic NO emissions, but also of high abiotic HONO emissions.

2.3.2 Reactions of $\text{NO}_2^-$ with metals

The reaction of $\text{NO}_2^-$ with metals in their reduced state, leading to the formation of gaseous N products, has long been recognized as another non-enzymatic pathway of N loss from soils. Wullstein and Gilmour (1964) introduced a mechanism, wherein $\text{NO}_2^-$ is reduced to NO by certain transition metals in sterile, moderately acidic soils by the following reaction (with $\text{Fe}^{2+}$ representing redox-active transition metals):

$$\text{Fe}^{2+} + \text{NO}_2^- + 2 \text{H}^+ \rightarrow \text{Fe}^{3+} + \text{NO} + \text{H}_2\text{O}$$ (7)

Transition metals promoting $\text{NO}_2^-$ losses in their reduced state were found to be iron, copper, and manganese (Wullstein and Gilmour, 1964). Nelson and Bremner (1970b) found that only cuprous ($\text{Cu}^+$), ferrous ($\text{Fe}^{2+}$), and stannous ($\text{Sn}^{2+}$) ions promote $\text{NO}_2^-$ decomposiiton, and showed on the basis of the standard electrode potentials that manganous ($\text{Mn}^{2+}$) ions cannot reduce $\text{NO}_2^-$ in acidic media. However, the authors stated that there is no evidence that the concentrations of $\text{Fe}^{2+}$, $\text{Cu}^+$, and $\text{Sn}^{2+}$ in soils under conditions promoting chemodenitrification are high enough to play a significant role in $\text{NO}_2^-$ decomposition. Studies in buffered solutions indicate that $\text{Fe}^{2+}$ and $\text{Cu}^+$ concentration must at least be 56 and 318 mg l$^{-1}$, respectively, before the mechanism according to Equation (7) can proceed (Jones et al., 2000).

A reaction between $\text{NO}_3^-$ and dissolved $\text{Fe}^{2+}$ has also been shown in the literature (Postma, 1990):

$$10 \text{Fe}^{2+} + 2 \text{NO}_3^- + 14 \text{H}_2\text{O} \rightarrow 10 \text{FeOOH} + \text{N}_2 + 18 \text{H}^+$$ (8)

However, this reaction appears to be very slow at lower temperatures, and is, therefore, not of high relevance in natural systems (Postma, 1990). Ottley et al. (1997) suggested that such an abiotic reduction of $\text{NO}_3^-$ may only be feasible in groundwater with solid phase copper as a catalyst on a timescale of months to years.
Controlling factors

Controlling factors of Fe$^{2+}$-mediated NO$_2^-$ decomposition are commonly reported to be pH and the concentrations of NO$_2^-$, NO$_3^-$, and Fe$^{2+}$ (with NO$_3^-$ having a much lower reactivity than NO$_2^-$) (Moraghan and Buresh, 1977; Nelson and Bremner, 1970b; Sørensen and Thorling, 1991; van Cleemput and Baert, 1983). Equation (7) requires that hydrogen ions are present in soil solution, so that at least moderate acidity is a prerequisite for this reaction. However, a considerable variability on the influence of pH on NO$_2^-$ decomposition by Fe$^{2+}$ is reported in the literature. While Chao and Kroontje (1966) found a decreasing reduction of NO$_2^-$ by Fe$^{2+}$ as pH increases from 5 to 6, which is in conjunction with the results of van Cleemput and Baert (1983), Moraghan and Buresh (1977), in contrast, observed a higher N trace gas production when raising the pH in a Fe$^{2+}$-containing medium from 6 to 8. At pH 6, however, a catalytic effect of Cu$^{2+}$ ions on NO$_2^-$ decomposition by Fe$^{2+}$ was found that appears to have little environmental significance yet (Sørensen and Thorling, 1991). Jones et al. (2000) argued that the reduction of NO$_2^-$ by reduced metal ions is the only abiotic NO$_2^-$ decomposition reaction in soils that does not require HNO$_2$ and is therefore possible at higher pH. The influence of pH is very complex, as it changes the equilibrium between NO$_2^-$ and free HNO$_2$ and the speciation of iron through the formation of different Fe$^{3+}$-precipitates and oxihydroxide species (Kampschreur et al., 2011). From their experiments, van Cleemput and Baert (1983) concluded that all conditions leading to high amounts of Fe$^{2+}$ in solution, i.e., a low redox potential and low pH, but also the physical structure of the iron compound, enhance NO$_2^-$ decomposition. Less crystalline iron oxides, and even more so amorphous iron, are more soluble than crystalline iron and stimulate NO$_2^-$ decomposition by bringing more Fe$^{2+}$ into solution (van Cleemput and Baert, 1984).

Kinetics and mechanisms

The kinetics of the Fe$^{2+}$-enhanced NO$_2^-$ decomposition were found to be second order (van Cleemput and Baert, 1983). The rate constant increased with increasing Fe$^{2+}$-concentration, but decreased with increasing pH. The dependence of the reaction on temperature increased at lower pH levels or at increasing Fe$^{2+}$-concentrations (van Cleemput and Baert, 1983). The half-life of NO$_2^-$ is obviously reduced by lower pH values and higher Fe$^{2+}$ concentrations, yet, the influence of Fe$^{2+}$ decreases at very high concentrations. Kampschreur et al. (2011) found the kinetics to be more affected by NO$_2^-$ than by iron.

While conditions for NO$_2^-$ and Fe$^{2+}$ accumulation do not occur coincidently in one soil layer, the possibility for the reaction exists at the interface between an aerobic zone overlaying an anaerobic zone where NO$_2^-$ moving downwards meets Fe$^{2+}$ moving upwards (Sørensen and Thorling, 1991; van Cleemput and Baert, 1983). Still, concentrations of Fe$^{2+}$ ions in most soils are considered too
low to promote NO$_2^-$ decomposition (Nelson and Bremner, 1970b; van Cleemput and Samater, 1996). The impact of metal cations on NO$_2^-$ decomposition is controversial, while some authors rule out the possibility that soil minerals and metal cations play a significant role completely (Bremner, 1997), others argue that iron, especially in highly soluble compounds, has a stimulating effect, and therefore intensifies NO$_2^-$ decomposition even in slightly acidic soils (van Cleemput, 1998).

Recently, Kampschreur et al. (2011) argued that the iron turnover rate is highly dependent on NO$_2^-$ concentration, and that it is fast enough to be responsible for significant N trace gas emissions, especially when NO$_2^-$ accumulates. This mechanism suggests a coupling between the iron and N cycles. While Fe$^{3+}$ is reduced to Fe$^{2+}$ biologically, Fe$^{2+}$ subsequently reduces NO$_2^-$ to NO by the simultaneous oxidation of Fe$^{2+}$ in a purely chemical reaction (Brons et al., 1991). Some of the NO can then be loosely bound to excess Fe$^{2+}$, so the Fe$^{2+}$–NO complex may not work as a permanent sink for NO (Brons et al., 1991). Moreover, NO can react further with Fe$^{2+}$ in a similar reaction as NO$_2^-$ to form N$_2$O:

$$2 \text{Fe}^{2+} + 2 \text{NO} + 2 \text{H}^+ \rightarrow 2 \text{Fe}^{3+} + \text{N}_2\text{O} + \text{H}_2\text{O} \quad (9)$$

As the purely chemical reactions proceed very fast, the system will be limited by the availability of Fe$^{2+}$ (Sørensen and Thorling, 1991). Kampschreur et al. (2011) endorsed the idea of a two-step mechanism for the reduction of NO$_2^-$ by Fe$^{2+}$ according to Equations 7 and 9, leading to N$_2$O emission, with NO as an intermediate. In their study, the emission of NO occurred immediately after the addition of NO$_2^-$ to Fe$^{2+}$, while N$_2$O emission was only observed after a significant amount of Fe$^{2+}$ had been oxidized by NO$_2^-$, leading to the formation of Fe$^{3+}$ precipitates. The reduction rate of NO was found to be strongly dependent on the size of the reactive surface and the Fe$^{2+}$ sorption to it (Kampschreur et al., 2011). Thus, compared to NO formation kinetics, which appear to be first order, the formation of N$_2$O via metal cations seems to be more complex due to the Fe$^{3+}$ mineral formation kinetics (Kampschreur et al., 2011).

Another reaction mechanism for the reduction of NO$_2^-$ by Fe$^{2+}$ directly to N$_2$O was proposed by Sørensen and Thorling (1991), who used a lepidocrocite suspension for their laboratory study:

$$4 \text{Fe}^{2+} + 2 \text{NO}_2^- + 5 \text{H}_2\text{O} \rightarrow 4 \text{FeOOH} + \text{N}_2\text{O} + 6 \text{H}^+ \quad (10)$$

A different formulation of this reaction, that takes into account that magnetite formation is favored under the experimental conditions used by Sørensen and Thorling (1991), is:

$$6 \text{Fe}^{2+} + 2 \text{NO}_2^- + 5 \text{H}_2\text{O} \rightarrow 2 \text{Fe}_3\text{O}_4 + \text{N}_2\text{O} + 10 \text{H}^+ \quad (11)$$
In their findings the presence of lepidocrocite was required for the initiation of the reaction between Fe$^{2+}$ and NO$_2^-$, while the reaction was very slow in the absence of lepidocrocite. It is assumed that rather a complexed, more reactive form of Fe$^{2+}$ is involved than dissolved Fe$^{2+}$. Therefore, two simultaneous processes are important: the initial binding of Fe$^{2+}$ to build a simple Fe$^{2+}$-containing complex with a Fe$^{3+}$ oxyhydroxide, and the subsequent reaction between NO$_2^-$ and the Fe$^{2+}$-containing compound (Sørensen and Thorling, 1991).

An increased N$_2$O production due to the reduction of NO$_2^-$ and the simultaneous oxidation of Fe$^{2+}$ to Fe$^{3+}$ was also observed in the presence of goethite (Cooper et al., 2003). These results are in accordance with Coby and Picardal (2005), who also found this reaction to be surface-catalyzed and failed to detect any significant N$_2$O production in systems lacking a surface catalyst. It is likely that surface reactions are not restricted to pure iron oxide, but can also be catalyzed by iron-containing sediments and soil particles. Coby and Picardal (2005) discussed the possibility of such reactions on microbial cell surfaces which have adsorbed Fe$^{2+}$. A recent study confirmed the assumption of a surface-catalyzed reaction (Tai and Dempsey, 2009). The authors used hydrous ferric oxide as their Fe$^{3+}$ source and did not observe any reduction of NO$_2^-$ in the absence of hydrous ferric oxide. Another interesting finding is that solid-bound Fe$^{2+}$ does not react with NO$_2^-$ in the absence of dissolved Fe$^{2+}$, so that the reaction stopped once dissolved Fe$^{2+}$ was depleted (Tai and Dempsey, 2009).

Another, yet similar mechanism was proposed by Samarkin et al. (2010) only recently, who found N$_2$O fluxes similar to those from fertilized tropical soils from a hypersaline playa lake without any biological activity in Antarctica. They observed N$_2$O emissions higher than expected on the lakeshore and tested the hypothesized chemical reaction in the laboratory. The suggested mechanism is a reaction between NO$_2^-$-rich brine and Fe$^{2+}$-containing minerals derived from the surrounding igneous Ferrar Dolerite. The proposed reaction equation is the same as in Equation (11). Samarkin et al. (2010) found that the production of N$_2$O is 100 times higher with NO$_2^-$ than with NO$_3^-$, thus the reduction of NO$_3^-$ to NO$_2^-$ will be the rate-limiting step. The reaction was observed at –20 °C with an increasing N$_2$O production rate with increasing temperature. The Arrhenius activation energy ($E_A$) of this reaction was 18.4 kJ mol$^{-1}$, indicating a diffusion-controlled reaction (Samarkin et al., 2010).

2.3.3. Reactions of NO$_2^-$ with SOM

NO$_2^-$ decomposition

Several studies published in the past noted a connection between the rate of NO$_2^-$ decomposition and SOM content (e.g., Blackmer and Cerrato, 1986; Bremner and Führ, 1966; Kappelmeyer et al., 2003; Nelson and Bremner, 1970a; Thorn and Mikita, 2000). Most of these studies showed an en-
enhanced decomposition of NO$_2^-$ in the presence of SOM in soils. Nelson and Bremner (1969) found in a study with several soils that the removal of SOM by combustion markedly reduced the soils’ ability for NO$_2^-$ decomposition. In the same study, they noticed that the soils with the highest C content were characterized by the largest recovery of added N due to fixation to SOM, and that SOM promoted decomposition of NO$_2^-$ at a pH above 5, where self-decomposition of NO$_2^-$ is very limited.

NO$_2^-$ fixation

The ability of soils to fix NO$_2^-$ by a purely chemical reaction with SOM was first demonstrated by Bremner and Führ (1966). They found that this fixation did not only occur in C-rich acidic, but also in mildly alkaline soils, and pointed out that the mechanism behind the fixation reaction likely involves aromatic structures of organic material. Lignin and the lignin-like fraction of SOM have been found to have the largest capacity for NO$_2^-$ fixation of all tested organic substances (Bremner and Führ, 1966). It is known that phenolic materials react with HNO$_2$ to form nitroso compounds, which is likely the mechanism behind the reaction between NO$_2^-$ and lignin, a phenolic substance of high molecular weight.

Führ and Bremner (1964a) found a fixation of 11% and 16% of added NO$_2^-$ in two mineral soils with a gaseous N loss of 79% and 77%, respectively. In an acidic peat soil, a NO$_2^-$ fixation of even 28% of the added NO$_2^-$ and a reduced gaseous N loss of 63% were reported in the same study. In another experiment, Führ and Bremner (1964b) found that even at neutral to slightly alkaline conditions up to 25% of added NO$_2^-$-N were fixed by SOM, although the reaction is pH-dependent and fixation is promoted by acidity.

Besides pH, soil C content is another important controlling factor, as a high correlation between SOM and the amount of N fixed was observed (Führ and Bremner, 1964b). The fixation mechanism was shown even at low NO$_2^-$ concentrations, at which relatively more NO$_2^-$ was fixed (Führ and Bremner, 1964b). The authors found the chemical fixation of NO$_2^-$ to be faster than the self-decomposition of NO$_2^-$, as it occurred at very low pH where NO$_2^-$ self-decomposition is very rapid.

Stevenson et al. (1970) assumed phenolic groups being also involved in the formation of NO, so that the reaction of NO$_2^-$ with SOM not only enhances NO$_2^-$ fixation, but may also lead to the abiotic production of NO. Bremner and Führ (1966) concluded that, although SOM is involved in the gaseous loss of NO$_2^-$, high SOM contents tend to reduce the gaseous loss of N due to chemical fixation and by the promotion of microbial nitrification.
Nitrosation reaction mechanism

Stevenson et al. (1970) presented the idea of nitrosation reactions, i.e., the formation of a nitroso group due to the reaction of $\text{NO}_2^-$ with an organic molecule, as the mechanism behind $\text{NO}_2^-$ fixation by SOM, but stated that the formed nitroso compounds are labile and may undergo further reactions to form gaseous N compounds. They obtained NO as the main gaseous product beside $\text{N}_2$, $\text{N}_2\text{O}$, and $\text{CO}_2$ as minor components in the reaction of $\text{NO}_2^-$ with lignin and humic preparations even at neutral pH, where self-decomposition of $\text{NO}_2^-$ is negligible.

Blackmer and Cerrato (1986) suggested that the highest abiotic production of NO can be associated with soils having high SOM content and low pH. The formation of nitrosophenols and oximes from phenolic entities and activated methylene groups upon nitrosation of fulvic and humic acids was demonstrated by Thorn and Mikita (2000). Like Stevenson et al. (1970) before, Thorn and Mikita (2000) also observed formation of $\text{N}_2\text{O}$ upon reaction between $\text{NO}_2^-$ and SOM at neutral to mildly alkaline pH values. They discussed the reaction of a second $\text{HNO}_2$ molecule with an oxime group, formed during the reaction of $\text{HNO}_2$ with a phenol or an activated methylene group, as a possible mechanism, in which the formation of an N–N-bond constitutes the first step. The authors observed $\text{CO}_2$ as another gaseous product upon nitrosative decarboxylation of certain aromatic acids.

Boudot and Chone (1985) presented a scheme of the internal N cycle in humiferous acidic soils implying importance to the effect of SOM on abiotic fixation and decomposition of $\text{NO}_2^-$. In an acid colluvial soil with a high humic acid content they found a $\text{NO}_2^-$ fixation of 68%, with only 20% being available for further nitrification. In these acidic soils with high organic C contents, Boudot and Chone (1985) found nitrification of $\text{NO}_2^-$ to $\text{NO}_3^-$ even completely suppressed by enhanced self-decomposition of $\text{HNO}_2$ and fixation of $\text{NO}_2^-$ to SOM.

In a field study on the effects of long-term N inputs in two temperate forest soils, Magill et al. (2000) found a near complete retention of added ammonium nitrate and discussed three possible mechanisms for the incorporation of N into SOM: N immobilization by microbes, mycorrhizal assimilation and exudation of organically bound N, and chemical reactions with SOM. Their results suggest a greater importance for abiotic N transformation pathways than previously thought, with nitrosation reactions playing a prominent role.

In a recent laboratory study (Islam et al., 2008), a fast $^{15}\text{N}$ incorporation of about 20% into the SOM pool was observed after the addition of $^{15}\text{N}$-labelled $\text{NO}_2^-$ to two soils with a pH of about 5 and an organic C content above 6%, while in an alkaline soil (pH 8.1) no $^{15}\text{N}$ was recovered from SOM at all, pinpointing the importance of soil pH. The fast nature of these chemical reactions could also explain low nitrification in some soils. In contrast, Kappelmeyer et al. (2003) found indeed enhanced abiotic N trace gas formation by artificial humic matter, but failed to find N incorporation. Venterea and Rolston (2000a) observed a strong positive correlation between the pro-
duction rates of NO ($r^2 = 0.9$) and N$_2$O ($r^2 = 0.82$) in sterile agricultural soils with organic C contents from 0.3–1.4%. This data is consistent with further experiments in which a non-linear relation between abiotic NO production and NO$_2^-$ concentration was observed that may be related to reactions of NO$_2^-$ with SOM (Venterea and Rolston, 2000b).

**Abiotic NO$_3^-$ fixation**

A study by Dail et al. (2001) with live and sterilized soils suggested that NO$_3^-$ added to an acidic forest soil undergoes a fast abiotic immobilization via incorporation into insoluble SOM. This was in obvious contrast to the results of Colman et al. (2007), who could not find any evidence of abiotic NO$_3^-$ incorporation in samples from 45 North and South American soils. They argued that reported abiotic incorporation of NO$_3^-$ was due to analytical artifact. As an explanation, the possibility of a partial reduction of NO$_3^-$ to NO$_2^-$ by biological respiration and/or abiotic processes followed by an incorporation of NO$_2^-$ into SOM was discussed (Dail et al., 2001; Fitzhugh et al., 2003).

Based on these findings, Davidson et al. (2003) proposed the “ferrous wheel hypothesis” for the abiotic NO$_3^-$ immobilization in soils. In this conceptual model, Fe$^{3+}$ is reduced by SOM to Fe$^{2+}$, the produced reactive Fe$^{2+}$ species in turn reduce NO$_3^-$ to NO$_2^-$, and NO$_2^-$ subsequently reacts with phenolic organic matter to form dissolved organic N compounds (Davidson et al., 2003). However, there is some controversy about this hypothesis. Colman et al. (2008) questioned the second step of the “ferrous wheel hypothesis”, i.e., the reaction between Fe$^{2+}$ and NO$_3^-$: This reaction is very slow in absence of a catalyst, and the suggested catalysts (copper, green rust or surface bound Fe$^{2+}$) would rather produce NH$_3$ or N$_2$ (see Equation (8)) than NO$_2^-$ (Hansen et al., 1994; Ottley et al., 1997; Postma, 1990).

As illustrated above, SOM plays a complex role in NO$_2^-$-driven abiotic processes in soils, on the one hand being able to fix N in soil and on the other hand enhancing gaseous N emissions. There is still controversy about its relevance for N trace gas emissions, and not all mechanisms are adequately understood. Beyond NO$_2^-$, several other potential pathways for abiotic nitrosation reactions with low molecular weight N compounds, such as NH$_3$, NH$_2$OH, and amino acids, add to the complexity (Thorn and Mikita, 2000). The role of SOM suggests that agricultural management practices, which are aligned to increase soil C storage, may have the unintentional side-effect of enhanced N$_2$O emission that could counteract greenhouse gas benefits of C sequestration (Venterea, 2007).

### 2.3.4. Reactions involving hydroxylamine

NH$_2$OH is the first intermediate in the enzymatic transformation of NH$_3$ to NO$_2^-$ by autotrophic nitrifying bacteria and archaea. Although there has long been no evidence for the extracellular re-
lease of NH$_2$OH and no reports of detection in soil, there is a possibility of fast chemical decompo-
sition of NH$_2$OH leading to the formation of N$_2$O and N$_2$ (Bremner et al., 1980). Nelson (1978)
found that NH$_2$OH is quantitatively decomposed by HNO$_2$ at a soil pH below 5:

\[
\text{NH}_2\text{OH} + \text{HNO}_2 \rightarrow \text{N}_2\text{O} + 2 \text{H}_2\text{O}
\]  

(12)

This is long known in chemistry and was first described by Meyer (1875). In aqueous solution,
NH$_2$OH and HNO$_2$ react readily at room temperature (Hughes and Stedman, 1963). The reaction is
pH-dependent and increases rapidly below pH 4, as HNO$_2$ rather than NO$_2^-$ is the reactive species
(Stevens and Laughlin, 1994). Döring and Gehlen (1961) found that the mechanism of HNO$_2$ on
NH$_2$OH involves a primary nitrosation of free NH$_2$OH. N$_2$O$_3$ likely acts as the nitrosating agent
(Hughes and Stedman, 1963). The nitrosation reaction shows a high kinetic complexity with differ-
ent intermediates. Döring and Gehlen (1961) argued that especially in buffered solution the nucle-
ophilic buffer anion can work as a carrier for nitrosating species and exert great influence on the
reaction kinetics. Bothner-By and Friedman (1952) found that at neutral pH a symmetrical inter-
mediate is involved, presumably hyponitrous acid (HO–N=N–OH) or rather its anion hyponitrite
(O–N=N–O–), while at lower pH values, the initial nitrosation leads to the formation of an unsym-
metrical intermediate, presumably N-nitrosohydroxylamine. The exact mechanism is still not com-
pletely understood, mainly because of the fast sequential isomerization stages and proton transfer
from NH$_2$OH to the end product N$_2$O (Fehling and Friedrichs, 2011). Nitroxyll (HNO), the mono-
mer of hyponitrous acid, has long been recognized as an important direct precursor of chemical
N$_2$O formation (Bonner and Hughes, 1988). A recent modeling study proved that N$_2$O production
was initialized by the formation of HNO, followed by a fast dimerization leading to hyponitrous
acid (Fehling and Friedrichs, 2011).

In soils, the simplified reaction mechanism of N$_2$O formation from NH$_2$OH is composed of two
steps: first the enzymatic production of NH$_2$OH, and second the chemical decomposition of
NH$_2$OH (Stüven et al., 1992). N$_2$O has been reported to be the major product of this reaction, and
only small amounts of N$_2$ were produced, except for calcareous soils, in which N$_2$ formation ex-
cceeded that of N$_2$O (Bremner et al., 1980). Bremner et al. (1980) observed that the production of
N$_2$O by NH$_2$OH decomposition significantly exceeded the production of N$_2$O via NO$_2^-$ self-
decomposition from different sterilized soils in a laboratory incubation experiment, in which NO
was the major product.

From laboratory experiments, Minami and Fukushi (1986) presumed that the interaction between
NO$_2^-$ and NH$_2$OH is largely of chemical origin. However, Bremner et al. (1980) and Minami and
Fukushi (1986) found that the addition of NO$_2^-$ to sterilized soils treated with NH$_2$OH did not
prominently increase the production of N$_2$O. Furthermore, the formation of N$_2$O was found to be
faster than the reaction between NO$_2^-$ and NH$_2$OH in the absence of soil (Bremner, 1997). This suggests that another reaction must also be responsible for the decomposition of NH$_2$OH in soil. Bremner et al. (1980) detected a correlation between the formation of N$_2$O through the chemical decomposition of NH$_2$OH and oxidized forms of iron and Mn. A scheme for the reaction of NH$_2$OH and an iron-containing compound was presented by Butler and Gordon (1986):

$$4 \text{Fe}^{3+} + 2 \text{NH}_2\text{OH} \rightarrow 4 \text{Fe}^{2+} + \text{N}_2\text{O} + \text{H}_2\text{O} + 4 \text{H}^+ \quad (13)$$

However, although Fe$^{3+}$ is generally much more abundant in soils than Mn$^{4+}$, NH$_2$OH will preferably react with manganese because of the higher redox potential of the Mn$^{4+}$/Mn$^{2+}$ redox pair compared to the Fe$^{3+}$/Fe$^{2+}$ redox pair. This fact also allows the selective extraction of manganese with NH$_2$OH in soil chemical analyses, while leaving the major part of the iron unaffected (Chao, 1972). It has also been shown that small amounts of buffer substances can significantly reduce the rate of the reaction represented by Equation (13) (Heil et al., 2014). In soils iron might be complexed too tightly to be available as a reaction partner for NH$_2$OH. Thus, it appears that a reaction with manganese, as postulated by Nelson (1978), could be more important despite the generally lower manganese content in soils compared to iron:

$$2 \text{MnO}_2 + 2 \text{NH}_2\text{OH} \rightarrow 2 \text{MnO} + \text{N}_2\text{O} + 3 \text{H}_2\text{O} \quad (14)$$

These two equations (13) and (14) are a simplistic representation of the chemical mechanisms, as there are more intermediate steps involved, and again HNO was proposed as the key intermediate (Butler and Gordon, 1986). Spott and Stange (2011) recently proved in a laboratory experiment in a slightly alkaline soil suspension that NH$_2$OH was rapidly transformed into N$_2$O within a few hours after NH$_2$OH application, independently from the presence of NO$_2^-$ or NO$_3^-$. These results are in accordance with Bremner et al. (1980) who reported a major conversion of NH$_2$OH to N$_2$O within two hours by a redox reaction with oxidized metal species.

Besides oxidation by soil constituents, NH$_2$OH can also be oxidized by O$_2$, or undergo disproportionation (Bonner et al., 1978; Moews and Audrieth, 1959). However, both processes are considerably slower than the processes mentioned above. The oxidation of NH$_2$OH by O$_2$ is faster than the disproportionation, which was not observed at pH 3, and was only slightly higher at elevated pH (Bonner et al., 1978). However, both processes are significantly catalyzed by transition metals, especially copper, so that they cannot be completely excluded, and should be mentioned in this context.

In an experiment it was found that after NH$_2$OH addition to soil not all of the NH$_2$OH–N could be accounted for as gaseous N loss or as inorganic forms of N (Nelson, 1978). The amount of non-
recovered NH$_2$OH–N exhibited a strong positive correlation with SOM content (Bremner et al., 1980; Nelson, 1978). On average, 25% of added NH$_2$OH–N was apparently organically bound and not easily extractable after fixation. It is known that NH$_2$OH reacts with several SOM sources, especially carbonyl groups, to produce oximes (Porter, 1969), so the observed NH$_2$OH fixation likely occurs via the formation of oximes in a reaction of NH$_2$OH with carbonyl groups in SOM and humic acids (Nelson, 1978):

$$R_1R_2–C=O + NH_2OH \rightarrow R_1R_2–C=N–OH + H_2O$$

This reaction was also made use of to measure carbonyl groups in SOM (Porter, 1969).

A mechanism by which NH$_2$OH is released from microorganisms into the medium was proposed by Stüven et al. (1992): when electrons are generated by the oxidation of pyruvate or formate, an imbalance between NH$_3$ and NH$_2$OH oxidation is induced, and NH$_2$OH is released. It was shown that during nitrification by mutant strains of *Nitrosomonas europaea* high amounts of NH$_2$OH were released into the medium and N$_2$O was subsequently emitted most likely via chemical decomposition of NH$_2$OH (Schmidt et al., 2004b). Although a release of NH$_2$OH may not be favorable for AOB, as NH$_2$OH oxidation to NO$_2^–$ is an energy-generating step for them (Arp and Stein, 2003), abiotic reactions of NH$_2$OH may still take place, owing to the high reactivity of NH$_2$OH. The usual lack of detection of NH$_2$OH in soils may be due to the high reactivity of NH$_2$OH in four competing processes: (i) the chemical formation of N$_2$O, (ii) the abiotic incorporation of NH$_2$OH in SOM, (iii) the oxidation of NH$_2$OH to NO$_2^–$ during nitrification, and (iv) biotic N$_2$O formation (Spott and Stange, 2011).

It cannot be ignored that N$_2$O is generated from the decomposition of NH$_2$OH, still most authors are in agreement that it is not a main mechanism for the production of N$_2$O in soils under field conditions (Bremner, 1997; Bremner et al., 1980; Conrad, 1996; Minami and Fukushi, 1986). Nevertheless, if the non-detection of NH$_2$OH in soils is only due to the fast chemical decomposition, emissions from chemical reactions of NH$_2$OH can probably be much higher than widely expected. Spott and Stange (2011) presumed that due to the rapid chemical NH$_2$OH conversion, N$_2$O produced during nitrification may be abiotically produced from NH$_2$OH rather than derived microbially. Depending on soil conditions, such as pH, redox state, and content of redox active metals (e.g., manganese, iron), the consideration of this coupled biotic–abiotic N$_2$O formation in mechanistic models for simulation of soil N$_2$O emissions might become relevant.
2.4. Implications of abiotic processes in terrestrial N trace gas emissions

Although some purely abiotic processes, as described above, have been shown to contribute to the formation of N trace gases from soils under certain soil conditions, most studies merely focus on enzymatic pathways as the sole source of N trace gases. While some authors endorse the idea of the abiotic production of N trace gases, they still do not account for it in their studies (e.g., Baggs et al., 2010; Stevens et al., 1997; Zhang et al., 2011; Zhu et al., 2011). This may lead to a potential overestimation of biological processes and thus an underestimation of the significance of abiotic pathways.

Relatively few studies have tried to quantify the contribution of chemical processes to total N trace gas emissions from soils, especially at field-scale. Yamulki et al. (1997) observed NO fluxes from a sterilized soil core (pH 3.9) of 29% compared to the equivalent soil core before sterilization, that were sharply reduced with increasing pH. N₂O production was found to be negligible. These results coincide with more recent studies, which contributed higher NO emissions from acidic agricultural soils partially to chemical decomposition of NO₂⁻ (Cheng et al., 2004). In a laboratory study, Kesik et al. (2006) noticed that up to 62% of the production of NO was due to chemical decomposition of NO₂⁻ in acidic soils (pH <4), while contribution to N₂O production was only minor (0.8%). They observed no abiotic N trace gas production at a pH above 4.5. Ding et al. (2010) observed NO emissions from a sterilized agricultural soil at pH 8, possibly by a reaction of NO₂⁻ with SOM. At this high pH, NO emissions were reduced to 7–13% of the emissions prior to sterilization. Nevertheless, Venterea (2007) estimated that abiotic processes account for 31–75% of the total N₂O production of fertilized agricultural soils.

The effects of abiotic N trace gas production have been implemented into a process-oriented model for N₂O and NO emissions from temperate forest soils by Li et al. (2000). However, in this model, chemical processes are only responsible for the production of NO, as this model only considers “classical” chemodenitrification, i.e., chemical processes involving NO₂⁻/HNO₂. In their model, Li et al. (2000) calculated abiotic NO production as a function of nitrification rate, soil temperature (after Yamulki et al., 1997), and soil pH (after Blackmer and Cerrato, 1986). The implementation of the nitrification rate into the calculation emphasizes the interaction between biotic and abiotic processes.

The model validation by Stange et al. (2000), run on data from different field sites, showed that at a beech site with a soil pH of 4 approximately 9% of total NO was produced abiotically, while at a spruce site in the same forest with a lower pH of 3.2, abiotic NO production increased markedly to 30% of the total NO emission. However, the assumption of the model was a uniform bulk soil pH, but microsite variability in acid forest soils has been shown to be up to a scale of 3 pH units (Bruehlheide and Udelhoven, 2005; Kesik et al., 2005). The model was used by Kesik et al. (2005)
A review of chemical nitrogen trace gas formation reactions in soils

for an inventory of European forest soils. Highest NO emissions (7.0 kg N ha\(^{-1}\) yr\(^{-1}\)) were simulated for the Netherlands and the neighboring areas in Belgium and Germany, due mainly to the low soil pH in that region, high atmospheric N deposition rates, and a higher abiotic production of NO (Kesik et al., 2005).

Further approaches had been made using stable isotope labeling techniques, where the N substrates are labeled with \(^{15}\)N. Stevens et al. (1997) made experiments to differentiate between microbial nitrification and denitrification by labeling different substrate sub-pools and quantifying the \(^{15}\)N enrichments in \(N_2O\) emissions. However, the distinction between biotic and abiotic processes relying on the same substrate is difficult. Recent approaches of incorporating the information gained from these isotope tracing techniques into a \(^{15}\)N tracing model could help to further quantify \(N_2O\) emissions from different pathways (Müller et al., 2014). Although the authors did not include abiotic pathways in their model, they found that the majority of \(N_2O\) emissions from old grassland soil were not associated with autotrophic nitrification or denitrification.

In the recent past, \(N_2O\) emissions by nitrifying bacteria have been attributed to the so-called nitrifier denitrification (Wrage et al., 2001). Nitrifier denitrification is a facultative pathway of nitrifiers in which NO\(_2^-\) is reduced anaerobically to N\(_2\) via NO and \(N_2O\) by the same autotrophic microorganism (Wrage et al., 2001). The pathway from NH\(_4^+\) via NO\(_2^-\) to \(N_2O\) was introduced by Poth and Focht (1985), and further to \(N_2\) by Poth (1986). NH\(_3\) oxidizers may use NO\(_2^-\) as an alternative electron acceptor for O\(_2\) when being subjected to oxygen deficiency (Ritchie and Nicholas, 1972). Poth and Focht (1985) hypothesized that nitrite reductase and nitric oxide reductase, the same enzyme as utilized by microorganisms involved in denitrification, are responsible for the transformation. The presence of the two enzymes has been confirmed in the genome sequence of \(Nitrosomonas europaea\) by Chain et al. (2003). However, the enzyme responsible for \(N_2O\) reduction has not been identified in nitrifiers yet (Chapuis-Lardy et al., 2007). The genome revealed the interesting feature that this bacterium requires NH\(_3\) as a substrate, and no capability was found to use other inorganic sources of energy. There is still a lot of uncertainty about the contribution to \(N_2O\) emitted from soils that reach from insignificant to about 30% of total produced \(N_2O\) (Wrage et al., 2001). In a recent study it was shown, that nitrifier denitrification can be a major contributor to total \(N_2O\) production from soil and can outbalance conventional denitrification at WFPS of 50–70% (Kool et al., 2011).

However, most studies only show that biological NH\(_3\) oxidation is a crucial process for soil \(N_2O\) production under certain conditions, and all known microbial \(N_2O\) production pathways require anaerobic conditions, whereas the presented chemical reactions involving NH\(_2OH\) can lead to \(N_2O\) production also under aerobic conditions. Hooper and Terry (1979) already stated that the incomplete oxidation of the nitrification intermediate NH\(_2OH\) can lead to the development of \(N_2O\). This matches the “nitrification-unstable intermediate” hypothesis formulated but abandoned by Poth and...
Focht (1985), in which N$_2$O is produced during nitrification by various reactions of intermediates formed during NH$_3$ oxidation. This does not mean that all N trace gas production other than through classical, nitrifier-, or co-denitrification has to be completely chemical per se, but it is likely that there is an interlink between the biological nitrification pathway and purely chemical reactions: nitrification providing the intermediates, NH$_2$OH and NO$_2^-$, as reactants for chemical N$_2$O formation reactions. Thus, biological NH$_3$ oxidation to NO$_2^-$ can be a source of N$_2$O, indirectly via chemical reactions of reactive and unstable intermediates, as depicted in a conceptual model in Figure 2.1.

The proposed coupled mechanism of biological NH$_3$ oxidation and chemical oxidation of NH$_2$OH could also easily explain the non-detection of N$_2$O emission from sterile soil samples, even with added NO$_2^-$, by some authors (e.g., Ding et al., 2010; Mørkved et al., 2007; Mummey et al., 1994; Yamulki et al., 1997). While NO$_2^-$ decomposition mainly produces NO, NH$_2$OH seems to be the key component for the abiotic N$_2$O production in soils. All chemical processes that have been found relevant to produce N$_2$O in soils require NH$_2$OH as precursor (Udert et al., 2005). This can be explained in the sense that the reactions proceed as an N comproportionation (Spott et al., 2011). N$_2$O is formed by the reaction between a nucleophilic N with the formal oxidation state $\text{−}1$ (NH$_2$OH) with an electrophilic N species when the formal oxidation state is $\text{+}3$, like in NO$_2^-$ or a nitrosonium cation (NO$^+$) in a nitrosation reaction (Spott et al., 2011). Due to the highly reactive character, NH$_2$OH will usually not accumulate in soils. However, Liu et al. (2014) developed a highly sensitive method for the detection of NH$_2$OH in soils and found a significant linear relationship between NH$_2$OH and N$_2$O formation under aerobic conditions in an acidic spruce forest soil.

As NH$_2$OH might also be immobilized by SOM, making it unavailable for reactions leading to N$_2$O, it requires active nitrification, providing new NH$_2$OH, to trigger abiotic N$_2$O production. Therefore, it can possibly not be observed in sterile soils and will lead to an underestimation of abiotic N$_2$O production in soil and a misinterpretation of the underlying mechanisms.

Another important step for a better understanding and future modeling of N in soils will be the coupling of different biogeochemical cycles. A good example is the ferrous wheel hypothesis (Davidson et al., 2003). Although being challenged by some authors (Colman et al., 2008; Schmidt and Matzner, 2009), the approach of Davidson et al. (2003) to combine different cycles is definitely pointing to the right direction. Biogeochemical cycles cannot be looked at in an isolated manner when trying to understand soil processes. The gearing of different element cycles into each other will make modeling of N transformation in soil more complex than it already is, but is needed for a better understanding of the different processes involved and will improve predictions of N trace gas emissions from soils.
2.5. Outlook

This review highlights the tight connection between biotic and abiotic processes in the terrestrial N cycle and likely other biogeochemical cycles. At the same time it demonstrates the challenge to disentangle the diverse processes all competing for N in soil. Experiments with sterilized soils failed to detect abiotic N\textsubscript{2}O production because, as this review emphasizes, it involves very likely a coupled biotic–abiotic production mechanism, so that experiments with non-sterile soils are needed to be able to show that chemical N trace gas production from microbiological intermediates is happening, and to define the relative importance of different processes. The greatest issue in studying N trace gas emissions from soils is the identification of the origin of the N trace gas, especially at the field scale. With previous methods it is not possible to separate biotic and abiotic sources and sinks satisfactorily, so that at present it is not possible to clearly determine the contribution of abiotic processes to total N trace gas emission from soils.

While inhibition methods have proven to be not very reliable for specific soil conditions or specific microorganisms, stable isotopes have the greatest potential to overcome this problem (Wrage et al., 2005). Stable isotopes are of great value for determining the importance and regulation of previously ignored pathways, and the use of stable isotope techniques can show that the current view on these processes is over-simplistic (Baggs, 2008). Advances in mass spectrometry and, lately, in laser spectroscopy enable the use of stable isotopes for the differentiation between processes, but techniques need to be further developed to quantify the different processes. A source partitioning is essential for closing the N\textsubscript{2}O budget and for understanding controls of processes to develop appropriate mitigation strategies (Baggs, 2008).
One approach is the natural abundance of $^{15}\text{N}$ or $^{18}\text{O}$ in $\text{N}_2\text{O}$ that has the potential for source partitioning between different processes, but fractionation – especially for biological processes – can differ over a large range of $\delta^{15}\text{N}$ and $\delta^{18}\text{O}$ values. At present, the success is limited by insufficient knowledge on fractionation factors of different pathways (Well and Flessa, 2009), although some advances have been made with respect to $\text{N}_2\text{O}$ production during denitrification (Lewicka-Szczebak et al., 2015; Rohe et al., 2014). An emerging approach is the site-specific determination of $^{15}\text{N}$ in $\text{N}_2\text{O}$, the differentiation between $\text{N}_2\text{O}$ isotopomers. The site preference (SP) is defined as the difference between the $^{15}\text{N}$ isotope ratio in the central ($\alpha$) and terminal ($\beta$) N atom (SP $= \delta^{15}\text{N}_\alpha - \delta^{15}\text{N}_\beta$) (Toyoda and Yoshida, 1999). Since the introduction of the method, the source partitioning using isotopomers has been a central research objective. The intramolecular distribution of $^{15}\text{N}$ in $\text{N}_2\text{O}$ is supposed to reflect the production mechanism rather than the substrate because it is independent from the isotopic signature of the precursor N species (Samarkin et al., 2010; Well et al., 2008). The bulk $\delta^{15}\text{N}$ values for $\text{N}_2\text{O}$ produced by the different biotic and abiotic reactions are of limited use, as the reactions feed on various N sources and the $\delta^{15}\text{N}$ is affected by the substrate. Yoshida and Toyoda (2000) found that the SP of $^{15}\text{N}$ in $\text{N}_2\text{O}$ varied significantly throughout the atmosphere, with low SP in the troposphere indicating local emissions. They stated that the SP is a tool with the potential to increase the partitioning of $\text{N}_2\text{O}$ sources and sinks. Recently, Heil et al. (2014) showed that the SP of $\text{N}_2\text{O}$ produced from different abiotic NH$_2$OH oxidation processes was very stable over time and in the same range as $\text{N}_2\text{O}$ production observed from AOB or AOA. By this, the latest results indicate that the SP can only be used to distinguish between reductive (denitrification and nitrifier denitrification) and oxidative (NH$_2$OH oxidation, both abiotically and by bacteria and archaea) $\text{N}_2\text{O}$ production (Decock and Six, 2013). However, there is still insufficient data available, so that more research is required.

This review showed a variety of purely abiotic pathways involved in soil NO and $\text{N}_2\text{O}$ emissions, being neglected in most studies, although being feasible over a wide range of soil properties. The review also emphasizes the tight coupling between the biotic nitrification and abiotic trace gas production. These processes could have great implications for our general understanding and modeling of N trace gas emissions from soils. However, more research is needed to quantitatively assess the importance of these abiotic pathways at field-scale and ecosystem level.
Chapter 3

Site-specific $^{15}\text{N}$ isotopic signatures of abiotically produced N$_2$O

Based on:
3.1. Introduction

N$_2$O is a powerful greenhouse gas with an approximately 300 times higher global warming potential than CO$_2$. Moreover, N$_2$O is today’s single most important ozone depleting substance (Ravishankara et al., 2009). The atmospheric mixing ratio of N$_2$O has increased from pre-industrial 270 ppb to 324 ppb in 2011 and is still increasing due to anthropogenic activities, at a rate of 0.8 ppb yr$^{-1}$ (WMO, 2013). Soils, predominantly agricultural soils, are a major source of N$_2$O, contributing an estimated 50–60% to global N$_2$O emissions (USEPA, 2010). However, it has recently been observed that soils can also, at least temporarily, function as a significant N$_2$O sink (Chapuis-Lardy et al., 2007; Goldberg and Gebauer, 2009). Based on top-down estimates, global N$_2$O emissions can be derived from the stratospheric loss and the atmospheric increase. Still the strength of individual source processes and possible sinks is rather uncertain. This is reflected by the large uncertainty of the bottom-up estimate of global N$_2$O emissions ranging from 8.5 to 27.7 Tg N yr$^{-1}$ (Denman et al., 2007). This great uncertainty can be caused by either overestimating N$_2$O sources or disregarding N$_2$O sinks (Billings, 2008).

Microbial nitrification and denitrification are considered the major processes responsible for soil N$_2$O emissions. The N$_2$O release during both processes has been conceptualized by Firestone and Davidson (1989) in their ‘hole-in-the-pipe’ model, which attributes N$_2$O emissions from soils during nitrification and denitrification to a ‘leaky’ N flow from NH$_4^+$ to NO$_3^-$ and the incomplete stepwise reduction of nitrate to molecular nitrogen (N$_2$). Today, the two main processes considered responsible for N$_2$O production during nitrification are hydroxylamine (NH$_2$OH) oxidation, i.e., the production of N$_2$O as a by-product of biological NH$_2$OH oxidation (Sutka et al., 2003), and nitrifier denitrification, the reduction of nitrite (NO$_2^-$) by nitrifying bacteria under oxygen-limiting conditions or at elevated NO$_2^-$ concentrations (Wrage et al., 2001). However, abiotic reactions were also identified as potential sources of N$_2$O (Bremner, 1997). Possible substrates are the two nitrification intermediates NH$_2$OH and NO$_2^-$, in the presence of iron compounds or other transition metals. Although these reactions have been known for several years (Bremner et al., 1980), they are neglected in most current nitrogen (N) trace gas studies.

Stable isotope techniques, especially $^{15}$N-isotopomer analysis of N$_2$O, have a great potential to disentangle the different processes leading to N$_2$O formation. The first site-specific analysis of N$_2$O produced by the chemical processes NO$_2^-$ reduction and NH$_2$OH oxidation indicated a constant SP of approximately 30‰ for both reactions (Toyoda et al., 2005). Only recently, Wunderlin et al. (2013) confirmed this finding for abiotic NH$_2$OH oxidation. However, the SP of N$_2$O produced via different NH$_2$OH oxidation pathways and experimental conditions has not yet been studied. Hence, the influence of these factors on the isotopic signature of N$_2$O is still unknown.
The aim of the present laboratory study was to investigate $\delta^{15}\text{N}_{\text{bulk}}$, $\delta^{18}\text{O}$, and SP values of N$_2$O formed by different abiotic reactions that could potentially occur in soils under a range of process conditions. Substrates were the nitrification intermediates NH$_2$OH and NO$_2^-$ that have been shown to produce significant amounts of N$_2$O. In preliminary experiments those nitrification intermediates have been reacted with each other and iron oxides at different pH levels to observe the production of the N trace gases NO and N$_2$O using quantum cascade laser absorption spectroscopy. The site-specific isotopic ratios of N$_2$O from reactions that have shown to produce significant amounts of N$_2$O were then analyzed in real time using the same technology.

3.2. Materials and Methods

3.2.1. Preliminary experiments

In preliminary experiments reactions were conducted in aqueous solution using deionized water. Solutions contained 1 mM NH$_2$OH (NH$_2$OH–HCl) and, depending the treatment, combinations of 1 mM NO$_2^-$ (NaNO$_2$), and 2 mM Fe$^{3+}$ (FeCl$_3$·6 H$_2$O) (all chemicals were reagent grade or better and obtained from Merck, Germany). Reaction solutions were either unbuffered or buffered, with citrate buffer for the pH range 3–5, and Tris–maleate buffer for pH 6. Single solutions were mixed in a 250-mL beaker to total up to 100 mL and placed in reaction chambers that were immediately closed afterwards. The custom-made chambers consisted of quartz glass cylinders equipped with PTFE cover plates. The PTFE plates and the quartz glass cylinder were clamped together with seven aluminum rods at the outside of the chamber and the contact surfaces were sealed with Viton O-rings. Uniform mixing of the gas phase inside the chamber was assured with a fan. The chamber volume was 2 L.

The reaction chamber was purged with 3.4 L min$^{-1}$ of dried, pressurized air (Forschungszentrum Jülich GmbH, Germany) controlled by a mass flow controller (MFC, Brooks Instruments, Germany). The sample gas was then split, and one part was directed to a QCLAS (Dual Laser Quantum Cascade Trace Gas Monitor, Aerodyne Research Inc., USA) for N$_2$O measurement and the other part to a NO chemiluminescence detector (AC32M, Ansyco GmbH, Germany) via PTFE tubing. The QCLAS was operated with two mid-infrared lasers for simultaneous measurements of N$_2$O, CO$_2$, CH$_4$, and H$_2$O concentrations at a high temporal resolution. Water vapor was measured together with the other gases to be able to correct for volumetric dilution and pressure broadening effects due to changing water vapor concentrations in the sample air. Prior to every experiment, the QCLAS was flushed with synthetic air, and a new background spectrum was taken. The instrumental precision of the measurements, given as standard deviation of the average N$_2$O mixing ratio at atmospheric level, was <0.3 ppb. The chemiluminescence detector was based on the principle of
measuring light emissions from the chemical reaction between NO and ozone. The reaction is selective for NO and linear over a range of 0 to 10000 ppb, which embraced the range of NO mixing ratios during the preliminary experiments. The detector had a detection limit of 1 ppb.

3.2.2. Laboratory setup for isotope-specific N₂O measurements

Reactions were conducted in aqueous solution using ultrapure water (18.2 MΩ cm). All solutions contained NH₂OH (NH₂OH–HCl; δ¹⁵N = −1.93 ± 0.11‰) and, depending on the treatment, combinations of NO₂⁻ (NaNO₂; δ¹⁵N = −27.70 ± 0.08‰), Fe³⁺ (FeCl₃ · 6 H₂O), Fe²⁺ (FeCl₂ · 4 H₂O) and Cu²⁺ (CuSO₄ · 5 H₂O) (all chemicals were reagent grade or better and obtained from Merck, Germany) (see Table 3.1). Solutions were freshly prepared every day and mixed immediately before measurement. Reaction solutions were either unbuffered or buffered, with phosphate–citrate buffer for the pH range 3–4, and Tris–maleate buffer for the pH range 5–8 (Table 3.1). The pH of the reaction solutions was measured with a pH electrode (SenTix® 81, WTW, Germany), calibrated at pH 4 and 7. Single solutions were mixed in three 250-mL beakers and placed in three replicate reaction chambers that were immediately closed. The same custom-made chambers as in the preliminary experiments were used, but the chamber volume of 2 L was reduced to 500 mL by a polypropylene insert with a recess for a 250-mL beaker.

<table>
<thead>
<tr>
<th>experiment No.</th>
<th>substrates and concentrations</th>
<th>volume [ml]</th>
<th>buffer system</th>
<th>pH</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>NH₂OH 1 mM + NO₂⁻ 1 mM</td>
<td>100</td>
<td>phosphate–citrate</td>
<td>3.0</td>
</tr>
<tr>
<td>2</td>
<td>NH₂OH 1 mM + NO₂⁻ 1 mM</td>
<td>100</td>
<td>phosphate–citrate</td>
<td>4.0</td>
</tr>
<tr>
<td>3</td>
<td>NH₂OH 1 mM + Fe³⁺ 2 mM</td>
<td>100</td>
<td>unbuffered</td>
<td>2.8</td>
</tr>
<tr>
<td>4</td>
<td>NH₂OH 1 mM + Fe³⁺ 5 mM</td>
<td>200</td>
<td>Tris–maleate</td>
<td>5.0</td>
</tr>
<tr>
<td>5</td>
<td>NH₂OH 1 mM + Cu²⁺ 0.1 mM</td>
<td>100</td>
<td>Tris–maleate</td>
<td>8.0</td>
</tr>
<tr>
<td>6</td>
<td>NH₂OH 0.5 mM + NO₂⁻ 0.5 mM + Fe³⁺ 1 mM</td>
<td>100</td>
<td>unbuffered</td>
<td>2.8</td>
</tr>
<tr>
<td>7</td>
<td>NH₂OH 1 mM + NO₂⁻ 1 mM + Fe³⁺ 2 mM</td>
<td>200</td>
<td>Tris–maleate</td>
<td>5.5</td>
</tr>
<tr>
<td>8</td>
<td>NH₂OH 0.25 mM + NO₂⁻ 0.25 mM + Fe³⁺ 0.5 mM + Fe²⁺ 0.5 mM</td>
<td>100</td>
<td>unbuffered</td>
<td>2.8</td>
</tr>
<tr>
<td>9</td>
<td>NH₂OH 2 mM + NO₂⁻ 2 mM + Fe³⁺ 4 mM + Fe²⁺ 4 mM</td>
<td>200</td>
<td>Tris–maleate</td>
<td>5.5</td>
</tr>
</tbody>
</table>
Figure 3.1: Schematic representation of the laboratory setup for the determination of abiotic N₂O production with dynamic flow-through chambers coupled to a quantum cascade laser absorption spectrometer (QCLAS).

As illustrated in Figure 3.1, reaction chambers were purged with 50 mL min⁻¹ of high purity synthetic air (20.5% O₂ in 79.5% N₂, purity 99.999%, Messer, Switzerland) controlled by three MFCs (red-y smart series, Vögtlin, Switzerland), via a multi MFC process control unit (PCU 1000, Vögtlin, Switzerland). At the outlet of each chamber, sample gas flows of individual chambers were dehumidified by separate Nafion® permeation dryers (MD-050-72S-1, Perma Pure, USA). The sample gas flows from the three reaction chambers were combined and guided through a Sofnocat® column (9.1 g) to remove traces of CO which would otherwise lead to spectral interferences in the quantum cascade laser absorption spectrometer (QCLAS). The sample gas was then directed to the QCLAS, where a flow of around 46 mL min⁻¹ was passing the spectrometer multipath absorption cell. The approximate flow rate was adjusted with a manual valve at the outlet of the cell, while the exact inflow was controlled by a pressure controller (red-y smart series, Vögtlin, Switzerland) keeping the cell pressure at 33.3 hPa. Switching between measuring and calibration gas was accomplished by two 3-way valves (series 9, Parker Hannifin, USA).
Table 3.2: Summary of the results for the studied abiotic reactions. The uncertainty is given as the standard deviation of the 60-s moving average.

<table>
<thead>
<tr>
<th>experiment</th>
<th>N₂O mixing ratio [ppm]</th>
<th>N turnover [%]</th>
<th>$\delta^{15}N^{bulk}$ [%]</th>
<th>$\Delta \delta^{15}N$ [%]</th>
<th>$\delta^{18}O$ [%]</th>
<th>site preference (SP) [%]</th>
</tr>
</thead>
<tbody>
<tr>
<td>No.</td>
<td>avg.</td>
<td>max</td>
<td>120min total</td>
<td>start</td>
<td>end</td>
<td>avg.</td>
</tr>
<tr>
<td>1</td>
<td>36.8</td>
<td>117.6</td>
<td>24.2 64.9</td>
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<td>-19.9</td>
<td>-22.9 ± 2.9</td>
</tr>
<tr>
<td>2</td>
<td>25.1</td>
<td>63.7</td>
<td>12.2 53.8</td>
<td>-32.6</td>
<td>-22.4</td>
<td>-26.9 ± 3.3</td>
</tr>
<tr>
<td>3</td>
<td>37.6</td>
<td>89.2</td>
<td>18.5 18.5</td>
<td>-2.7</td>
<td>3.6</td>
<td>1.4 ± 1.9</td>
</tr>
<tr>
<td>4</td>
<td>23.6</td>
<td>41.4</td>
<td>8.0 15.6</td>
<td>-23.0</td>
<td>-19.1</td>
<td>-21.7 ± 0.9</td>
</tr>
<tr>
<td>5</td>
<td>39.4</td>
<td>53.9</td>
<td>19.4 19.4</td>
<td>-7.2</td>
<td>-19.3</td>
<td>-15.1 ± 4.0</td>
</tr>
<tr>
<td>6</td>
<td>20.4</td>
<td>88.0</td>
<td>20.5 42.1</td>
<td>-14.2</td>
<td>-14.6</td>
<td>-14.4 ± 0.1</td>
</tr>
<tr>
<td>7</td>
<td>6.2</td>
<td>8.4</td>
<td>0.9 3.3</td>
<td>-19.6</td>
<td>-23.5</td>
<td>-22.5 ± 1.2</td>
</tr>
<tr>
<td>8</td>
<td>25.2</td>
<td>46.8</td>
<td>24.9 24.9</td>
<td>-14.0</td>
<td>-13.1</td>
<td>-13.8 ± 0.5</td>
</tr>
<tr>
<td>9</td>
<td>17.4</td>
<td>20.1</td>
<td>0.9 4.6</td>
<td>-18.2</td>
<td>-22.9</td>
<td>-22.7 ± 0.2</td>
</tr>
</tbody>
</table>

*In experiments with two N substrates a 1:1 reaction between the educts was assumed and the average of the $\delta^{15}N$ of the educts used to calculate $\Delta \delta^{15}N$.

*b concentrations in this experiment were below the calibration limit.
3.2.3. Instrumentation

The employed laser spectrometer (custom-made, Aerodyne Research Inc., USA) was based on continuous wave quantum cascade laser (cw-QCL) technology, and was originally developed for high precision NOx measurements (Tuzson et al., 2013). For the measurement of N₂O isotopologues, the QCL was replaced, the optics optimized and the software reconfigured. Major improvements over the spectrometer previously reported for N₂O isotopic analysis (Waechter et al., 2008) include: a) optimally selected spectral range around 2203 cm⁻¹, which allows for the first time the simultaneous quantification of the four most abundant N₂O isotopologues (¹⁴N¹⁴N¹⁶O, ¹⁵N¹⁴N¹⁶O, ¹⁴N¹⁵N¹⁶O, and ¹⁴N¹⁴N¹⁸O); b) increased sensitivity due to cw operation of the QCL (Alpes Lasers, Switzerland) and employing a longer astigmatic Herriott multi-pass absorption cell (204 m path length, AMAC-200, Aerodyne Research Inc., USA); and c) a reference path with a short (5 cm) N₂O–filled cell to lock the laser emission frequency to well-defined absorption lines.

The spectrometer was operated in a flow-through mode and mixing ratios of N₂O isotopologues were measured at 1 Hz temporal resolution. For N₂O mixing ratios of 50 ppm and a cell pressure of 3.3 kPa the spectrometer enables high precision (< 0.05‰) analysis of δ¹⁵N⁺, δ¹⁵N⁻, and δ¹⁸O with 450 s spectral averaging. The 1-s precision for delta values is < 0.6‰. This is a factor 2–3 superior to our previously published results (Mohn et al., 2010; Mohn et al., 2012). The multi-pass cell was flushed with synthetic air every three hours to record a new background spectrum. For this period no measurement data are available. The spectroscopically determined isotope ratios were related to the international isotope ratio scales (air–N₂ for ¹⁵N/¹⁴N, VSMOW for ¹⁸O/¹⁶O) through analysis of calibration gases before and after every experiment. Calibration gases were prepared in the laboratory at EMPA based on gravimetric and dynamic dilution methods from pure medical N₂O (Messer, Switzerland) supplemented with distinct amounts of isotopically pure ¹⁵N¹⁴N¹⁶O (> 98%, Cambridge Isotope Laboratories, USA) and ¹⁴N¹⁴N¹⁶O (> 99.95%, ICON Services, USA). Primary calibration gases have been analyzed for δ¹⁵N⁺, δ¹⁵N⁻, δ¹⁸O by IRMS at Tokyo Institute of Technology (Toyoda and Yoshida, 1999). Additionally, comparability of our results with IRMS for δ¹⁵N⁺ and δ¹⁵N⁻ has been demonstrated in an inter-comparison approach (Köster et al., 2013). The N₂O turnover was calculated from total N₂O emissions and N input.

The abundances of different isotopic species are reported in the δ-notation. The δ¹⁵N bulk value is defined as δ¹⁵Nbulk = (δ¹⁵N⁰ + δ¹⁵N⁸)/2, where δ¹⁵N⁰ denotes the relative ¹⁵N abundance at the central (¹⁴N¹⁵N¹⁶O) and δ¹⁵N⁸ at the terminal N position (¹⁵N¹⁴N¹⁶O) of the asymmetric N₂O molecule (Brenninkmeijer and Röckmann, 1999; Toyoda and Yoshida, 1999). The SP is defined as difference in isotope deltas at central and terminal position δ¹⁵N⁰ – δ¹⁵N⁸. The net N isotope effect (NIE) is the difference in δ¹⁵N between the relevant substrate and the released N₂O and was approximated as Δδ¹⁵N = δ¹⁵Nsubstrate – δ¹⁵Nbulk (Sutka et al., 2003).
The first 15 min of each experiment were discarded due to mixing with ambient air from the chambers. Averages of isotopic delta values were calculated for the first 5 min after that period, for the last 5 min, as well as over the complete experimental run. The NIE was only calculated for the initial 5 min of the experiment after the discarded period. N$_2$O threshold mixing ratios for the use of isotope data were 18 ppm as lower and 90 ppm as upper boundary, respectively. Measurement uncertainties were calculated as the standard deviation of the 60-s moving average.

**Figure 3.2:** Mixing ratios of N$_2$O (red line) and NO (blue line) produced by the abiotic reaction between 1 mM NH$_2$OH and 1 mM NO$_2^-$ buffered (citrate buffer) (A) at pH 3, (B) at pH 4, (C) at pH 5, and (D) unbuffered (pH 3.4) with the addition of 2 mM Fe$^{3+}$; All values are 1 Hz data.

### 3.3. Results

#### 3.3.1. NO and N$_2$O production from nitrification intermediates NH$_2$OH and NO$_2^-$

The first N trace gas production reactions studied were the reaction between NH$_2$OH and NO$_2^-$ at different pH levels, and additionally in the presence of Fe$^{3+}$ (Fig. 3.2). The reaction between NH$_2$OH and NO$_2^-$ buffered at pH 3 (Fig. 3.2a) resulted in a strong NO and N$_2$O production right at the beginning of the experiment. NO mixing ratios reached the maximum only a few minutes after the onset of the reaction at 398 ppb. N$_2$O mixing ratios increased for about one hour until a maximum of about 680 ppb was reached. After reaching their peak values, both mixing ratios decreased slowly for the rest of the experiment and approached zero after approximately 15 h. The same reac-
tion at pH 4 changed the kinetics of the NO and N₂O production (Fig. 3.2b). NO production also reached the maximum several minutes after start of the experiment, but at a lower value of only 133 ppb. The N₂O production kinetics were also different, with a lower maximum (490 ppb), but a longer tailing with N₂O mixing ratios still at about 30 ppb after 18 h. At pH 5 (Fig. 3.2c), the same reaction resembled completely different kinetics that resulted in a low but constant production of NO and N₂O after an initial small NO peak and an initial slow increase of N₂O mixing ratios. The final mixing ratios of NO and N₂O at the end of the experiment, after 15 h, were 4 and 88 ppb, respectively. The addition of Fe³⁺ to the two nitrifications intermediates NH₂OH and NO₂⁻, unbuffered at pH 3.4, changed the kinetics of NO and N₂O productions dramatically (Fig. 3.2d). The reaction showed an immediate production of both NO and N₂O, with the maximum value reached right after the beginning of the experiment. The N₂O maximum was also much higher, at about 2850 ppb, than previously observed without the addition of Fe³⁺. The maximum value of NO (398 ppb) was at the same level as without Fe³⁺ addition at pH 3.

Figure 3.3: Mixing ratios of N₂O (red line) and NO (blue line) produced by the abiotic reaction between 1 mM NH₂OH and 2 mM Fe³⁺, (A) unbuffered at pH 3.4 and (B) buffered (Tris-maleate) at pH 6; All values are 1 Hz data.

Experiments in the absence of NO₂⁻, demonstrating the reaction between NH₂OH and a redox-active transition metal, in particular Fe³⁺, are presented in Figure 3.3. In unbuffered solution, the reaction was associated with high N₂O formation that led to a maximum N₂O mixing ratio of about 390 ppb only 15 min after the onset of the experiment (Fig. 3.3a). Thereafter, the N₂O mixing ratio decreased until no more N₂O formation was observed after 10 h. Buffered at pH 6, the same reaction showed similar kinetics, however, the observed maximum was about four times lower (Fig.3.3b). In both experiments no significant NO formation was observed.
3.3.2. Isotopic signatures of abiotically produced N₂O

The first mechanism of abiotic N₂O production studied was the reaction between NH₂OH and NO₂⁻ at pH 3 and 4 (experiments 1 and 2 in Table 3.1). Both reactions resulted in a considerable N₂O production with a higher N₂O yield and maximum mixing ratio at the lower pH (Table 3.2). In Figure 3.4, N₂O mixing ratios and isotopic composition are given for the reaction at pH 4. At the onset of the experiment a fast increase in the N₂O mixing ratio from ambient to 64 ppm was observed, followed by a steady decline to 6 ppm within the following 17 h (Fig. 3.4a). Within this period, about 50% of the added N had been converted to N₂O-N (Table 3.2). The strong increase of N₂O mixing ratios in the headspace of the chamber in the beginning co-occurred with a decrease of δ¹⁵Nα and δ¹⁵Nβ (Fig. 3.4b), caused by mixing of newly formed N₂O with ambient N₂O (324 ppb, δ¹⁵Nbulk = 6.6‰, δ¹⁸O = 44.2‰, SP = 18‰; Toyoda et al., 2013) that was present in the reaction chamber during closure. This initial decrease in δ¹⁵Nα and δ¹⁵Nβ was followed by a constant increase in both isotope deltas, due to a constant enrichment of the substrate in ¹⁵N due to kinetic fractionation. As δ¹⁵Nα and δ¹⁵Nβ became enriched at the same rate, their difference, i.e., the SP, remained constant (Fig. 3.4d). The consistency of SP at 34.3 ± 0.4‰ (experiment 1) and 34.0 ± 0.5‰ (experiment 2) was not affected by an increase in the measurement uncertainty with decreasing N₂O mixing ratios. The NIE was calculated from the average δ¹⁵N of the two N substrates and the δ¹⁵Nbulk of initially emitted N₂O. It was slightly higher for the reaction at pH 4 (17.8‰) than at pH 3 (15.7‰), which could be explained by the lower reaction rate at higher pH. A similar, yet less pronounced behavior as for δ¹⁵N was observed for δ¹⁸O, with a moderate rise of 3.9‰ over the experiment after a strong increase in the first 10 min (Fig. 3.4c).

The second abiotic N₂O production mechanism analyzed was the oxidation of NH₂OH with Fe³⁺ in unbuffered solution as well as in solution buffered at pH 5 (experiments 3 and 4 in Table 3.1 and 3.2). The experiment in unbuffered solution yielded higher maximum N₂O mixing ratios and a faster N turnover rate and led to a minor ¹⁵N isotopic fractionation. For the experiment with Tris-maleate buffer (Fig. 3.5), a rapid increase of N₂O mixing ratio up to 41 ppm was observed within 40 min after the start of the experiment, followed by a slow decrease to 11 ppm after 5 h. Within these 5 h, 16% of the added N substrate had been transformed to N₂O (Table 3.2). The δ¹⁵Nα and δ¹⁵Nβ values showed a slow but steady increase of 3.5‰ during the experiment (Fig. 3.5b). A similar behavior was also observed for δ¹⁸O values (Fig. 3.5c), while the SP averaged at 34.9 ± 0.4‰ (Fig. 3.5d). The NIE for the buffered experiment was 21.1‰, while it was significantly lower for the unbuffered experiment (0.8‰) as explained above (Table 3.2).
Figure 3.4: Results of the isotopic analysis of N₂O produced by the abiotic reaction between 1 mM NH₂OH and 1 mM NO₂⁻, buffered at pH 4 (citrate–phosphate buffer): N₂O mixing ratio in ppm (A), δ¹⁵Nbulk, δ¹⁵Nα, and δ¹⁵Nβ of N₂O (B), δ¹⁸O of N₂O (C), and the N₂O site preference (SP) (D); All values are 1 Hz data, red lines show the 60 s moving average.

Figure 3.5: Results of the isotopic analysis of N₂O produced by the abiotic reaction between 1 mM NH₂OH and 5 mM Fe³⁺ buffered at pH 5 (Tris–maleate buffer): N₂O mixing ratio in ppm (A), δ¹⁵Nbulk, δ¹⁵Nα, and δ¹⁵Nβ of N₂O (B), δ¹⁸O of N₂O (C), and the N₂O site preference (SP) (D); All values are 1 Hz data, red lines show the 60 s moving average.
The last reaction mechanism investigated was the autoxidation of NH$_2$OH catalyzed by Cu$^{2+}$ at pH 8 (Fig. 3.6; experiment 5 in Table 3.1 and 3.2). This third reaction led also to a fast increase in N$_2$O mixing ratios to a maximum of 54 ppm after 50 min, before it rapidly declined to 19 ppm within another 90 min (Fig. 3.6a). The NH$_2$OH conversion rate to N$_2$O was 19% within this period. The initially emitted N$_2$O was depleted in $^{15}$N as compared to the NH$_2$OH substrate ($\Delta \delta^{15}$N = 5.3‰). Both $\delta^{15}$N$^a$ and $\delta^{15}$N$^b$ steadily decreased afterwards, once ambient air had been flushed out of the chamber. As both $\delta^{15}$N$^a$ and $\delta^{15}$N$^b$ decreased at the same rate, the SP remained constant at 34.4 ± 0.3‰ (Fig. 3.6d). The $\delta^{18}$O of N$_2$O showed a rapid increase during the whole experiment by over 20‰ to a maximum of 65‰, much higher than in the other reactions (Fig. 3.6c).

Figure 3.6: Results of the isotopic analysis of N$_2$O produced by abiotic autoxidation of 1 mM NH$_2$OH catalyzed by 0.1 mM Cu$^{2+}$ buffered at pH 8 (Tris–maleate buffer): N$_2$O mixing ratio in ppm (A), $\delta^{15}$N$_{\text{bulk}}$, $\delta^{15}$N$^a$, and $\delta^{15}$N$^b$ of N$_2$O (B), $\delta^{18}$O of N$_2$O (C), and the N$_2$O site preference (SP) (D); All values are 1 Hz data, red lines show the 60 s moving average.

Ternary and quaternary unbuffered reaction mixtures (experiments 6 and 8, Table 3.1) with two possible reaction mechanisms, i.e., the reaction of NH$_2$OH with NO$_2^-$ and the oxidation of NH$_2$OH by Fe$^{3+}$, led to a fast and sharp increase in N$_2$O production. Due to the addition of Fe$^{3+}$, the solution was acidic with a pH of 2.8. Although the concentration of both N substrates was only half or a quarter of experiments 1 to 5, 15 min after the start the maximum N$_2$O mixing ratio peaked at 88 ppm in experiment 6. However, the N turnover to N$_2$O within the first 2 h (20.5%) was slightly lower compared to the reaction between NH$_2$OH and NO$_2^-$ at a similar pH (experiment 1; 24.2%);
The $\delta^{15}N_{\text{bulk}}$ of N$_2$O emitted at the onset of the experiment was similar to the average $^{15}N$ content of the two N substrates ($\Delta\delta^{15}N = -1.4$). Nevertheless, one has to consider that $\Delta\delta^{15}N$ is only indicative for reactions involving both N substrates, as N$_2$O production could also be from NH$_2$OH only.

In experiment 8, the combined influence of Fe$^{2+}$ and Fe$^{3+}$ on the reaction between NH$_2$OH and NO$_2^-$ was investigated in unbuffered solution. Concentrations of both N substrates were half compared to experiment 6. The maximum observed N$_2$O mixing ratio was also nearly half at 47 ppm. There was no significant difference between $\delta^{15}N$ and $\delta^{18}O$ of N$_2$O produced in experiments 6 and 8, and in both experiments only a marginal isotopic fractionation was observed assuming a 1:1 reaction of NH$_2$OH and NO$_2^-$ as N$_2$O educts. The reaction in the presence of both Fe$^{2+}$ and Fe$^{3+}$ had a slightly higher N turnover within the first 2 h of the experiment (24.9%) as compared to experiment 6 with Fe$^{3+}$ alone (20.5%).

![Figure 3.7: Relationship between all 1-min average $\delta^{15}N_{\text{bulk}}$ and $\delta^{18}O$ values of the produced N$_2$O of all experiments.](image)

In buffered solution (pH 5.5), the results of the reaction of NH$_2$OH and NO$_2^-$ with Fe$^{3+}$ and with Fe$^{2+}$/Fe$^{3+}$, respectively (experiments 7 and 9, Table 3.1), were completely different from those of the unbuffered reactions (experiments 6 and 8). Although N substrate concentrations and solution volume were higher, N$_2$O production was much lower (Table 3.2). Also the kinetics of the buffered reactions was completely different from the unbuffered reactions. In experiment 7, an increase in N$_2$O mixing ratio in the headspace up to 8.4 ppm after 45 min was observed, followed by a slow gradual decrease. In experiment 9 the initial fast increase was followed by a very slow but succes-
sive increase until the end of the experiment after 500 min. $\delta^{15}$N$^a$ and $\delta^{15}$N$^b$ for experiments with or without Fe$^{2+}$ addition showed a similar behavior with minor isotopic fractionation at the onset ($\Delta \delta^{15}$N$^a = 4.8\%$, experiment 7; $\Delta \delta^{15}$N$^b = 3.4\%$, experiment 9). In the course of the experiment $\delta^{15}$N$^a$ and $\delta^{15}$N$^b$ gradually decreased by around 5%. In addition, both experiments were characterized by a very low N turnover rate in the first 2 h of 0.9%.

The 1-min averages of the $\delta^{15}$N bulk and $\delta^{18}$O values plotted against each other showed a strong relationship between the two delta values for all individual experiments (Fig. 3.7). N$_2$O produced by the different abiotic reactions showed a linear correlation between $\delta^{15}$N bulk and $\delta^{18}$O in dependence on the reaction time, those relationships, however, differed completely between different experiments, showing the great variability in fractionation between the different reactions, although all were abiotic NH$_2$OH oxidations. Only same reactions in the same buffer medium showed similar fractionation patterns (experiment 1 and 2).

### 3.4. Discussion

#### 3.4.1. NO and N$_2$O production from nitrification intermediates NH$_2$OH and NO$_2^-$

Preliminary experiments have shown that reactions involving the nitrification intermediates NH$_2$OH and NO$_2^-$ can lead to the production of N trace gases NO and N$_2$O (Fig. 3.2 and 3.3). The production of NO was restricted to reactions involving NO$_2^-$, while in the absence of NO$_2^-$ no NO formation could be observed. NO formation was further restricted to low pH levels, as an increase in pH led to a sharp decline in NO formation. The addition of Fe$^{3+}$ to the reaction did not enhance NO formation, as NO formation with added Fe$^{3+}$ was similar to the NO formation without Fe$^{3+}$ at a similar pH. The strong pH dependency of the observed NO formation can be explained by the fact that HNO$_2$ rather than NO$_2^-$ is the reactive species in the self-decomposition of NO$_2^-$ which leads to the production of NO (van Cleemput and Samater, 1996). Due to the protonation of NO$_2^-$ to HNO$_2$ (pKa = 3.3) the reaction became very slow at pH 5, as the equilibrium between HNO$_2$ and NO$_2^-$ shifts towards the NO$_2^-$ side. NO$_2^-$ is stable above pH 5.5 (van Cleemput and Baert, 1978), so that an abiotic NO formation in soils seems to be restricted to acidic ecosystems, as e.g., coniferous forests (Stange et al., 2000).

N$_2$O formation, however, could only be observed in the presence of NH$_2$OH. Two N$_2$O production mechanisms could be observed, the reaction between NH$_2$OH and NO$_2^-$ and the oxidation of NH$_2$OH by a transition metal. Like in the abiotic formation of NO, HNO$_2$ was the reactive species in the reaction with NH$_2$OH. The reaction showed the same pH dependency as the self-decomposition of NO$_2^-$, so that this pathway is also limited to acidic soils. As HNO$_2$ is involved in both, abiotic NO and N$_2$O formation, both reactions are competing for HNO$_2$. After an initial peak
in NO formation abiotic N₂O production becomes the dominant pathway, so that the reaction with NH₂OH seems to be the preferred or faster reaction when both precursors are available. An even higher and faster formation of N₂O was observed when Fe³⁺ was added to the NO₂⁻ and NH₂OH reaction. N₂O production from the oxidation of Fe³⁺ could also be observed in the absence NO₂⁻, albeit at a lower level. These results are in conjunction with Minami and Fukushi (1986) and Bremner et al. (1980) who found the oxidation of NH₂OH by transition metals to be faster than the reaction between NH₂OH and NO₂⁻. Also buffers were found to attenuate the reaction with Fe³⁺, the reaction could potentially be of more relevance, as it is possible over a larger pH range than the reaction with NO₂⁻.

Based on the above results, the assumption that reactions involving NH₂OH lead to the formation of N₂O and reactions only involving NO₂⁻ lead to the formation of mainly NO as postulated by Bremner (1997) could be confirmed. Abiotic NO formation from NO₂⁻ self-decomposition as in classical chemodenitrification could potentially play a role in acidic soils, while abiotic N₂O can be hypothesized to be feasible over a wider pH range with NH₂OH as the key component in the formation.

3.4.2. Comparison of reaction mechanisms

The average δ¹⁵Nbulk of N₂O produced in all abiotic reactions studied ranged between −26.9 and 1.4‰ (Table 3.2). It reflected the δ¹⁵N of the substrate partially, hence the information obtained from δ¹⁵Nbulk of emitted N₂O to identify the N substrate is not straightforward, and the use of δ¹⁵Nbulk to identify the N substrate may be limited. Reactions involving NO₂⁻ produced N₂O with the lowest δ¹⁵Nbulk, which could be attributed to the low δ¹⁵N of the substrate, but reactions with NH₂OH as the only N substrate yielded N₂O with a δ¹⁵Nbulk nearly as low. The reason for this is a variable degree of fractionation in favor of the lighter isotope for different reaction pathways, which is mirrored by the NIE covering a range of −0.8 to 21.1‰ and strongly depending on experimental conditions (e.g., buffer medium and pH) for certain pathways (Table 3.2). Correspondingly, fractionation was lowest in the unbuffered experiments but much higher in experiments e.g. with citrate–phosphate buffer, although both were associated with high N₂O production rates.

The decomposition of NH₂OH catalyzed by Cu²⁺ could clearly be distinguished from the pattern of the other reactions, as it showed an inverse fractionation pattern with N₂O getting lighter in ¹⁵N over time. This may be attributed to one of the complex intermediate formation steps, such as fractionation during dimerization in favor of a ¹⁵N–¹⁴N bond as discussed below.

The average δ¹⁸O of N₂O in the different experiments showed a similar order and spanned a similar range as δ¹⁵Nbulk, with average values between 33.9 and 60.2‰ (Table 3.2). The same substrates in buffered or unbuffered medium led to significantly different δ¹⁸O values, whereas the same sub-
strate in the same buffer medium, but at different pH, produced similar δ18O values (experiments 1 and 2). Values were highest for unbuffered experiments, while reactions in citrate–phosphate buffer produced intermediate and those in Tris–maleate buffer the lowest δ18O values. A steady increase in δ18O was observed in all experiments as reactions proceeded, which is consistent with Rayleigh fractionation behavior. The interpretation of δ18O values might be complicated by possible, buffer-dependent exchange of O between water and NO2−, while no exchange is assumed for NH2OH (Kool et al., 2011). However, there was again one exception from this pattern, i.e., the autoxidation of NH2OH catalyzed by Cu2+, which showed high δ18O values and the largest increase in δ18O. Figure 3.7 additionally illustrates the great variability in δ18O, as well as in δ15Nbulk fractionation, between the different experiments, although relationships between the two delta values were found for each individual experiment, which is in agreement with Schmidt et al. (2004a) who found a correlation of both delta values as a function of reaction time or turnover for the NH2OH oxidation with *Nitrosomonas europaea*.

The combined reaction between NH2OH, NO2−, and iron in unbuffered solution (experiments 6 and 8) resulted in similar kinetics as the reaction between NH2OH and Fe3+ in an unbuffered medium, yet the δ15Nbulk of N2O gave no indication about the N substrate and revealed no isotopic fractionation during the experiment. The N turnover in the first 120 min was similar to experiments 1–4, yet the maximum N2O mixing ratio in experiment 6 was at a similar level, although NH2OH concentration was only half as compared to experiments 1–4. This could have had different reasons: two reaction mechanisms occurring simultaneously and leading to higher N2O production than both mechanisms alone, and the experiment being conducted in an unbuffered medium. In buffered solution, the buffer anion had a considerable influence on reactions involving Fe3+, most probably by building iron complexes, which reduced the free iron concentration, thus preventing a reaction with NH2OH.

There was no indication for a different reaction mechanism caused by the addition of Fe2+. In previous studies, the reaction between NH2OH and NO2− had been shown to be second-order (Döring and Gehlen, 1961) and the oxidation of NH2OH by Fe3+ to be pseudo-first-order with respect to NH2OH and Fe3+ concentration (Butler and Gordon, 1986). Most likely, Fe2+ was immediately oxidized to Fe3+, leading to a higher overall N turnover. The proportionally higher Fe3+ concentration as compared to the N substrate in experiment 8 vs. experiment 6 probably enhanced the reaction velocity and could have made up for part of the kinetic effect of the lower N substrate concentration, making the reaction appear first-order.

There were probably two reasons leading to slow kinetics in experiments 7 and 9 in buffered medium as compared to experiments 6 and 8 in unbuffered solution: (a) a higher pH slowing down the reactions of NH2OH with NO2− and/or with Fe3+ (Butler and Gordon, 1986; van Cleemput and Samater, 1996), and (b) buffer anions forming very stable or even insoluble iron complexes, re-
stricting the reactivity of Fe$^{3+}$. Further experiments showed that the oxidation of NH$_2$OH by Fe$^{3+}$ is suppressed in buffered solution independent of pH (data not shown). N$_2$O production in experiments 7 and 9 seemed to be determined by the slow degree of protonation of NO$_2^-$ – the process that forms HNO$_2$ (pK$_a$ = 3.3) – at pH 5.5, as not NO$_2^-$, but HNO$_2$ is the reactant in the reaction of NH$_2$OH and NO$_2^-$. 

### 3.4.3. Comparison with other studies

The observed $^{15}$N site preference of N$_2$O formed in the abiotic reactions of this study was in a similar range to recent studies reporting on N$_2$O production via microbial NH$_2$OH oxidation. In batch pure culture studies, SP values of 33.5 ± 1.2‰, 32.5 ± 0.6‰, 35.6 ± 1.4‰, and 30.8 ± 5.9‰ had been observed for *Nitrosomonas europaea*, *Nitrosomonas multiformis*, *Methylosinus trichosporium*, and *Methylococcus capsulatus*, respectively (Sutka et al., 2006; Sutka et al., 2003, 2004). For marine environments (*Nitrosomonas marina* C-113a) a SP of 36.3 ± 2.4‰ was found (Frame and Casciotti, 2010). Recently, Santoro et al. (2011) reported a SP of N$_2$O from marine AOA that was in the same range as for abiotically produced N$_2$O (30.3‰). In a recent mixed culture experiment the SP of N$_2$O ranged from 26.4 to 30.7‰ at conditions where NH$_2$OH oxidation was expected to be the dominant N$_2$O producing process (Wunderlin et al., 2013). However, in mixed population systems additional pathways cannot be completely excluded and may have led to a lower SP (Wunderlin et al., 2013). The SP of N$_2$O produced abiotically via NH$_2$OH oxidation had been determined for the first time by Toyoda et al. (2005). In a laboratory experiment with addition of MnO$_2$ to a NH$_2$OH solution, the authors found a SP of 29.5 ± 1.1‰. Wunderlin et al. (2013) observed a similar SP by adding NH$_2$OH to tap water (SP 30.3 ± 0.2‰) in a batch reactor. Both studies confirm a constant SP for N$_2$O produced by abiotic reactions as found in this study, although their results are about 4–5‰ lower than the average SP obtained in this study. Additionally, this study showed for the first time that the SP was constant during all reaction stages for different reaction mechanisms and process conditions.

### 3.4.4. Mechanism leading to high positive SP

In the present study, a characteristic, strongly positive SP was observed for N$_2$O formation via NH$_2$OH oxidation. The exact reaction mechanism is unknown, but it is hypothesized that the high positive SP is indicative of a mechanism via a symmetric intermediate, presumably cis-hyponitrous acid (HO–$^{14}$N=15N–OH) or rather its anion cis-hyponitrite (–O–$^{14}$N=15N–O–), with a kinetic isotope effect leading to a preferential cleavage of the $^{14}$N–O bond over the $^{15}$N–O bond (Schmidt et al., 2004a; Toyoda et al., 2002). However, the detailed mechanism is poorly understood, mainly because of the rapid sequential isomerization steps and proton transfer during the reaction of NH$_2$OH.
Site-specific $^{15}$N isotopic signatures of abiotically produced N$_2$O (Fehling and Friedrichs, 2011). Nitrooxyl (HNO) has also been discussed as an important initial intermediate (Bonner and Hughes, 1988). In a recent modeling study, Fehling and Friedrichs (2011) found N$_2$O production to be initialized by formation of HNO, followed by a fast dimerization leading to hyponitrous acid and, after rapid deprotonation, finally to the decomposition of the hyponitrite anion. The fast reaction makes N$_2$O the only measurable indicator. Our findings suggest that all NH$_2$OH oxidation processes have a common intermediate in which the N–O bond cleavage is occurring. It also suggests that the SP of N$_2$O is an indicator of a formation mechanism via a symmetric intermediate but not necessarily of the originating process.

### 3.4.5. Source partitioning

The $\delta^{15}$N$_{\text{bulk}}$ in N$_2$O emitted from soils is supposed to be determined by the isotopic signature of the substrate, and not by the production process (Sutka et al., 2006). This prevents the use of $\delta^{15}$N$_{\text{bulk}}$ alone in N$_2$O source partitioning, as different processes in soils can feed on the same substrate pools. Our study demonstrated that $\delta^{15}$N$_{\text{bulk}}$ was only of very limited use for identifying the N substrate of N$_2$O production, even though it was a laboratory study under controlled conditions. In field studies, where the isotopic signature of the substrate is often unknown and different intermediate N transformation steps affect the final isotopic signature of emitted N$_2$O, it will become impossible to disentangle different N$_2$O production pathways using only $\delta^{15}$N$_{\text{bulk}}$. Our results also showed that the $\Delta \delta^{15}$N was not constant for the different abiotic reactions studied and was influenced by reaction conditions and perhaps other unknown factors.

The $^{18}$O signature of N$_2$O has been increasingly used to characterize its production processes in soils (Wrage et al., 2005). However, it has been shown that up to 100% of the O in N$_2$O can originate from H$_2$O via O exchange mainly between H$_2$O and NO$_2^-$, but potentially also between other intermediates of the diverse N transformation and N$_2$O production pathways (Kool et al., 2009a; Kool et al., 2009b). This entails that the $\delta^{18}$O in N$_2$O reflects the isotopic signature of the substrate O only partially or not at all. With our first continuous measurement of $\delta^{18}$O in N$_2$O, we could confirm the great variability of $\delta^{18}$O for abiotic N$_2$O production. A positive correlation between $\delta^{18}$O and SP, as found by Frame and Casciotti (2010) for N$_2$O production via NH$_2$OH oxidation, could not be confirmed by our experiments. However, we found a strongly positive correlation between $\delta^{18}$O and $\delta^{15}$N$_{\text{bulk}}$ of N$_2$O for experiments with a $^{15}$N-depletion over time, and a strongly negative correlation for experiments with an inverse $^{15}$N fractionation ($\alpha < 0.05$). Thus, the O exchange during N$_2$O production in the terrestrial N cycle poses a great challenge for the interpretation of O isotopes in N$_2$O and their application to biogeochemical studies (Kool et al., 2009b).

Lately, the N$_2$O SP has been given the greatest potential in overcoming the influence of substrate, variable fractionation and O exchange on isotopic signatures of N$_2$O. The SP is independent of the
isotopic signature of the substrate and does not change significantly during production (Ostrom and Ostrom, 2011). Thus, the SP provides a conservative tracer of the N\textsubscript{2}O production process (Opdyke et al., 2009; Toyoda et al., 2005). The constant SP for the abiotic reactions observed in our study were in the same range as reported for microbial NH\textsubscript{2}OH oxidation, fungal denitrification, and AOA (Santoro et al., 2011; Sutka et al., 2008). In contrast, N\textsubscript{2}O derived from heterotrophic and nitrifier denitrification can be clearly distinguished from other sources. All other known N\textsubscript{2}O production processes are characterized by similar SP. Our results rather demonstrated that the SP does not necessarily reflect the production process per se, but the crucial intermediate which undergoes N–O bond cleavage.

Other pathways like DNRA and anammox can add to soil N\textsubscript{2}O production. DNRA can produce N\textsubscript{2}O as a side product (Stevens et al., 1998), whereas anammox is thought to bypass the potential N\textsubscript{2}O production in aquatic systems (Yang et al., 2012). However, NH\textsubscript{2}OH as a possible intermediate of anammox might lead to the formation of small amounts of N\textsubscript{2}O (van der Star et al., 2008). It can be assumed that N\textsubscript{2}O produced via these pathways has a similarly high SP as N\textsubscript{2}O from the other pathways with NH\textsubscript{2}OH as the essential intermediate.

3.5. Conclusions

In the present study we could show that the application of quantum cascade laser absorption spectroscopy for monitoring single reactions is a promising and adequate tool for getting deeper insight in the N cycle, as biogeochemical reactions and characteristics like inverse fractionation can be followed in real time. The \textsuperscript{15}N SP of N\textsubscript{2}O from purely abiotic reactions involving the highly reactive nitrification intermediate NH\textsubscript{2}OH has been determined for the first time with both high precision and high temporal resolution for different abiotic reaction mechanisms over a wide pH range. Although three different reaction pathways from NH\textsubscript{2}OH to N\textsubscript{2}O over a wide range of pH values were tested, the SP remained constant within the experimental uncertainty in all reactions, with average SP values ranging only from 33.9 to 35.6‰. Thus, no evidence for an effect of different experimental conditions like pH or buffer medium on SP was found.

This laboratory study puts emphasis on coupled biotic–abiotic reactions in soils and adds a new perspective to N\textsubscript{2}O production during nitrification. In most of the reactions studied, conversion of NH\textsubscript{2}OH to N\textsubscript{2}O was fast as expected due to the very high reactivity of NH\textsubscript{2}OH. This implies that NH\textsubscript{2}OH could also be quickly converted to N\textsubscript{2}O upon release to the soil matrix by ammonium-oxidizing organisms. In addition, this study provides new information, which might be helpful for the source partitioning of N\textsubscript{2}O emissions from soils. The SP of N\textsubscript{2}O is deemed to have the potential to be a powerful tool in disentangling the different N\textsubscript{2}O production and consumption processes in
soils, but in the present study we also showed its limitations. It seems that SP can only be used to differentiate between oxidative and reductive N₂O production, as obviously all processes relevant for N₂O formation during nitrification lead to the same SP. It can be argued that (1) microbial and abiotic processes share the same underlying intermediate steps leading to the same SP, or (2) the observed N₂O formation during nitrification is not of microbial origin but a purely chemical reaction of the nitrification intermediate NH₂OH.
Chapter 4

Abiotic N₂O production from hydroxylamine in soils and their dependence on soil properties

Based on:
4.1. Introduction

$\text{N}_2\text{O}$ is an important greenhouse gas. It has an about 300 times higher global warming potential than $\text{CO}_2$ over a time frame of 100 years and contributes approximately 6% to anthropogenic radiative forcing, making it the third-most important contributor after $\text{CO}_2$ and methane (WMO, 2013). Furthermore, $\text{N}_2\text{O}$ is known to be partly responsible for the catalytic destruction of ozone in the stratosphere (Crutzen, 1970). While other historically dominant ozone depleting substances have been successfully regulated by the Montreal Protocol, $\text{N}_2\text{O}$ is still unregulated and, if present trends continue, will become the dominant ozone depleting substance in the 21st century (Ravishankara et al., 2009). The atmospheric mixing ratio of $\text{N}_2\text{O}$ has increased by 20% from a pre-industrial level of 270 ppb to 325 ppb in 2012 at a rate of 0.80 ppb yr$^{-1}$ over the last decade (WMO, 2013). The increase in atmospheric $\text{N}_2\text{O}$ is tightly coupled to increasing anthropogenic $\text{N}$ fixation, mainly applied as fertilizer and manure on agricultural fields.

Soils have been identified as the major source of $\text{N}_2\text{O}$, contributing an estimated 50–60% to global $\text{N}_2\text{O}$ emissions (USEPA, 2010). However, there is still a large uncertainty associated with estimates of global $\text{N}_2\text{O}$ emissions from natural and anthropogenic sources, ranging from 8.1 to 30.7 Tg N yr$^{-1}$ (Ciais et al., 2013). This great range of estimated values is mainly a reflection of the great uncertainty of the individual source and sink strengths of the diverse processes involved in $\text{N}_2\text{O}$ formation and consumption in soils (Billings, 2008).

Two microbial $\text{N}$ transformation processes, autotrophic nitrification and heterotrophic denitrification, are considered the major $\text{N}_2\text{O}$ sources, contributing an estimated 70% of the global $\text{N}_2\text{O}$ emissions from soils (Butterbach-Bahl et al., 2013). However, as discussed in great detail above, there is a lot of uncertainty about the sole production of $\text{N}_2\text{O}$ by microbial processes, as several abiotic production mechanisms involving the nitrification intermediate $\text{NH}_2\text{OH}$ have been shown to be feasible in soils.

Lately, stable isotope techniques have developed great potential for disentangling the variety of different $\text{N}_2\text{O}$ formation processes; especially the intramolecular distribution of $^{15}\text{N}$ in $\text{N}_2\text{O}$, the so-called site preference (SP), has been in the focus of recent research (Decock and Six, 2013). The site-specific isotopic signature of $\text{N}_2\text{O}$ produced by several microbial pathways has been studied (Frame and Casciotti, 2010; Opdyke et al., 2009; Sutka et al., 2008; Sutka et al., 2006; Well et al., 2006; Wunderlin et al., 2013) as well as for abiotic $\text{N}_2\text{O}$ production via $\text{NH}_2\text{OH}$ oxidation (Heil et al., 2014). However, until now it is impossible to unambiguously differentiate between $\text{N}_2\text{O}$ production and consumption processes using SP information (Ostrom and Ostrom, 2011).

For better $\text{N}_2\text{O}$ mitigation strategies it is vital to understand the multitude of underlying microbial and abiotic processes of $\text{N}_2\text{O}$ production in the terrestrial $\text{N}$ cycle and their controlling factors, as it is likely that $\text{N}_2\text{O}$ emissions from soils will increase at an ever growing rate due to an increasing
demand for food, accompanied by an increased use of N fertilizer (Ciais et al., 2013). A better understanding is also prerequisite for lowering the high model uncertainty related to N₂O emissions that is caused by the multitude of simultaneous processes involved in N₂O formation, but also by the high temporal and spatial variability of these processes.

The chemical oxidation of NH₂OH in the presence of several transitions metals commonly found in soils was recognized more than 30 years ago (Bremner et al., 1980), but is still neglected in most current N trace gas studies. The present study was designed to test for the potential of a coupled biotic–abiotic mechanism of N₂O production under aerobic conditions, in which NH₂OH microbiologically produced during nitrification is leaking to a certain extent out of autotrophic and heterotrophic nitrifiers into the soil matrix, where it is readily oxidized to N₂O by transition metals, such as manganese or iron, or by NO₂⁻, which is also excreted by ammonium-oxidizers. To test for this potential mechanism, we added NH₂OH to soil samples from different ecosystems (forest, grassland, cropland), both under non-sterile conditions and after sterilization with three different sterilization methods. The guiding hypothesis of the study was that at least in some soils this coupled biotic–abiotic mechanism might play a significant role in aerobic N₂O formation during nitrification.

4.2. Material and methods

4.2.1. Sample collection

Soil samples were collected from three field sites (cropland, grassland, coniferous forest) that are part of the TERENO network, and additionally from a deciduous forest on the campus of Forschungszentrum Jülich (50°54'38"N, 6°24'44"E). The coniferous forest site (Wüstebach; 50°30'15"N, 6°18'15"E) was situated in the low mountain ranges of the Eifel National Park. The main vegetation at this site is Norway spruce (*Picea abies* (L.) Karst.). The soil type was a Cambisol with a loamy silt texture. The grassland site (Rollesbroich; 50°37'18"N, 6°18'15"E) was located in the Northern Eifel region. The soil type was also Cambisol. The agricultural site (Selhausen; 50°52'09"N, 6°27'01"E) was intensively used with regular lime and fertilizer applications. The soil type was classified as a Luvisol with a silty loam texture. At each site, soil samples were collected from the top 20 cm. At the coniferous forest site, the top 20 cm were divided into litter layer (L), organic topsoil horizon (Oh), and humic mineral topsoil layer (Ah) and collected separately. After collection, samples were transferred to the institute, where they were sieved to 2 mm and stored at 4 °C under well aerated conditions for further analysis.
4.2.2. Soil chemical analyses

Soil samples were analyzed for chemical parameters by the central analytical laboratory of Forschungszentrum Jülich. The total C and total N content were determined using an elemental analyzer (vario EL Cube, Elementar Analysensysteme, Hanau, Germany). For measurements, 20–50 mg sample material, in replicates of three, were analyzed. Concentrations of a range of elements were determined by ICP-OES. Sample extraction was done using lithium borate by extracting the mixture at 1000 °C for 30 min in a muffle furnace. The melt was dissolved in 30 mL HCl (5%) and filled up to a volume of 50 mL before analysis for total Ca, Cu, Fe, K, Mg, Mn, Na, and Sn content. Soil pH was determined in 0.01 M CaCl₂ solution. An overview of all soil chemical parameters determined can be found in Table 4.1.

4.2.3. Incubation experiments

To study the formation of N₂O in the different soils in dependence of NH₂OH content, incubation experiments were conducted in the laboratory. One gram (dry weight) of soil (0.5 g for litter) were weighed into 22-mL GC headspace vials (VWR International, Darmstadt, Germany), and the water content of the soil samples was adjusted to 50% water holding capacity (WHC) with deionized water or NH₂OH solution, respectively. Vials were closed gastight immediately afterwards with butyl septa and aluminum crimp caps (VWR International). Each treatment was carried out in replicates of three. The standard application rate of NH₂OH was 5 nmol (5 nmol per g soil), but experiments with a higher application rate of 10 nmol (10 nmol per g soil) were also conducted. Incubation experiments were performed with non-sterile as well as sterile soils. We used three different methods of sterilization: autoclaving, chloroform fumigation, and methyl iodide fumigation. Through autoclaving, samples were sterilized by exposing them to a temperature of 121°C and a pressure of 0.3 MPa for 30 min. For sterilization with chloroform fumigation, soil samples were placed into an 18.5 L desiccator together with 100 mL of chloroform (Merck, Darmstadt, Germany) in a separate beaker. The desiccator was closed and evacuated, and the soil samples were exposed to chloroform at room temperature for 24 h. Methyl iodide fumigation was conducted in an analogous manner, only exchanging chloroform with 5 mL methyl iodide (99% reagent plus grade, Sigma-Aldrich, Steinheim, Germany) as described in Oswald et al. (2013). Incubations were performed at the same day sterilization procedures were finished.
Table 4.1: Overview of the different soil samples and their chemical parameters that were used in this study.

<table>
<thead>
<tr>
<th>Site</th>
<th>pH</th>
<th>C</th>
<th>N</th>
<th>C/N</th>
<th>Ca</th>
<th>K</th>
<th>Mg</th>
<th>Na</th>
<th>Fe</th>
<th>Mn</th>
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<td>[−]</td>
<td>[mass-%]</td>
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<td></td>
<td></td>
<td></td>
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</tr>
<tr>
<td>Litter</td>
<td>3.4</td>
<td>45.70</td>
<td>2.02</td>
<td>22.60</td>
<td>0.29</td>
<td>0.21</td>
<td>0.09</td>
<td>0.03</td>
<td>0.52</td>
<td>0.02</td>
</tr>
<tr>
<td>Oh</td>
<td>2.9</td>
<td>29.30</td>
<td>1.43</td>
<td>20.50</td>
<td>0.11</td>
<td>0.73</td>
<td>0.13</td>
<td>0.10</td>
<td>2.05</td>
<td>0.00</td>
</tr>
<tr>
<td>Ah</td>
<td>3.1</td>
<td>14.10</td>
<td>0.72</td>
<td>19.50</td>
<td>0.05</td>
<td>1.15</td>
<td>0.17</td>
<td>0.15</td>
<td>3.34</td>
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<tr>
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<td></td>
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<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>grassland</td>
<td>3.6</td>
<td>12.60</td>
<td>0.63</td>
<td>19.90</td>
<td>0.27</td>
<td>1.00</td>
<td>0.16</td>
<td>0.43</td>
<td>1.23</td>
<td>0.08</td>
</tr>
<tr>
<td>cropland</td>
<td>4.9</td>
<td>2.88</td>
<td>0.27</td>
<td>10.50</td>
<td>0.19</td>
<td>1.63</td>
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<td>0.32</td>
<td>2.90</td>
<td>0.11</td>
</tr>
<tr>
<td></td>
<td>6.4</td>
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<td>0.37</td>
<td>1.46</td>
<td>0.33</td>
<td>0.57</td>
<td>2.25</td>
<td>0.09</td>
</tr>
</tbody>
</table>
Experiments were conducted with field moist as well as with pre-dried samples. The pre-dried samples were dried slowly at 60 °C until the weight was constant. The incubation time in all experiments was 6 h at room temperature (20 °C). At the end of the incubation time, the headspace of the sample vials was analyzed using a gas chromatograph (Clarus 580, PerkinElmer, Rodgau, Germany) equipped with an electron capture detector (ECD) for N₂O detection. The instrument was calibrated using three different standard gases with 250, 500, and 750 ppb N₂O balanced with N₂ (99.5% purity, Linde, Munich, Germany).

4.2.4. Reaction kinetics analyses

Experiments to determine the reaction kinetics of NH₂OH oxidation to N₂O were conducted for the different soils with a flow through-reaction chamber connected to an infrared laser absorption spectrometer for online real-time analysis of N₂O mixing ratio (Dual Laser Quantum Cascade Trace Gas Monitor, Aerodyne Research, Inc., Billerica, MA, USA). The instrument consists of two mid-infrared lasers and is able to measure N₂O, CO₂, CH₄, H₂O simultaneously at a temporal resolution of up to 10 Hz. Water vapor is measured to correct for volumetric dilution and pressure broadening effects caused by increasing water vapor concentration in the sample air. Prior to every experiment the cell was flushed with synthetic air and a new background spectrum was taken. The instrumental precision of the measurements given as standard deviation of the average N₂O mixing ratio at atmospheric level was <0.3 ppb.

The first experiment was carried out with non-sterile as well as autoclaved and chloroform-fumigated soil samples. For this purpose, 10 g of soil (dry weight) were weighed into a 250-mL beaker, so that the bottom of the beaker was completely covered. The soil water content was adjusted to 50% WHC with NH₂OH solution. Depending on the initial moisture level of the soil samples, different amounts of solution had to be added, but NH₂OH concentration of the solutions was individually adapted, so that the same amount of 5 µmol NH₂OH (500 nmol per g soil) was added to each sample. A hundredfold higher amount of NH₂OH per g soil compared to the incubation experiments had to be chosen for sufficiently high N₂O formation to achieve a high measurement precision at the high air flow rate through the incubation chamber. Immediately after addition of the solution to the soil sample, the beaker was placed in the flow-through chamber, which was instantly closed. The chambers were custom-made of a quartz glass cylinders fitted with PTFE cover plates. The PTFE plates were hold together by seven aluminum rods at the outside of the chamber. The contact surfaces were sealed with Viton® O-rings. A fan inside the chamber assured uniform mixing of the gas phase. The chamber volume was 2 L and it was purged with 2.5 L min⁻¹ of pressurized air controlled by a mass flow controller (Brooks Instruments, Dresden, Germany) during the experiment. The outlet of the chamber was directly connected to the laser analyzer via PTFE tubing.
A second experiment was conducted to quantify N\textsubscript{2}O formation from NH\textsubscript{2}OH in soil at different temperatures. A refrigerating and heating water circulator (RC 20 CS, LAUDA, Lauda-Königshofen, Germany) was used to maintain constant temperatures. In this experiment, 25 g of autoclaved cropland soil (dry weight) were transferred to an autoclaved 500-mL bottle (SCHOTT DURAN®, DURAN Group GmbH, Wertheim, Germany). The bottle was then placed in the water bath, so that only the bottleneck was above the water surface. To monitor soil temperature, a mercury thermometer was pushed into the soil that had been sterilized with ethanol immediately before use. Then, like in the first experiment, NH\textsubscript{2}OH solution was added to adjust soil water content to 50% WHC, corresponding to a NH\textsubscript{2}OH amount of 5 µmol (200 nmol per g soil). The bottle was closed gastight with custom-made lids with gas inlet and outlet ports immediately afterwards and flushed with 2.5 L min\textsuperscript{-1} pressurized air as described above.

4.2.5. N\textsubscript{2}O isotopic analyses

For the determination of N\textsubscript{2}O isotopic composition and \textsuperscript{15}N isotopomer signatures, 5 g of soil (2.5 g for litter) were weighed into 100 mL headspace vials (VWR International) and treated the same way as samples for GC incubation experiments. According to the five times larger headspace in the IRMS vials, we used five times more sample material and added a five times higher amount of NH\textsubscript{2}OH to the soil (25 nmol; 5 nmol per g soil). Samples were incubated under the same conditions as for the GC incubation. After 6 h incubation time, the headspace of the vials was analyzed using an isotope ratio mass spectrometer (IRMS) (IsoPrime 100, Elementar Analysensysteme, Hanau, Germany) coupled to a pre-concentration unit (TraceGas, Elementar Analysensysteme) for online separation and purification of N\textsubscript{2}O.

With the IRMS, the mass-to-charge ratios (m/z) 44, 45, and 46 of the N\textsubscript{2}O\textsuperscript{+} ion and m/z 30 and 31 of the NO\textsuperscript{+} fragment ion of N\textsubscript{2}O were measured and used to determine the isotopologue and isotopomer signatures of N\textsubscript{2}O (Toyoda and Yoshida, 1999). More specifically, \(\delta^{15}\text{N}_{\text{bulk}}\) (i.e., the average \(\delta^{15}\text{N}\) over the N\textsubscript{2}O molecule), \(\delta^{15}\text{N}_a\) (i.e., \(\delta^{15}\text{N}\) at the central position of the N\textsubscript{2}O molecule), and \(\delta^{18}\text{O}\) of N\textsubscript{2}O were determined. The \(\delta^{15}\text{N}\) at the terminal position of the N\textsubscript{2}O molecule, \(\delta^{15}\text{N}_\beta\), was calculated according to \(\delta^{15}\text{N}_\beta = 2 \cdot \delta^{15}\text{N}_a - \delta^{15}\text{N}_\alpha\). The \(\delta^{15}\text{N}\) SP is then defined as SP = \(\delta^{15}\text{N}_a - \delta^{15}\text{N}_\beta\). A correction for \(\delta^{17}\text{O}\) was performed, assuming a mass-dependent fractionation of \(\delta^{17}\text{O}\) and \(\delta^{18}\text{O}\) and using the calculations according to Kaiser et al. (2003), with \(17\text{R} = 0.00937035 \cdot (18\text{R})^{0.516}\). We used pure N\textsubscript{2}O (99.999%, Linde, Munich, Germany) as working standard for isotope analysis, and \(\delta^{15}\text{N}_{\text{bulk}}\), \(\delta^{18}\text{O}\), and SP were calibrated against two reference gases (Ref 1: \(\delta^{15}\text{N}_a\): 15.70 ± 0.31‰, \(\delta^{15}\text{N}_\beta\): –3.21 ± 0.37‰, \(\delta^{15}\text{N}_{\text{bulk}}\): 6.24 ± 0.11‰, SP: 18.92 ± 0.66‰, \(\delta^{18}\text{O}\): 35.16 ± 0.35‰; Ref 2: \(\delta^{15}\text{N}_a\): 5.55 ± 0.21‰, \(\delta^{15}\text{N}_\beta\): –12.87 ± 0.32‰, \(\delta^{15}\text{N}_{\text{bulk}}\): –3.66 ± 0.13‰, SP: 18.42 ± 0.50‰, \(\delta^{18}\text{O}\): 32.73 ± 0.21‰) provided by EMPA (Dübendorf, Switzerland) and as described in Mohn et
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al. (2014). Analytical precision, expressed as standard deviation, was 0.1‰, 0.2‰, and 0.2‰ for $\delta^{15}$N, $\delta^{18}$O, and SP, respectively.

Isotope values are reported in the delta notation, with $\delta = (R_{\text{sample}}/R_{\text{standard}} - 1) \times 1000$, where $R_{\text{sample}}$ and $R_{\text{standard}}$ are the ratio of heavy to light isotope ($^{15}$N/$^{14}$N or $^{18}$O/$^{16}$O) in the sample and an international standard, i.e., atmospheric N$_2$ for nitrogen and Vienna Standard Mean Ocean Water (VSMOW) for oxygen, respectively.

4.2.6. Calculations

N$_2$O emission rates following NH$_2$OH addition were calculated by subtracting N$_2$O emission rates from corresponding control samples. The mass of N$_2$O-N produced was calculated using the ideal gas law at standard laboratory conditions ($p = 1013$ mbar; $T = 293.15$ K).

To interpret isotopic data of N$_2$O following NH$_2$OH addition, a simple two-source mixing model was used. From the mass-weighted $\delta$-value of the sample (N$_2$O with NH$_2$OH addition), the mass-weighted $\delta$-value of the background (N$_2$O with H$_2$O addition only) was subtracted and divided by the sum of the two masses: $\delta_{\text{sample}} = (\delta_{\text{NH}_2\text{OH}}m_{\text{NH}_2\text{OH}} - \delta_{\text{H}_2\text{O}}m_{\text{H}_2\text{O}})/(m_{\text{NH}_2\text{OH}} + m_{\text{H}_2\text{O}})$.

All statistical analyses were performed using OriginPro 8 (OriginLab Corp., Northampton, MA, USA). An unpaired t-test was used to test for treatment differences at a significance level $\alpha < 0.05$. The uncertainty of the provided data is given as the standard deviation of the replicates. For calculated values, uncertainties of all individual measured parameters propagate to the error of the derived value on the basis of the standard deviation.

4.3. Results

4.3.1. Soil incubation experiments

While neither the L nor the Oh horizon of the spruce forest soil showed significantly higher N$_2$O evolution upon the addition of NH$_2$OH compared to the H$_2$O only treatment, the Ah horizon exhibited N$_2$O formation only after NH$_2$OH addition, while in the H$_2$O treatment there was no N$_2$O formed at all (Fig. 4.1.). In contrast, N$_2$O formation in the deciduous forest soil could be observed for both treatments, but significantly more with NH$_2$OH than with H$_2$O only. The strongest reaction to NH$_2$OH addition was found in the grassland and cropland soils, whereas there was no N$_2$O formation after H$_2$O addition. The turnover of NH$_2$OH-N to N$_2$O-N during the incubation time of 6 h was 44% for the grassland, and 56% for the cropland soil.
Abiotic N$_2$O production from hydroxylamine in soils

**Figure 4.1:** N$_2$O emission rates in ng N per g soil from six different non-sterile soil samples after the addition of deionized H$_2$O or NH$_2$OH solution (5 nmol) in 6-h incubation experiments. Samples of ambient air were measured and subtracted as backgrounds from the results. Error bars represent the SD of three replicates. Lower case letters indicate significant differences between different treatments at one site, capital letters between the different sites within one treatment ($\alpha < 0.05$).

In a second series of incubations under the same conditions we used different sterilization methods to test the hypothesis of an abiotic production mechanism. N$_2$O formation after NH$_2$OH addition differed between sterile and non-sterile soil samples, and in addition also between sterilization methods (Fig. 4.2). Autoclaving had the strongest effect on NH$_2$OH-induced N$_2$O emissions in all samples. In the coniferous forest soil samples it reduced N$_2$O emission rates to zero in all three soil horizons. Also in the grassland soil a strong attenuation effect was observed, with N$_2$O emission rates after NH$_2$OH addition being reduced by almost 99%. The only soil showing significant N$_2$O emissions upon NH$_2$OH addition after autoclaving was the cropland soil, for which the amount of N$_2$O formed was only reduced by approx. 50%. Chloroform fumigation did not have such a pronounced effect on N$_2$O emission rates as autoclaving. It did not have a significant effect in the L and Oh horizon of the coniferous forest soil, led to a decrease of about 20% in both the grassland and cropland soil, and lowered N$_2$O formation by about 50% both in the Ah layer of the coniferous forest and in the deciduous forest soil. The third sterilization method, the fumigation of samples with methyl iodide, led to similar results as chloroform fumigation (Fig. 4.2).
Abiotic N\textsubscript{2}O production from hydroxylamine in soils

Figure 4.2: N\textsubscript{2}O emission rates in ng N per g soil from six different non-sterile and sterilized soil samples after the addition of NH\textsubscript{2}OH solution (5 nmol) in 6-h incubation experiments. Replicates with the addition of deionized H\textsubscript{2}O were used as backgrounds and subtracted from the results. Error bars represent the SD of three replicates. Lower case letters indicate significant differences between different treatments at one site, capital letters between different sites within one treatment (\( \alpha < 0.05 \)).

Drying of the soil prior to the incubation led to lower NH\textsubscript{2}OH-induced N\textsubscript{2}O emission rates (Fig. 4.3). The largest decrease could be observed for the grassland and deciduous forest site, while for the cropland soil it was only marginal compared to fresh soil. However, also the reduction of N\textsubscript{2}O emission rates by autoclaving was lower when soil samples were dried before incubation. Both non-sterile and sterile cropland soil samples had NH\textsubscript{2}OH-to-N\textsubscript{2}O turnover rates higher than 50%. A two-fold increase in NH\textsubscript{2}OH addition (10 nmol) led to an increase in N\textsubscript{2}O emission rates in all soils (Fig. 4.3). The three coniferous forest soil horizons showed a slight increase after adding twice the amount of NH\textsubscript{2}OH, but still remained at a very low level. The other three non-sterile soils reacted to doubling of added NH\textsubscript{2}OH amount with an increase in N\textsubscript{2}O emission rates by factors of 2.4, 2.1, and 2.9, respectively. In sterilized cropland samples about 49% of the added 10 nmol NH\textsubscript{2}OH-N had been converted to N\textsubscript{2}O-N. This was again by far the highest value of all soils of this study (Fig. 4.3).
**Figure 4.3**: N$_2$O emission rates in ng N per g soil from six different non-sterile and sterilized soil samples after the addition of two different amounts of NH$_2$OH (5 and 10 nmol) in 6-h incubation experiments. Replicates with the addition of deionized H$_2$O were used as backgrounds and subtracted from the results. Error bars represent the SD of three replicates. Lower case letters indicate significant differences between different treatments at one site, capital letters between the different sites within one treatment ($\alpha < 0.05$).

### 4.3.2. Kinetics of N$_2$O formation

Infrared laser absorption measurements at a temporal resolution of 1 Hz provided insight into the kinetics of the oxidation of NH$_2$OH in the different soils of this study. The course of the reaction was very similar for all soils: N$_2$O formation peaked only seconds after the addition of NH$_2$OH, followed by a rapid decline (Fig. 4.4). Already 30 min after NH$_2$OH addition N$_2$O mixing ratios were back to baseline levels in most samples. However, the amplitude of the N$_2$O peaks was significantly different between different soil samples. The differences in peak heights were similar to the differences in N$_2$O formation in the GC incubation experiments. Highest N$_2$O production was found in the non-sterile cropland soil, where chloroform fumigation decreased N$_2$O formation only slightly, while autoclaving reduced the N$_2$O by more than half (Fig. 4.4 A). In all soil samples and treatments, N$_2$O emissions dropped quickly, and most of the reaction was completed in less than 5 min. At this point in time, already 48.5% of the NH$_2$OH–N added to the non-sterile cropland soil had been converted to N$_2$O–N. This rate reached 57.1% after 15 min and 58.5% after 30 min. In contrast, autoclaving reduced the turnover rate to 31.5%.
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Figure 4.4: N2O mixing ratios in ppb measured at a temporal resolution of 1 Hz emitted from different soils after the addition of NH2OH solution (5 µmol): (A) from non-sterile and sterile cropland soil, (B) from non-sterile and sterile grassland soil, (C) from the three non-sterile coniferous forest soil layers, and (D) non-sterile deciduous forest soil.

The addition of the same amount of NH2OH to the grassland soil led to a maximum peak of N2O that was three times lower than for the cropland soil (Fig. 4.4 B). The total N turnover from NH2OH to N2O after 30 min was also lower (31.9%), owing to the lower reaction velocity, which was also reflected in a longer tailing of N2O mixing ratio. No significant difference was observed between non-sterile and chloroform-fumigated soil samples, whereas autoclaving reduced N2O formation significantly, with only 9.3% of the added NH2OH–N converted to N2O–N.

The different layers of the coniferous forest soil showed a NH2OH-induced N2O emission that was two orders of magnitude lower than in the cropland soil, with a small N2O peak within the first 5 min of the experiment (Fig. 4.4 C). A similar, only slightly higher, N2O formation was observed for the deciduous forest soil (Fig. 4.4 D), but the reaction progress was clearly different compared to the coniferous soil, with a slower decrease and longer tailing of N2O mixing ratio.

To ultimately test whether N2O formation from NH2OH occurred via chemical and not a biological reaction, a last experiment was conducted with the laser spectrometer to study the effect of different temperatures on NH2OH oxidation in soils. For this experiment we chose autoclaved, sterile cropland soil only. The experiment revealed that increasing temperature from 10 °C to 50 °C was associated with a steady increase in the initial N2O peak height and faster reaction kinetics, i.e., a faster decline of N2O mixing ratios back to the background level (Fig. 4.5). However, despite dif-
ferent kinetics there was no significant differences in turnover of added NH$_2$OH–N to N$_2$O–N between the different temperatures from the cropland soil, which amounted to about 25% at all temperature levels after one hour. Furthermore, the findings demonstrated that the oxidation of NH$_2$OH proceeded extremely fast and that no further oxidation of the added NH$_2$OH to N$_2$O could be observed after less than one hour.

![Figure 4.5](image)

Figure 4.5: N$_2$O mixing ratios in ppb measured at a temporal resolution of 1 Hz emitted from sterile (autoclaved) cropland soil after the addition of NH$_2$OH solution (5 µmol) at different temperatures from 10 to 50 °C.

4.3.3. Isotopic signature of produced N$_2$O

The isotopic signature of N$_2$O emissions from the L and Oh horizon of the coniferous forest site could not be determined because N$_2$O production was too low. The $\delta^{15}$N$_{bulk}$ values of N$_2$O from Ah horizon of the coniferous soil as well as of the grassland and cropland soils were in the range of –5 to –8‰, i.e., $^{15}$N-depleted compared to the NH$_2$OH substrate, which had a $\delta^{15}$N of $-1.93 \pm 0.11$‰ (Table 4.2). In contrast, $\delta^{15}$N$_{bulk}$ of N$_2$O from the deciduous forest soil was slightly $^{15}$N-enriched over the $\delta^{15}$N of NH$_2$OH, but this value was also associated with a much higher uncertainty due to the low amount of N$_2$O formed. The $\delta^{18}$O values spread out over a larger range than $^{15}$N values, did not reveal a clear pattern, and showed a much higher variability among replicates than $\delta^{15}$N values. The $^{15}$N SP for the three soils with highest amount of N$_2$O formed, i.e., grassland, non-sterile and autoclaved cropland was on average 35‰. The SP values of the other two soils were 2–3‰ lower, but also afflicted with a much higher uncertainty, which was negatively correlated with the amount of N$_2$O produced.
**Table 4.2**: Turnover rate of added NH₃OH-N to N₂O-N, isotopic signatures and ¹⁵N site preference (SP) of N₂O emitted after addition of NH₂OH to several soils.

<table>
<thead>
<tr>
<th>Site</th>
<th>Turnover rate</th>
<th>δ¹⁵Nbulk [‰ vs. air N₂]</th>
<th>δ¹⁸O [‰ vs. VSMOW]</th>
<th>SP [%]</th>
</tr>
</thead>
<tbody>
<tr>
<td>coniferous forest (Ah)</td>
<td>2.0 ± 0.2</td>
<td>−8.09 ± 1.53</td>
<td>35.95 ± 9.25</td>
<td>33.02 ± 4.63</td>
</tr>
<tr>
<td>deciduous forest</td>
<td>3.5 ± 0.9</td>
<td>0.47 ± 5.68</td>
<td>47.20 ± 18.05</td>
<td>31.81 ± 14.93</td>
</tr>
<tr>
<td>grassland</td>
<td>36.9 ± 2.4</td>
<td>−6.10 ± 0.73</td>
<td>43.57 ± 4.86</td>
<td>35.21 ± 1.94</td>
</tr>
<tr>
<td>cropland (non-sterile)</td>
<td>45.8 ± 1.5</td>
<td>−5.12 ± 0.50</td>
<td>45.74 ± 3.03</td>
<td>35.66 ± 1.56</td>
</tr>
<tr>
<td>cropland (autoclaved)</td>
<td>43.7 ± 3.6</td>
<td>−7.02 ± 0.70</td>
<td>42.63 ± 4.76</td>
<td>34.12 ± 1.85</td>
</tr>
</tbody>
</table>

### 4.4. Discussion

#### 4.4.1. Abiotic N₂O formation from different soils

There has been some controversy in the past about exclusively microbial formation of N₂O during nitrification (Arp and Stein, 2003; Beaumont et al., 2002; Schmidt et al., 2004b; Yu et al., 2010). However, most studies relate N₂O production during nitrification to microbial pathways. Positive correlations between high ammonia oxidation activity and N₂O production in chemostat and mixed culture experiments have been found (Wunderlin et al., 2012; Yu et al., 2010). Although it is believed that the ammonia oxidation intermediate NH₂OH is generally not released in the environment, NH₂OH release by AOB had been reported by Stüven et al. (1992) and Schmidt et al. (2004b). Nevertheless, a large part of the soil NH₄⁺ pool will pass through NH₂OH during nitrification (Arp and Stein, 2003), and a release of NH₂OH could be followed by a fast chemical oxidation of NH₂OH leading to a non-detection. The possibility of such a mechanism has been recently emphasized by results obtained from a newly developed highly sensitive method for determination of soil NH₂OH, which revealed a positive correlation between NH₂OH and N₂O (Liu et al., 2014).

Our assumption of a possible abiotic N₂O production at aerobic conditions via oxidation of the nitrification intermediate NH₂OH could be partially verified. We demonstrated that for the agricultural and grassland soils of our study the oxidation of NH₂OH could be a potential pathway of N₂O formation, while for forest soils this mechanism seems to be of no or only minor importance. This is supported by the observation that chloroform fumigation did not lead to a complete reduction of N₂O formation from NH₂OH, and in most treatments the reduction was only small compared to non-sterile samples. As chloroform fumigation is the standard method for determining total soil microbial biomass (Jenkinson and Powlson, 1976), which requires complete lysis of microorgan-
isms, one should assume that – despite controversy whether this method is indeed completely accounting for all microorganisms in the soil – the soil should be sterile or that at least microbial activity should be reduced to a minimum. Autoclaving, on the other hand, is one of the standard methods in microbiology and medicine for sterilization. Thus, the fast formation of significant amounts of N\textsubscript{2}O in both autoclaved and chloroform-fumigated grassland and cropland soil indicated an abiotic production mechanism. An even stronger proof of an abiotic, i.e., purely chemical oxidation of NH\textsubscript{2}OH, was obtained from the incubations at different temperatures (Fig. 4.5). The increasing reaction velocity with increasing temperature, even clearly beyond common microbiological temperature optima, was in accordance with the thermodynamics of a chemical reaction. The reaction also resembled the pseudo-first order kinetics of the oxidation of NH\textsubscript{2}OH with manganese dioxide (MnO\textsubscript{2}) that we had found in supporting experiments (data not shown). For microbial processes potentially responsible for N\textsubscript{2}O formation from NH\textsubscript{2}OH added to soil, such as nitrification, a different behavior with temperature would be expected. For example, the ammonium-oxidizing bacterium *Nitrosomonas europaea* has its temperature optimum of growth and activity at around 35 °C (Grunditz and Dalhammar, 2001), and a linear relationship between activity of *N. europaea* and temperature was found between 10 and 30 °C (Groeneweg et al., 1994). Potentially there are thermophilic AOB (Lebedeva et al., 2005) or archaea (De La Torre et al., 2008) still active at elevated temperatures; however, it is unlikely that such populations developed in the short period under the prevailing experimental conditions. Thus, the faster turnover of NH\textsubscript{2}OH at 50 °C compared to 40 °C in our sterile agricultural soil clearly demonstrated the potential of this soil to oxidize NH\textsubscript{2}OH abiotically to N\textsubscript{2}O.

In our study, we found lower N\textsubscript{2}O formation in autoclaved soils than in samples fumigated with chloroform and methyl iodide. A possible explanation for this observation is that during autoclaving the soil is exposed to high temperatures and high pressure. These conditions are likely to alter the soil organic matter composition by favoring hydrolysis or organic molecules, thereby enhancing the concentration of dissolved organic matter and increasing the accessibility of reactive functional groups, like carbonyl groups, that can readily react with NH\textsubscript{2}OH, e.g., to oximes (Bremner et al., 1980). This increased competition for NH\textsubscript{2}OH between dissolved organic matter binding NH\textsubscript{2}OH and oxidants converting NH\textsubscript{2}OH to N\textsubscript{2}O, subsequently could lead to less N\textsubscript{2}O formation, as discussed in more detail below. This could also explain, why the cropland soil was least affected by autoclaving because it has the lowest organic C content, while N\textsubscript{2}O formation in autoclaved grassland soil, with a much higher organic C content, was almost zero.

The recovery rates of added NH\textsubscript{2}OH-N as N\textsubscript{2}O-N reached from insignificant in forest to about 50% in cropland soils. This recovery rates were quite stable for individual soils, thus they seemed to be influenced by soil parameters. However, sterilization methods, especially autoclaving, generally led to a reduction of the recovery rates. As autoclaving is quite a harsh sterilization method, it is
likely that it altered soil chemistry. Still, 50% or more of added NH$_2$OH-N was not detected as N$_2$O-N. The gaseous products NO or NO$_2$ can be excluded as possible further products, as no significant amounts of these gases had been observed during our studies (data not shown). It cannot be excluded that N$_2$ was a product of these incubations; however, we were not able to measure N$_2$ against the high atmospheric background, and the gas would also be of no environmental concern. A fraction of the added NH$_2$OH-N was likely bound to organic matter as oxime, and NH$_2$OH might have been also oxidized to other mineral N forms, such as nitrite, but further research is needed in this respect.

4.4.2. Factors influencing abiotic N$_2$O formation

To identify control factors of NH$_2$OH-induced N$_2$O formation and to explain the differences in NH$_2$OH oxidation potential between the different soils, we correlated N$_2$O emission rates from non-sterile as well as sterile soils with soil physical and chemical parameters (Table 4.3). The correlation of soil pH and C/N ratio with N$_2$O emission rates yielded the highest correlation coefficients of all parameters tested, especially for non-sterile and chloroform-fumigated soils. Correlation coefficients for the same correlation in autoclaved soils were lower. While the correlation between soil pH and N$_2$O emissions was positive, i.e., more N$_2$O was formed at higher pH, a negative correlation was observed for the relationship between N$_2$O emission and C/N ratio. Thus, the lower the C/N ratio the more N$_2$O was formed from the same amount of NH$_2$OH. Correlations between N$_2$O emission and soil C and N alone were also negative, but not significant, clearly indicating that the C/N ratio as an indicator of soil organic matter quality was the better predictor of N$_2$O emissions via NH$_2$OH oxidation.

Several soil cations also featured partially significant correlations with emitted N$_2$O, especially Mg. However, most of these cations were also closely correlated with soil pH (data not shown), so that the relationship between non-oxidative cation, such as Ca and Mg and N$_2$O emission was likely a cross-correlation. This assumption was supported by additional experiments in aqueous solution, affirming that neither Na and K, nor Mg and Ca were able to convert NH$_2$OH to N$_2$O (data not shown). The ability of the two remaining cations, iron and Mn, to oxidize NH$_2$OH to N$_2$O has been shown elsewhere (Bremner et al., 1980; Butler and Gordon, 1986). However, in our soils the iron content was only weakly correlated with NH$_2$OH-related N$_2$O formation. In contrast, much higher correlation coefficients were found for Mn, albeit just below significance.
Table 4.3: Pearson correlation coefficients of the linear correlation between the N$_2$O emission rates after the addition of NH$_2$OH from non-sterile, chloroform-fumigated, and autoclaved soils and several measured soil chemical parameters.

<table>
<thead>
<tr>
<th></th>
<th>pH</th>
<th>C</th>
<th>N</th>
<th>C/N</th>
<th>Ca</th>
<th>K</th>
<th>Mg</th>
<th>Na</th>
<th>Fe</th>
<th>Mn</th>
</tr>
</thead>
<tbody>
<tr>
<td>N$_2$O emission rate, non-sterile soil</td>
<td>0.97*</td>
<td>-0.78</td>
<td>-0.79</td>
<td>-0.98*</td>
<td>0.52</td>
<td>0.80</td>
<td>0.97*</td>
<td>0.79</td>
<td>0.36</td>
<td>0.79</td>
</tr>
<tr>
<td>N$_2$O emission rate, chloroform-fumigated</td>
<td>0.98*</td>
<td>-0.73</td>
<td>-0.75</td>
<td>-0.98*</td>
<td>0.54</td>
<td>0.76</td>
<td>0.96*</td>
<td>0.76</td>
<td>0.33</td>
<td>0.77</td>
</tr>
<tr>
<td>N$_2$O emission rate, autoclaved soil</td>
<td>0.87*</td>
<td>-0.50</td>
<td>-0.53</td>
<td>-0.67</td>
<td>0.66</td>
<td>0.43</td>
<td>0.73</td>
<td>0.74</td>
<td>0.09</td>
<td>0.40</td>
</tr>
</tbody>
</table>

*significant at a 0.05 level
Despite the much lower Mn content compared to the iron content of the soils (Table 4.1), this can be easily explained by the difference in redox potential of the two redox pairs Fe^{2+}/Fe^{3+} and Mn^{2+}/Mn^{4+}, which highly favors the reaction of NH_{2}OH with Mn^{4+} over the reaction with Fe^{3+}. Thus, it becomes obvious that much less abundant Mn can exert a higher control on NH_{2}OH oxidation than iron.

On the basis of the results of the correlation analysis, we conclude that there are three soil parameters, of which one or more could explain the abiotic oxidation of NH_{2}OH in soils: soil pH, C/N ratio, and soil Mn content. However, the correlation analysis is based only on a few data points, and especially for C/N ratio basically only two values are available, so that collinearity could be a problem. Thus, more research on the individual parameters and their influence on abiotic N_{2}O production is needed to verify this hypothesis. Yet, there is a chemical mechanism explaining the influence of each of the parameters. The strong influence of soil pH on abiotic N_{2}O formation can be explained chemically, as NH_{2}OH as the precursor of N_{2}O has a pK_{a} of 5.95. Below this pH value NH_{2}OH also exists in its protonated (NH_{3}OH^{+}), which is more stable than free NH_{2}OH. With decreasing pH, the ratio between protonated and unprotonated form increases, thus, at a lower pH more NH_{3}OH^{+} is present and less free NH_{2}OH is available for oxidation. The effect of C/N ratio on N_{2}O emission rates can be explained by a competitive reaction of NH_{2}OH with organic C that is higher at higher soil carbon content and even more so at wider C/N ratios. Soil organic C contains carbonyl groups, and at wider C/N ratios there are potentially more of these groups available. It was shown that NH_{2}OH can react with carbonyl groups to form oximes (R^{1}R^{2–}C=–N–OH) (Porter, 1969), and Bremner et al. (1980) found a highly significant correlation between oxime N and organic C after the addition of NH_{2}OH to sterile soils. These initial oximes can further undergo Beckmann rearrangements and form secondary organic soil constituents (Thorn and Mikita, 2000). These findings are in accordance with our results of lower N_{2}O formation with higher organic C content and also support our assumption of a strong influence of C/N ratio on abiotic N_{2}O formation. The third important soil parameter identified was soil Mn content, although the correlation coefficient is just below significance. It is known that several transition metals can oxidize NH_{2}OH to N_{2}O (Butler and Gordon, 1986). Especially Fe^{3+} had been in the focus of research, as it is ubiquitously present in almost every soil. However, Fe^{3+} ions are frequently bound in insoluble form and thus, are not readily available as reaction partner for NH_{2}OH. Therefore, despite a much lower Mn content in our soils, NH_{2}OH will preferably react with Mn^{3+} or Mn^{4+}, rather than with Fe^{3+}. This phenomenon is also made use of in soil chemical analyses for the selective extraction of Mn with NH_{2}OH, while leaving the major part of the iron unaffected (Chao, 1972).
4.4.3. Isotopic signature of abiotically produced N$_2$O

The isotopic signature of N$_2$O, especially the $^{15}$N SP of N$_2$O, is considered as a promising tool for disentangling the different N$_2$O production and consumption processes in soils (Baggs, 2008). In recent studies, it has been shown that the SP can be used to differentiate between N$_2$O production and consumption processes, i.e., nitrification and denitrification, albeit with substantial uncertainty (Ostrom and Ostrom, 2011). Only recently, Heil et al. (2014) found a remarkably constant SP in the range of 33.9–35.6‰ for different abiotic NH$_2$OH oxidation reactions in aqueous solution, that was very stable over time and different experimental conditions. Earlier, Toyoda et al. (2005) found a SP of 29.5 ± 1.1‰ for the oxidation of NH$_2$OH by MnO$_2$. In the present work with natural soils, we found the SP of the N$_2$O to be in the same range as in Heil et al. (2014). We used a two-way mixing model to calculate the isotopic composition of N$_2$O, so that the values given in Table 4.2 only represent the portion of N$_2$O produced by oxidation of added NH$_2$OH. Even if we confirmed the SP for abiotic N$_2$O formation found in solutions for soils, it is also in agreement with recent studies reporting on N$_2$O production during nitrification via microbial NH$_2$OH oxidation. In pure culture batch experiments, SP values of 33.5 ± 1.2‰, 32.5 ± 0.6‰, 35.6 ± 1.4‰, and 30.8 ± 5.9‰ were observed for *Nitrosomonas europaea*, *Nitrosomonas multiformis*, *Methylosinus trichosporium*, and *Methylococcus capsulatus*, respectively (Sutka et al., 2006; Sutka et al., 2003, 2004). Frame and Casciotti (2010) reported similar SP values for a marine nitrifying bacterium (36.3 ± 2.4‰; *Nitrosomonas marina* C-113a) and Santoro et al. (2011) found values for marine AOA that were only slightly lower but still in a range comparable to our results (30.3‰). Lately, Jung et al. (2014) found similar SP for AOA, but also found a strain with a SP as low as 13.1 ± 1.2‰, and thereby further complicating the picture. Until now, the large and partially overlapping ranges for the diverse N$_2$O production and consumption processes in soils strongly limit the use of SP for N$_2$O source partitioning. Further research on this topic and more data on individual processes are needed.

4.5. Conclusions

This study showed that at least some soils have the potential to oxidize NH$_2$OH to N$_2$O in a purely abiotic reaction, which emphasizes the possibility of a coupled biotic–abiotic production of N$_2$O during nitrification. We found this potential to be highly dependent on soil properties. Three factors that were found to possibly explain the capacity of the different soils to oxidize NH$_2$OH to N$_2$O were pH, C/N ratio, and Mn content, but further research is needed to evaluate the influence of the single parameters. Even if only a small fraction of NH$_2$OH is “leaking” into the soil during nitrification, it could be transformed to N$_2$O via fast chemical oxidation. At high nitrification rates, this could have great environmental consequences. On the basis of our findings we suggest a revision
of the ‘hole-in-the-pipe’ model for nitrification. We propose an abiotic N$_2$O production during ni-
trification that is mainly controlled by the “leakage” of NH$_2$OH from ammonia-oxidizing microor-
ganisms and the properties of the surrounding soil. If this mechanism turns out to be relevant, this
would have great implications for N$_2$O mitigation strategies, especially for agroecosystems with
high fertilizer-induced nitrification rates in combination with a high soil pH and low organic car-
bon content with low C/N ratio.
Chapter 5

N₂O decomposition over hot and dry surfaces
5.1. Introduction

Although the findings by Junge et al. (1971) of low N$_2$O concentrations in air masses of Saharan origin have been explained by Rebbert and Ausloos (1978) with a suggested tropospheric photochemical decomposition mechanism, this mechanism has since then not been studied in more detail. The photochemical and thermal decomposition of N$_2$O is a well-known process at elevated temperatures (Kondratenko and Pérez-Ramírez, 2006), but an absorption on dry particulate matter could potentially allow this destruction of N$_2$O to proceed at temperatures found on earth. As photolysis has also shown to play a role in N$_2$O production by a surface-catalyzed reaction (Rubasinghege et al., 2011), the verification of a similar process for the decomposition of N$_2$O would be of great importance for the understanding of the N cycle and could help to reduce the large uncertainties of the global N$_2$O budget.

An important factor for the relevance of the reaction is the rate of air mass flowing across a desert region in the mixed layer, in which the air stream can be mixed downward and come into contact with the desert surface. Based on the assumption of characteristic meteorological parameters for desert regions (1 km height of the boundary layer; mean wind speed of 10 m s$^{-1}$), Alyea et al. (1978) calculated that over a long desert transverse of 3000 km about 75% of the boundary layer air mass will be mixed downward to the surface layer. They concluded that only few nominal deserts ($4.5 \times 10^6$ km$^2$ surface area; the Sahara equals two nominal deserts) might lead to 15-20 year time constants for N$_2$O, at relatively low destruction efficiencies. This proposed mechanism would make deserts an effective sink in the troposphere for N$_2$O (Alyea et al., 1978).

However, this mechanism has not been considered since its discovery, but would be of great importance with increasing anthropogenic N$_2$O emissions, and would greatly improve modeling of the global N$_2$O cycle. The discovery of an additional tropospheric sink of global significance would largely expand our understanding of the global N$_2$O cycle and the global N$_2$O budget.

To show the photochemical decomposition of N$_2$O over warm and dry sand surfaces, a laboratory experiment was set up, using a flow-through reaction chamber in a closed loop connected to an infrared laser absorption spectrometer for online real-time analysis of N$_2$O mixing ratio. Quartz sand as the main constituent of continental deserts was chosen as the reactive surface, also in mixtures with the transition metal oxides ferric oxide (Fe$_2$O$_3$) and manganese oxide (MnO$_2$) that are commonly found as components of desert sands. Both iron and manganese oxides, have been shown to have a catalytic effect on the decomposition of N$_2$O at elevated temperatures (Kondratenko and Pérez-Ramírez, 2006; Yamashita and Vannice, 1996), thus it can be assumed that they also influence a potential decomposition mechanism in hot desert regions. Experimental conditions were chosen to resemble conditions typically found in desert regions with high intensity UV radiation, high surface temperatures, and dry air.
The use of laser absorption spectroscopy allows for a high-precision measurement of N\textsubscript{2}O that is superior to that of gas chromatography, and additionally enables measurements at very high temporal resolution. Those two factors make laser spectroscopy an ideal tool to observe potentially low N\textsubscript{2}O degradation rates at the laboratory scale.

### 5.2. Materials and Methods

For the determination of N\textsubscript{2}O mixing ratios an infrared laser absorption spectrometer was used (Dual Laser Quantum Cascade Trace Gas Monitor, Aerodyne Research, Inc., Billerica, MA, USA). The instrument consists of two mid-infrared lasers that are able to measure N\textsubscript{2}O, CO\textsubscript{2}, CH\textsubscript{4}, and H\textsubscript{2}O simultaneously at a temporal resolution of up to 10 Hz. Water vapor is measured to correct for volumetric dilution and pressure broadening effects caused by increasing water vapor concentration in the sample air. Prior to every experiment the cell was flushed with synthetic air and a new background spectrum was taken. The instrumental precision of the measurements, expressed as standard deviation of the average N\textsubscript{2}O mixing ratio at atmospheric level was <0.3 ppb.

The laser was connected to a flow-through reaction chamber in a closed loop. Dried hardware store quartz sand was used as reactive surface in our experiments. For testing the effect of iron and manganese oxides on N\textsubscript{2}O decomposition, the quartz sand was also mixed with 2.5% (m/m) Fe\textsubscript{2}O\textsubscript{3}, and further additionally with 0.25% (m/m) MnO\textsubscript{2}. The sand was dried prior to each experiment at 105 °C for at least 24 h. At the beginning of each experiment, the hot sand was taken out of the drying oven and was immediately filled into a flow-through reaction chamber. The chamber was custom-made of a 30 cm long quartz glass cylinder fitted with PTFE cover plates, and one connection at each side for in- and outlet. The PTFE plates were hold together by seven aluminum rods at the outside of the chamber. The contact surfaces were sealed gastight with Viton® O-rings. A thermocouple was fitted inside the chamber to monitor the temperature close to the sand surface during the experiment. The 30 cm long chamber had a diameter of 14 cm, and was filled with sand just less than half corresponding to a sand surface area of approximately 420 cm\textsuperscript{2}. The chamber was then purged with 2.5 L min\textsuperscript{1} of a standard gas of known concentration of N\textsubscript{2}O (250, 500, or 750 ppb balanced with N\textsubscript{2}; 99.5% purity N\textsubscript{2}O, Linde, Munich, Germany) or pressurized air of variable N\textsubscript{2}O concentration (Forschungszentrum Jülich GmbH, Germany), controlled by a mass flow controller (Brooks Instruments, Dresden, Germany). When the whole system had equilibrated and a constant mixing ratio of N\textsubscript{2}O was reached after a few minutes, magnetic 3-way valves (series 9, Parker Hannifin, Cleveland, OH, USA) were switched and the chamber was put into a closed loop, with the outlet of the chamber connected to the inlet of the laser and the outlet of the laser connected to the inlet of the chamber via a membrane pump (MVP 070-3, Pfeiffer Vacuum, Asslar, Germany) to achieve constant circulation (Fig. 5.1). Two 300 W UV-light (Ultra-Vitalux®, Osram,
Augsburg, Germany) that produce a radiation spectrum similar to that of natural sunlight, and two 250 W (Sicatherm®, Osram) infrared (IR)-light lamps with a dominant wavelength of 1100 nm and a low visible light fraction were placed 60 cm above the reaction chamber for high intensity UV radiation and to keep the surface temperature of the sand high, as found in hot desert regions. A bypass loop to detach the chamber from the closed loop and to shut it off completely was added to be able to run experiments also in batch mode.

For this thesis, five different experiments were conducted to study N₂O photodecomposition. In the beginning, two experiments (1 and 2) in a closed loop mode were carried out with quartz sand filled into the reaction chamber, first using a reference gas with sub-ambient N₂O mixing ratio (281 ppb), and then a second gas with above-ambient N₂O mixing ratio (500 ppb) over periods of 17 and 29 h, respectively. During these experiments the chamber was exposed to the UV and IR radiation of the four light sources over the complete experimental run. Further, an experiment was conducted under the same conditions, but with a simulated diurnal day/night cycle (6 h/16 h) that was achieved by switching the UV and IR radiation off during night (experiment 3). In a next step, the experiments 1 and 2 with a permanent light source were repeated under the same conditions as before for approximately 24 h, but the sand was additionally mixed with Fe₂O₃ and MnO₂ as described above (experiment 4). In contrast to the previously described measurements that were run in continuous closed-loop mode, a final experiment was conducted in batch mode (experiments 5), in which the reaction chamber was disconnected from the closed-loop and N₂O mixing ratios were measured repeatedly in three-hour intervals. Light conditions were the same as in the experiments with continuous light before.

![Figure 5.1](image-url): Schematic representation of the laboratory setup for the detection of photochemical N₂O decomposition over a hot and dry sand surface with a dynamic flow-through chamber coupled to a quantum cascade laser absorption spectrometer (QCLAS).
5.3. Results

With the performed experiments it was not possible to find a clear evidence for a N₂O decomposition mechanism that could be a potential sink for N₂O in hot desert regions. When a reference gas with a sub-ambient N₂O mixing ratio of 281 ppb was utilized, the closed loop experiments with high UV radiation and at high temperature of the sand surface (53 °C) revealed an increase of N₂O mixing ratios close to ambient levels over time (experiment 1; Fig. 5.2 A). When the chamber was filled with a reference gas with a higher than ambient N₂O mixing ratio (500 ppb), the opposite trend was observed, showing a gradual decrease to the ambient N₂O mixing ratio (experiment 2; Fig. 5.2 B). Both experiments shown in Fig. 5.2 were conducted under the same conditions, using two UV lamps and one IR lamp during the complete experimental run. In the beginning of the experiment with sub-ambient reference gas, the N₂O mixing ratio slightly decreased by 1 ppb over a period of 30 min and stayed at a level of about 281 ppb for about half an hour (Fig. 5.2 A). After this initial phase, the mixing ratio continuously increased until the end of the experiment. The experiment with a 500 ppb reference gas (Fig. 5.2 B) on the other hand showed the opposite trend, with N₂O mixing ratio decreasing from the beginning of the experiment. The speed of decrease slowed down as the mixing ratio approached the ambient level.

![Figure 5.2](image-url): Nitrous oxide (N₂O) mixing ratios in ppb (red line) over time at a temporal resolution of 1 Hz and sand near-surface temperature in °C (black line) in closed-loop experiments with simulated hot desert daytime conditions in a dynamic flow-through reaction chamber filled about half-full with sand and irradiated with ultraviolet (UV) and infrared (IR) light. The system was flushed with pressurized air with 281 ppb N₂O in experiment 1 (A) and with a 500 ppb N₂O reference gas in experiment 2 (B) prior to the experiment.
Figure 5.3 shows the results of experiment 3 in which a diurnal day–night–day cycle (6 h/16 h/7 h) was simulated. The reference gas used to flush the system was dry pressurized air with a N$_2$O mixing ratio of 297 ppb. During daytime conditions, the chamber was irradiated with two UV lamps and one IR lamp, which we all switched off during night. During daytime conditions sand temperatures reached a maximum of 60 °C and cooled down to a temperature of 27 °C at the end of the night. At the beginning of the experiment, when daytime conditions were simulated, N$_2$O mixing ratios were relatively stable for about one hour. This stage was then followed by a steady increase of N$_2$O mixing ratio. With the beginning of nighttime conditions, the increase stopped, and for about three hours N$_2$O mixing ratios decreased. After the decrease phase, the mixing ratios gradually increased again until the next daytime light phase, when this increase was further enhanced up to the point, at which N$_2$O mixing ratios close to the atmospheric level (ca. 324 ppb) were reached. In contrast, water vapor mixing ratios behaved differently. Mixing ratios sharply decreased at the beginning of the experiment from the initial level of the pressurized air of 2400 ppm until a constant level of about 795 ppm at the end of the first daytime phase was reached. Upon the change from daytime to nighttime conditions, the same sharp decrease as at the beginning of the experiment could be observed. After this initial decrease, a low but steady increase in water vapor mixing ratio was observed at nighttime conditions. With the beginning of the second daytime phase a strong increase in water vapor was observed.
As desert sand also includes other substances than silica, such as transition metal oxides, and as transition metal oxides could play an important role in surface-bound reaction mechanisms, further experiments with sand and metal oxide mixtures were conducted under the same conditions as in experiment 1. Experiment 4 with sand and Fe₂O₃ and MnO₂ mixture showed no significant difference compared to experiment 1 with pure quartz sand (Fig. 5.4) that has been conducted under the same conditions, so that a possible involvement of transition metals in an adsorption mechanism by silica sand could not be confirmed. The N₂O mixing ratio increased from the beginning of the experiment on until the ambient level was reached. By the end of the experiment, no further change in N₂O mixing ratio was observed. The water vapor mixing ratio, on the contrary, strongly decreased at the start of the experiment for about four hours down to 230 ppm, after which it gradually increased up to slightly above 1000 ppm until the end of the experiment.

Figure 5.4: Nitrous oxide (N₂O) mixing ratios in ppb and water vapor (H₂O) mixing ratios in ppm over time at a temporal resolution of 1 Hz in a closed loop experiment (experiment 4) with simulated daytime conditions in a dynamic flow-through reaction chamber filled about half-full with sand mixed with 2.5% ferric oxide (Fe₂O₃) and manganese oxide 0.25% (MnO₂) and irradiated with ultraviolet (UV) and infrared (IR) light. The system was flushed with pressurized air (225 ppb N₂O) prior to the experiment.

As apparently a small leakage in the experimental setup led to the observed changes in N₂O mixing ratios, very likely at the stage of the membrane pump, experiment 5 was run in batch mode, thereby excluding the pump and the analyzer for the duration of the batch mode. For this, the chamber was filled with a mixture of a reference gas with a N₂O mixing ratio of 500 ppb and pressurized air (294
ppb N$_2$O) to obtain a N$_2$O mixing ratio close to the ambient level. After that, the chamber was disconnected from the pump and the analyzer by magnetic 3-way valves (Fig. 5.1). The closed chamber was exposed to the UV and IR radiation of two UV lamps and one IR lamp placed 60 cm above the chamber. This corresponded to a sand surface temperature of approximately 60 °C. After periods of three hours, the mixing ratio of N$_2$O in the chamber air was measured by switching the chamber back into loop mode for 5 min. The initial N$_2$O mixing ratio averaged over 5 min was 328.1 ± 0.2 ppb. After three hours the N$_2$O mixing ratio had decreased significantly to 325.9 ± 0.2 ppb. However, there was no further significant reduction in N$_2$O after another period of three hours, when the mixing ratio remained at a similar value of 325.7 ± 0.2 ppb. As the chamber itself was gastight, it can only be concluded that there was no photo-degradation of N$_2$O ongoing in the experiments, or the degradation rate was so low that a change in N$_2$O mixing ratio at the end of the experiment was below the detection limit.

5.4. Discussion

The results of the photo-degradation experiments clearly indicated a small but constant leakage in the loop system with ambient air diffusing into the system. There were several possible sources of leakage in the following way. The flow-through chamber and its connections could be excluded, as gas-tightness had been tested and no leakage had been detected at all. Another potential source of gas diffusion into the closed-loop system was the PTFE tubing. However, although N$_2$O can diffuse through the PTFE material (Dobson and Taylor, 1986), the permeability is low, but could explain part of the observed leakage. Another potential source of leakage was the membrane pump with its four membrane heads. Additionally, the adsorption cell of the laser spectrometer was found not to be gastight. These sources added up to the total leakage rate of the system that caused ambient air to slowly diffuse into the closed loop. By this, the results can only lead to the assumption that if the N$_2$O photo-degradation existed in hot desert regions and occurred in the experimental system simulating these desert conditions, it would have been smaller than the leakage rate found in the setup.

The simulated diurnal cycle in experiment 3 (Fig. 5.2) could also give no evidence of a N$_2$O photo-degradation. Moreover, it showed possible effects of adsorption and desorption on the sand surface that could, together with the leakage, explain the variability in the mixing ratios of N$_2$O and water vapor. At the beginning of the experiment, when the sand was still completely dry, the results showed signs of an adsorption of water vapor from the already relatively dry reference gas on the sand surface, until the adsorption capacity of the sand was reached. At the beginning of the night cycle, decreasing temperatures may have allowed even more water vapor to be adsorbed by the sand surface. Also N$_2$O might have been adsorbed during the cooling period of the sand. When the
temperature had stabilized to ambient laboratory conditions, and adsorption capacity was reached again, both mixing ratio curves of N$_2$O and water vapor describe the diffusion of ambient air into the loop through the leakage in the system. The strong increase of N$_2$O and water vapor mixing ratios with the warming up at the next daytime conditions could be explained by a desorption of previously adsorbed N$_2$O and water vapor. In this experiment, significantly more water vapor compared to N$_2$O was potentially adsorbed.

Diurnal variability of N$_2$O mixing ratios in the atmosphere has been reported in the past by several authors (Brice et al., 1977; Cicerone et al., 1978; Matthias et al., 1979; Pierotti et al., 1978) and has also been attributed to a potential tropospheric N$_2$O sink via photochemical decomposition as proposed by Rebbert and Ausloos (1978). Albeit a diurnal variation of N$_2$O mixing ratios could not be confirmed by Cofer et al. (1986), a potential sink for N$_2$O could exist at the regional scale, such as in hot desert regions (Pierotti et al., 1978). Although, the presented diurnal cycle results could not verify a N$_2$O decomposition process, they showed that potential absorption/desorption processes could explain small diurnal changes in N$_2$O mixing ratios.

Although some small decreases in N$_2$O mixing ratio were observed in experiments 1 and 3 (Fig. 5.2A and Fig. 5.3), they could more likely be associated with adsorption and not destruction of N$_2$O, as the decreases were only observed when the sand was still very dry at the beginning of the experiment or when it was cooling down from higher temperatures. Furthermore, these decreases where followed by strong increases in N$_2$O mixing ratio when temperatures increased again, likely by desorption. Beside potential adsorption of N$_2$O molecules to the sand surface, there was no evidence for any other N$_2$O-consuming process, such as photochemical destruction of N$_2$O.

Although it is known that transition metal oxides can have a catalytic effect on photochemical N$_2$O destruction (Kondratenko and Pérez-Ramírez, 2006; Yamashita and Vannice, 1996), this effect could not be shown for the potential N$_2$O decomposition over hot sand. As Rebbert and Ausloos (1978) stated, small amounts of water vapor would drastically reduce photolysis. This would mean that the presence of water vapor as low as in the present experiments would already suppress N$_2$O photolysis, and therefore this mechanism would not be of relevance in regions where water vapor mixing ratios exceed this threshold. On the contrary, if water vapor below this threshold does not suppress N$_2$O photo-degradation, the mechanism could still be of significance at longer timescales in very dry and hot desert regions, although it was not detectable in experiments of only several hours duration. The N$_2$O decomposition rate can anyway only be low, as Pierotti et al. (1978) only found small changes in N$_2$O mixing ratios in the large desert area of the Sahara.
5.5. Conclusions

With the presented experiments it could not be confirmed that a mechanism of \( \text{N}_2\text{O} \) photodecomposition over warm and dry sand surfaces existed, as suggested by Rebbert and Ausloos (1978). However, it can neither be confirmed nor denied that this mechanism exists. With the experimental setup it was not possible to detect the decomposition of \( \text{N}_2\text{O} \), which is potentially very low within a short timeframe, due to limitations of the equipment as discussed above. It is also possible, that the rate is too low to be observed with the existing leakage rate, but could still be of environmental importance in large desert areas that allow for long residence times of air masses in such areas. That is why further research on this process is necessary, as this potential sink for \( \text{N}_2\text{O} \) would be the only known tropospheric sink for \( \text{N}_2\text{O} \) besides denitrification that can act as both a source and sink of \( \text{N}_2\text{O} \). This would lead to a revision of the global \( \text{N}_2\text{O} \) budget, and a revision of existing models could help to close gaps in the global \( \text{N}_2\text{O} \) budget. A great part of this uncertainty is due to the uncertainty in the dominant loss term of \( \text{N}_2\text{O} \) (Ciais et al., 2013), and a newly discovered \( \text{N}_2\text{O} \) sink would imply that the previously estimated source strength of \( \text{N}_2\text{O} \), which is derived via a top-down approach, was too low.
Chapter 6

Synopsis
6.1. Summary

This thesis was laid out to characterize abiotic N trace gas formation processes in soils that have been known for several years, but are still not well understood and are widely neglected in most current studies. The first part of the thesis was a review gathering knowledge about abiotic N trace gas formation processes that built the basis for designing the experiments. The experiments mainly focused on the abiotic production of N\textsubscript{2}O from the nitrification intermediate NH\textsubscript{2}OH, characterized the isotopic signature of abiotically produced N\textsubscript{2}O, demonstrated abiotic N\textsubscript{2}O formation in soils, but also looked at potential photochemical N\textsubscript{2}O decomposition on hot and dry sand surfaces.

The second chapter of this thesis was a review that updated the information about the role of abiotic processes in the formation of gaseous N products in soils. Several reactions involving the nitrification intermediates NO\textsubscript{2}\textsuperscript{−} and NH\textsubscript{2}OH are known to produce the N trace gases NO and N\textsubscript{2}O. These reactions are: (i) the self-decomposition of NO\textsubscript{2}\textsuperscript{−}, (ii) reactions of NO\textsubscript{2}\textsuperscript{−} with reduced metal cations, (iii) the nitrosation of NO\textsubscript{2}\textsuperscript{−} by SOM, (iv) the reaction between NO\textsubscript{2}\textsuperscript{−} and NH\textsubscript{2}OH, and (v) the oxidation of NH\textsubscript{2}OH by iron and manganese. The reactions were presented in great detail with their gaseous products, potential environmental relevance, and influential soil parameters. While the gathered information showed that these reactions could occur over a broad range of soil characteristics, it was also found that these reactions are ignored in most current studies in favor of biological N trace gas formation. Relatively few studies were found that tried to quantify the contribution of abiotic processes to total N trace gas emissions of soils, which leaves great uncertainty in emission models and mitigation strategies. The negligence was found to be mainly due to the simultaneous occurrence of biotic and microbial processes in close vicinity, which makes it difficult to discriminate between the different processes. The review revealed participation of the nitrification intermediates NO\textsubscript{2}\textsuperscript{−} and NH\textsubscript{2}OH in all known abiotic N trace gas formation processes. By this, the review emphasized a coupled biotic–abiotic mechanism, with biotic nitrification supplying the substrates that are subsequently converted abiotically to NO and N\textsubscript{2}O. For the first time, these processes have been merged into a conceptual model explaining abiotic NO and N\textsubscript{2}O formation during nitrification. In an outlook, the review further showed the potential of stable isotope techniques to disentangle the different N\textsubscript{2}O production processes in soils. Based on the knowledge from the review, experiments were developed to characterize the abiotic processes.

The first experiments, presented in chapter three, looked at the site-specific isotopic signatures of abiotically produced N\textsubscript{2}O, but prior to this, preliminary experiments were conducted using a QCLAS to study the contribution of different abiotic processes to NO and N\textsubscript{2}O formation. It was observed that reactions involving NO\textsubscript{2}\textsuperscript{−} mainly led to the formation of NO while reactions with NH\textsubscript{2}OH mainly led to the formation of N\textsubscript{2}O, as did the comproportionation of NO\textsubscript{2}\textsuperscript{−} and NH\textsubscript{2}OH. This comproportionation as well as the NO\textsubscript{2}\textsuperscript{−} decomposition to NO were limited to low pH conditions (< pH 5.5), while reactions of NH\textsubscript{2}OH with transition metals occurred over a wide pH range.
As the SP of $^{15}$N in N$_2$O is a promising tool to give more insight into N$_2$O production processes, the SP of N$_2$O produced by different abiotic reactions that had been identified to lead to significant N$_2$O formation reactions, was determined in a laboratory study. All reactions involved the nitrification intermediate NH$_2$OH in combination with different soil constituents (NO$_2^-$, Fe$^{3+}$, Fe$^{2+}$, Cu$^{2+}$). The experiments were conducted in aqueous solution placed in flow-through reaction chambers. N$_2$O production and its four main isotopic species ($^{14}$N$^{14}$N$^{16}$O, $^{15}$N$^{14}$N$^{16}$O, $^{14}$N$^{15}$N$^{16}$O, $^{14}$N$^{14}$N$^{18}$O) were quantified simultaneously, online and at a temporal resolution of 1 Hz, using a QCLAS. Thereby, the study presents the first continuous analysis of $\delta^{18}$O in N$_2$O. The experiments revealed the possibility of purely abiotic reactions over a wide range of acidity (pH 3–8) by different mechanisms: the reaction of NH$_2$OH with NO$_2^-$ at low pH, the oxidation of NH$_2$OH by Fe$^{3+}$ (pH 3–8), and the Cu$^{2+}$ catalyzed autoxidation of NH$_2$OH at higher pH. The $\delta^{15}$N and $\delta^{18}$O of N$_2$O produced by these abiotic pathways were significantly different between reaction mechanisms and reaction conditions. The $\delta^{15}$N of N$_2$O reflected the $\delta^{15}$N of the precursors NH$_2$OH (-1.93‰) and NO$_2^-$ (-27.0‰) only partially. Reactions with the same N substrate resulted in significantly different $\delta^{15}$N values under different reaction conditions, which limits the identification of the N source from the bulk $^{15}$N isotopic signature of N$_2$O. For $\delta^{18}$O of N$_2$O, a similar picture emerged, very likely caused by complex O exchange processes between water and dissolved N species. All abiotic pathways showed a characteristic SP of $\delta^{15}$N in N$_2$O of about 35‰. The SP was unaffected by process conditions such as pH, as well as the type of reaction, and remained constant during the experiments. These new findings contribute new information to the challenge of source partitioning of N$_2$O emissions from soils. As hypothesized, a distinct SP of N$_2$O was found, but as also assumed, this constant SP was in the same range as previously found for microbial nitrification and fungal denitrification.

As the first experiments only looked at abiotic N$_2$O formation from NH$_2$OH in aqueous solutions, further experiments, presented in chapter four, were conducted with natural soils to analyze whether this abiotic N$_2$O formation from NH$_2$OH can occur in soils. For this, N$_2$O formation from NH$_2$OH was studied in laboratory incubation experiments in cropland, grassland, and forest soils. Incubations were conducted with and without the addition of NH$_2$OH to non-sterile and sterile soil samples. N$_2$O evolution was quantified with gas chromatography and further analyzed with online laser absorption spectroscopy to get insight into the N$_2$O formation dynamics. Additionally, the isotopic signature of the produced N$_2$O ($\delta^{15}$N, $\delta^{18}$O, and $^{15}$N SP) was analyzed with isotope ratio mass spectrometry. The different soils showed large differences in N$_2$O formation upon the addition of NH$_2$OH. While the forest soil samples showed hardly any N$_2$O evolution after addition of NH$_2$OH, immediate and very large formation of N$_2$O was observed in the cropland soil, also in sterilized samples. There were also differences in N$_2$O formation observed in soils subjected to different sterilization methods. Autoclaving significantly reduced N$_2$O emissions from most soils,
while chloroform fumigation reduced N<sub>2</sub>O formation only slightly compared to non-sterile soils. The experiments could show that sterile soils can oxidize added NH<sub>2</sub>OH abiotically, and that at least some of the soils used in the experiments, mainly the agricultural soil, had an oxidation potential for NH<sub>2</sub>OH as hypothesized before. This oxidation potential was highly dependent on soil pH, C/N ratio, and manganese content, in descending order, as revealed by a correlation analysis. N<sub>2</sub>O formation was lower at lower pH. This could be explained by the higher chemical stability of NH<sub>2</sub>OH at a lower pH due to a higher degree of protonation of the NH<sub>2</sub>OH molecule. A higher C/N ratio led to a significant reduction of abiotic N<sub>2</sub>O, which could be explained by an increasing incorporation of NH<sub>2</sub>OH into the organic material, especially with high C/N ratio, that could act as a competitive reaction for NH<sub>2</sub>OH. High manganese content correlated with high N<sub>2</sub>O formation, as manganese was shown to be able to oxidize NH<sub>2</sub>OH to N<sub>2</sub>O. Interestingly, no influence of iron on N<sub>2</sub>O formation was observed, as obviously iron is bound too tightly in the soil matrix to be able to participate in the oxidation of NH<sub>2</sub>OH to N<sub>2</sub>O. The kinetics of the observed N<sub>2</sub>O formation resembled the same immediate and strong formation of N<sub>2</sub>O as for the equivalent abiotic reaction in aqueous solution. Also the temperature dependency of the reaction in soils corresponded to the thermodynamic behavior of a purely chemical reaction, so that it could be concluded that the observed NH<sub>2</sub>OH-induced N<sub>2</sub>O was largely of abiotic origin. The SP of <sup>15</sup>N in N<sub>2</sub>O was in the same range as previously found for abiotic N<sub>2</sub>O formation as well as for microbial nitrification and fungal denitrification. With these experiments it was possible to show that at least some soils can oxidize NH<sub>2</sub>OH to N<sub>2</sub>O as hypothesized. The results suggest a possible coupled biotic–abiotic production of N<sub>2</sub>O during nitrification, e.g., due to leakage of the nitrification intermediate NH<sub>2</sub>OH with subsequent reaction in the soil matrix.

The possibility of a potential N<sub>2</sub>O decomposition mechanism via photolysis over hot sand surfaces was investigated in laboratory experiments as presented in chapter five. Laboratory experiments were designed, using a flow-through reaction chamber filled with sand coupled to a QCLAS in a closed loop to continuously monitor N<sub>2</sub>O mixing ratios. Experimental conditions were chosen to simulate hot desert daytime conditions, i.e., high surface temperatures and high UV radiation. However, with the conducted experiments it was not possible to show a photochemical destruction of N<sub>2</sub>O over hot sand surfaces. This was mainly due to a slight leakage of ambient air into the closed loop setup, so that no unambiguous conclusion could be drawn. However, it can be inferred that the potential photochemical process, if it exists, must have a low turnover rate, making it very hard to detect in a short timeframe. Thus, the existence of this process could neither be proven nor denied, and a potential involvement of transition metal oxides, such as iron and manganese oxide, could also not be observed.
6.2. Synthesis

The aim of this thesis was to evaluate the role of abiotic processes in N trace gas formation and decomposition in soils. Despite the fact that several reactions leading to the formation of NO or N₂O have been known for years, they are widely neglected in most current studies. This thesis compiles the scattered studies on known abiotic N trace gas formation processes and merges them into a conceptual model of abiotic N trace gas formation during microbial nitrification. Based on this model, experiments were designed to study N₂O formation by NH₂OH oxidation in soils. The experiments gave evidence of N₂O formation via abiotic NH₂OH oxidation, and influences of different soil parameters could be determined. With the obtained results, the hypothesis of a coupled biotic–abiotic N₂O formation, as presented in the conceptual model, could be partially confirmed.

The in-depth literature review, as presented in chapter two, allowed a comprehensive summary of all abiotic N₂O and NO production and consumption processes that are known to occur in soils. By compiling all known reactions, it was possible to divide abiotic N trace gas production processes into two groups. The first group included reactions involving NO₂⁻. These reactions largely lead to the formation of NO, with N₂O as a side product only, which have classically been labeled as chemodenitrification. The second group comprised the reactions involving NH₂OH, being responsible for the majority of abiotic N₂O formation. Additionally, an involvement of SOM in these mechanisms was found that has not been well investigated and that obviously plays an ambivalent role, i.e., on the one hand being able to abiotically fix mineral forms of N, and on the other hand being involved in N trace gas formation. Preliminary experiments could confirm this grouping into chemodenitrification, leading to mainly NO, and NH₂OH oxidation leading to N₂O formation. As the two precursors of abiotic N trace gas formation, NO₂⁻ and NH₂OH, are both intermediates of microbial nitrification, this thesis proposes a coupled biotic–abiotic production of N₂O, with nitrification providing the substrates, which are subsequently oxidized or reduced in a purely abiotic fashion. This coupled mechanism, with both abiotic and biotic processes proceeding at the same time, makes it very difficult to differentiate between both. This probably led to an underestimation of abiotic N trace gas formation in the past. While most of the occasional studies on abiotic N trace gas formation focused only on single processes or groups of processes, the comprehensive review in this thesis for the first time allowed a conceptualization of all known abiotic N₂O and NO, as well as HONO producing processes from nitrification intermediates NO₂⁻ and NH₂OH into one model. This conceptual model can be considered as an revision and extension of the ‘hole-in-the-pipe’ model (Firestone and Davidson, 1989). The model allows the explanation of the formation N trace gases N₂O, NO, and HONO during nitrification by all known abiotic processes involved.

The production of NO via chemodenitrification is generally better understood and accepted than NH₂OH oxidation, as the precursor NO₂⁻ is, unlike NH₂OH, always released by AOB into the soil matrix. Although it generally does not accumulate in soils (see chapter 2.2), part of the NO₂⁻ can
undergo chemical decomposition primarily to NO. As HNO₂ rather than NO₂⁻ is the reactive species in these NO₂⁻ decomposition reactions, they are highly pH-dependent. This is why chemodenitrification is considered an important process for N trace gas loss only in acidic soils, e.g., in temperate coniferous forests (Kesik et al., 2006; Stange et al., 2000). NH₂OH oxidation was for a long time disregarded, primarily because of the non-detection of NH₂OH in soils. This could be explained by the highly reactive character of NH₂OH and the lack of a sensitive detection method, that has only very recently been introduced (Liu et al., 2014). The high reactivity and non-detection may be two reasons why abiotic N₂O production via NH₂OH oxidation has been neglected in favor of microbial pathways, and this is why the experiments designed for this thesis focused on trying to confirm the relevance of abiotic NH₂OH oxidation.

First experiments with the nitrification intermediates NO₂⁻ and NH₂OH in combination with several transition metals in solution supported the assumption of an abiotic oxidation of NH₂OH to N₂O. Experiments confirmed for one the assumption that reactions involving NO₂⁻ only, in the absence NH₂OH, produce mainly NO and that NH₂OH is required to produce significant amounts of N₂O. The results presented in chapter three showed that reactions of NH₂OH-induced N₂O formation are possible over a wide pH range in combination with several transition metals.

Interesting information could be gained from the isotopic data collected. The study showed that new quantum cascade laser absorption spectroscopy technology is a suitable tool for getting more insight into the N cycle, and potentially other biogeochemical cycles, with high precision and at a high temporal resolution. The new technology revealed interesting features of the examined abiotic reactions, as an inverse isotopic fractionation during the reaction of NH₂OH and Cu²⁺, and the surprisingly stable and constant SP for all abiotic N₂O production processes that resembled the SP found for N₂O production during nitrification and fungal denitrification. It was shown that different experimental conditions exerted no influence on the measured SP. It could be assumed, that the SP is independent of the N substrate, and not necessarily represents the production mechanism, but rather the last intermediate product in the formation of N₂O, as assumed by some authors before (Fehling and Friedrichs, 2011; Toyoda et al., 2002). However, this limits the use of the SP for source partitioning N₂O emissions, as reflected in SP values in a similar range for abiotic NH₂OH oxidation and NH₂OH oxidation in cell cultures. Based on these results one can argue that both microbial and abiotic processes share the same last intermediate step in N₂O formation, or alternatively it could be interpreted that N₂O produced allegedly microbially during nitrification could in reality be derived from the chemical oxidation of the nitrification intermediate NH₂OH, released by AOB.

In chapter four experiments were brought to next level, in which reactions of NH₂OH with soil constituents and proposed mechanisms were tested in natural soils. The conducted experiments showed that at least some soils have a potential to oxidize NH₂OH in a purely abiotic way. This
potential was highly dependent on soil properties. The observed site-specific isotopic signature of N\textsubscript{2}O produced in soils was in the same range as in the previous experiments, which is also the same range as for N\textsubscript{2}O from nitrification and fungal denitrification. As the isotopic signatures could give no distinct evidence for an abiotic N\textsubscript{2}O formation, incubation experiments with sterilized soils clearly showed that some soils can oxidize NH\textsubscript{2}OH abiotically, as sterile samples showed similar N\textsubscript{2}O production upon NH\textsubscript{2}OH addition compared to non-sterile replicates. Even stronger evidence of an abiotic N\textsubscript{2}O formation was given by a reaction kinetics study with a QCLAS. The fast character of the reaction resembles the reaction progress of NH\textsubscript{2}OH oxidation found in the previous experiments in solution. Additionally, the temperature dependence of the reaction was as expected for a chemical reaction with pseudo-first-order kinetics, as observed for the oxidation of NH\textsubscript{2}OH by Fe or Mn (Butler and Gordon, 1986). These results clearly demonstrated that NH\textsubscript{2}OH added to grassland and agricultural soils was oxidized abiotically. In the environment, however, this N\textsubscript{2}O formation will be controlled by the NH\textsubscript{2}OH release rate by AOB and by the prevailing soil parameters as previously described in the conceptual model (Fig. 2.1).

An interesting feature when looking at the influential factors on abiotic N\textsubscript{2}O formation was that Fe seemed to have no influence on the abiotic oxidation of NH\textsubscript{2}OH, whereas Mn, albeit about a factor of ten lower in concentration than Fe, played a significant role in NH\textsubscript{2}OH oxidation. It seemed as if the soil Fe was too tightly bound to be available for NH\textsubscript{2}OH oxidation and that the role of Fe on NH\textsubscript{2}OH oxidation was previously overestimated due to its generally high abundance, although Mn has the higher redox potential. SOM seems to play another important role in abiotic N\textsubscript{2}O formation by binding NH\textsubscript{2}OH to functional groups of lignin or dissolved humic acid and, thereby, suppressing its oxidation to N\textsubscript{2}O, so that abiotic N\textsubscript{2}O formation seems to be of minor importance in soils rich in organic matter. This incorporation of NH\textsubscript{2}OH into SOM is expected to be highest at near neutral pH conditions (Thorn and Mikita, 2000).

This thesis gave strong evidence that abiotic N trace gas formation has to be considered when evaluating gaseous N emissions from soils. The abiotic NO formation via chemodenitrification is a process, which has to be considered for acidic soils. Although conditions favorable for NO\textsubscript{3}– accumulation usually do not coincide with conditions favorable for NO\textsubscript{2}– decomposition, the processes could be of significance in strongly acidic soils, in which NO\textsubscript{2}– can be quickly decomposed before being converted to NO\textsubscript{3}– via nitrification. The N\textsubscript{2}O formation via NH\textsubscript{2}OH oxidation, on the other hand, is obviously underrepresented in N\textsubscript{2}O emission studies, as this thesis demonstrated that it can be a relevant process, e.g., in agricultural soils with high nitrification rates, a high pH, and low organic C. It can be assumed that at least a certain part of N\textsubscript{2}O emissions previously attributed to microbial processes is of chemical origin, albeit with an uncertainty of its magnitude and contribution to total soil N\textsubscript{2}O emissions.
In the last part of the thesis, experiments have been conducted to verify a potential decomposition mechanism over hot and dry surfaces. Although an abiotic N\textsubscript{2}O trace gas decomposition reaction had been reported in the past, it could not be identified with the experimental setup used in this thesis. That said, the conducted experiments could neither confirm nor negate the existence of such a mechanism. As discussed above, the experimental setup had limitations that allowed ambient air to diffuse into the closed loop system and thus masked potential N\textsubscript{2}O decomposition in the closed system. It can only be concluded that if a photochemical decomposition of N\textsubscript{2}O in hot desert areas existed, the rate of this process is potentially very low in a short timeframe. However, with long desert transects resulting in long residence times of air masses in hot desert regions, this process could still be a significant sink of N\textsubscript{2}O at a global level.

6.3. Perspectives

6.3.1. Source partitioning of N\textsubscript{2}O using stable isotopes

Great expectations have been set into the source partitioning of different N\textsubscript{2}O emission sources using stable isotopes, especially using the site-specific \textsuperscript{15}N isotope data. Although at present this site-specific \textsuperscript{15}N isotope information can only be used to differentiate between oxidative and reductive N\textsubscript{2}O emission sources (Decock and Six, 2013), and the results gained in this thesis cannot be used to differentiate between microbial and abiotic NH\textsubscript{2}OH oxidation, recent advances in research on the isotopic composition of N\textsubscript{2}O from denitrification under oxic and anoxic conditions (Lewicka-Szczebak et al., 2015) and from fungal denitrification (Rohe et al., 2014) show a progress in this field by adding new fractionation factors and SP values for the differentiation between distinct conditions and microorganisms. Even if with this thesis no differentiation between biotic and abiotic processes was possible, new information about the behavior of fractionation over time could be gained using latest quantum cascade laser absorption spectroscopy technology that was not possible with older IRMS instruments.

With recent advances in technology, like infrared laser spectroscopy for SP measurements in N\textsubscript{2}O having become commercially available, more high-resolution data on different processes under a range of conditions could be gained in the near future, giving a deeper insight into the isotope fractionation by different processes and bringing new chances for a possible source partitioning. Another issue is the comparability of data from different laboratories because of the lack of international standards for $\delta^{15}$N\textsubscript{bulk} and SP in of N\textsubscript{2}O. However, although there still is no standard for SP, recent efforts in form of interlaboratory compression studies, such as by Mohn et al. (2014), will help to improve comparability of results between different laboratories.
6.3.2. Abiotic NH$_2$OH oxidation in soils

As suggested in chapter 4.4.2, abiotic N$_2$O formation could potentially be explained by three soil parameters: pH, C/N ratio, and Mn content. However, this study relies only on relatively few data points. For a proper prediction using soil parameters, a more substantial study would be needed. The problem of collinearity exists as well, so that studies on the influence of each single parameter are necessary to confirm the influence of the three identified soil parameters on the oxidation of NH$_2$OH in soils. It is possible that not all of the three parameters exert influence on abiotic N$_2$O formation. Although the influence of all three parameters can be explained chemically, especially the influence of the C/N ratio of SOM is unclear. Further studies with soils covering a broader range of C/N ratios and SOM composition are needed, including a characterization of the binding forms of N in the SOM.

Although this dissertation could show that at least some soils have the potential to oxidize the nitrification intermediate NH$_2$OH to form N$_2$O, the greatest uncertainty with respect to the coupled biotic–abiotic production of N$_2$O in soils is the release of NH$_2$OH by AOB or also AOA. While it has been shown in the past that different AOB strains released NH$_2$OH in cell cultures (Schmidt et al., 2004b; Stüven et al., 1992), this has not yet been reported from soils, although only recently it was possible to detect NH$_2$OH with a novel method in an acidic spruce forest soil (Liu et al., 2014). Further research on AOB and AOA and their release of NH$_2$OH is vital for a better understanding of biotic and abiotic N$_2$O formation, as this is the key step in the whole process. Without the release of NH$_2$OH there would potentially be no abiotic N$_2$O formation in soils. To determine the leakage rate of NH$_2$OH of AOB, AOA, or other soil microorganisms, will be a great effort for microbiologists because it would presumably be different for different bacterial strains and highly dependent on environmental conditions and stresses acting on the microorganisms, such as nutrient or oxygen availability. As it has been shown that AOA can outnumber AOB in most soils (Leininger et al., 2006), AOA could play an even more important role in this process than AOB; yet, little is known about their metabolism, that could be significantly different from that of AOB.

However, if an average leakage rate of NH$_2$OH from nitrification for different environments could be determined, this would allow – in conjunction with the knowledge about the basic soil parameters (as discussed above) – to estimate the abiotic N$_2$O production for single ecosystems based on the nitrification rate. Although there is still a lot of research on several unknown variables needed, this thesis shows that a prediction of abiotic N$_2$O formation based on nitrification rate and different soil parameters could be possible and should be the ultimate goal of studies on NH$_2$OH-induced N$_2$O formation, as it would greatly improve modeling of N$_2$O emissions from soil. Furthermore, mitigation strategies could be improved, as N$_2$O emissions might have been previously incorrectly connected to other production processes, so that better mitigation strategies could be designed aligned with the relevant processes.
6.3.3. N₂O decomposition over hot and dry surfaces

There should also be more research on the presented N₂O decomposition mechanism. Although this thesis could not confirm the existence of such a mechanism at conditions found in the environment, the failure to do so, however, could have been mainly due to the instrumentation and experimental setup. Recent instrumentation is already capable of detecting small changes in N₂O, but the setup needs to be improved to achieve a perfect gas-tightness of the system. New long-term experiments with such an improved gas-tightness might reveal the presence of this mechanism as a sink for N₂O in the troposphere, which could change our understanding of the atmospheric lifetime and cycling of N₂O dramatically.
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