Structure of Single-Chain Nanoparticles Under Crowding Conditions: A Random Phase Approximation Approach

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Abstract

The conformation of poly methyl methacrylate (PMMA)-based single-chain nanoparticles (SCNPs) and their corresponding linear precursors in presence of deuterated linear PMMA in deuterated dimethylformamide (DMF) solutions has been studied by small angle neutron scattering (SANS). The SANS profiles were analyzed in terms of a three-component random phase approximation (RPA) model. The RPA approach

described well the scattering profiles in dilute and in crowded solutions. Considering all the contributions of the RPA leads to an accurate estimation of the single chain form factor parameters and the Flory-Huggins interaction parameter between PMMA and DMF. The value of the latter in dilute regime indicates that the precursors and the SCNPs are in good solvent conditions, while in crowding conditions the polymer becomes less soluble.

Introduction

Macromolecules in biological systems in vivo are mostly in the cell's crowded environment, where the high concentration of other molecules creates an effective nano-confinement that can dramatically modify biological function through changes in the structural conformation as well as in the dynamics with respect to diluted in vitro conditions. In particular, unstructured polymer chains—including intrinsically disordered proteins and unfolded coils—compress under crowding. In fact, the coil collapse is considered a necessary factor that drives protein folding. Aspects such as concentration, chain architecture, intra- and intermolecular interactions will have an impact on the chain conformation. The understanding of the role of these factors requires the structural characterization in crowded media, which is challenging given the high concentration and the intrinsic complexity in these samples.

Neutron scattering is the most suited technique to measure the form factor of small concentrations of labeled chains in the presence of large concentrations of other species. It measures both the structure and thermodynamic quantities such as the Flory-Huggins interaction parameter. In dilute conditions, both small angle X-ray or neutron scattering (SAXS/SANS) access the overall chain size as well as the fractal dimension inside the particle. However, in the semi-dilute regime SANS and SAXS yield information about the chain correlations under full contrast conditions, i.e., when all the macromolecules have a different scattering length density compared to the solvent. To probe the single chain conformation in semi-dilute and concentrated solutions, the appropriate contrast conditions are needed.

This can be achieved for SANS by hydrogen/deuterium labeling. A strategy commonly used is masking out the contribution of one species in the mixture by contrast matching with the solvent. This is done by having a small amount of hydrogenated polymers in a mixture of deuterated polymers and deuterated solvent and assuming that the scattering signal arises exclusively from the hydrogenated polymer. However, to make sure that the scattering intensity yields the single chain form factor, the system must be in the so-called zero-average contrast conditions, which cancel out the intermolecular structure factor contribution to the total scattering.³

The chain size reduction of homopolymers in semi-dilute solutions is fairly well understood; theoretical approaches have led to the prediction that the radius of gyration scales with concentration as $R_{\rm g} \sim c^{-1/8}$ in good solvent and this dependence was experimentally supported by SANS in homopolymer solutions.^{3,4} The same behavior was found in solutions in which the probe and the crowder are chemically different.⁵ In binary polymer mixtures where the effects of the polymer-polymer interactions are not negligible, these must be taken into account to quantify the chain compression.⁶ Intrinsically disordered proteins (IDPs), as they are unfolded chains, are also expected to contract in crowding media. However, SANS studies of IDPs in contrast matched crowder solutions of diverse nature, from globular proteins to ramified polysaccharides, have yielded contradictory data. In some cases, a mild size reduction is reported,⁷ and in others even a size increase, attributed to a soft attraction with the crowders which are of different chemical nature than the protein under study.⁸

Indeed, biological systems are heterogeneous and complex in nature, and the use of simplified models helps to bridge the gap between polymer solutions and biological mixtures. In particular, single-chain nanoparticles (SCNPs) can help address the essential question of the effect of crowding on the structure. SCNPs are single-stranded polymers partially collapsed via intramolecular bonding. To avoid unwanted intermolecular bonding, collapse is induced under high dilution. These nano-objects serve as simplified models for IDPs⁹ due to their internal structure and inherent polydispersity in size and morphology. ¹⁰ General synthesis

methods involve the functionalization of a linear precursor and subsequent intramolecular cross-linking. ¹¹ Each cross-link between two reactive functional sites of the chain generates an internal loop and globally the precursor chain reduces its size. The formation of long-range loops in good solvent conditions is less probable and short range loops are favored due to the self-avoiding conformation of the precursor, resulting in sparse, non-globular SCNPs in solution. ¹² In addition, unlike globular proteins whose folding is driven by defined interactions, the collapse of synthetic SCNPs is the result of a stochastic process leading not only to a less controlled compaction but also to a polydispersity of resulting topologies. ^{12,13}

The structural changes of SCNPs under crowding have been previously addressed using model SCNPs embedded in analogue linear crowder solutions. ^{14,15} In one study, the effect of crowding of linear polystyrene (PS) of low- and high-molecular weights on the structure of PS-SCNPs showed that the crowders with different molecular weights have different effects: while long chains tend to impede their aggregation and lead to chain compression above their overlap concentration, short ones are found to mediate depletion interactions leading to aggregation. ¹⁵ On another study, the effect of poly(methyl methacrylate) (PMMA) on the structure of PMMA-based SCNPs was investigated by combining contrast variation SANS with molecular dynamics (MD) simulations. Results showed a crossover from unperturbed SCNP conformation in dilute conditions to a collapse starting when the total concentration reached the SCNP overlap concentration, and continuing decreasing in size with increasing crowder concentration. ¹⁴

In those studies, the SANS signal was assumed to result exclusively from the labeled chain form factor, assuming that any other contribution was removed by subtracting the scattering of the nearly contrast-matched crowder. However, in such concentrated systems, even mild polymer-solvent interactions contribute to the scattering. To account for such effects, the Random Phase Approximation (RPA) formalism applies, which considers the contribution of the effective Flory-Huggins interaction parameter between the different components in a polymer mixture to the total structure factor. This theoretical framework is

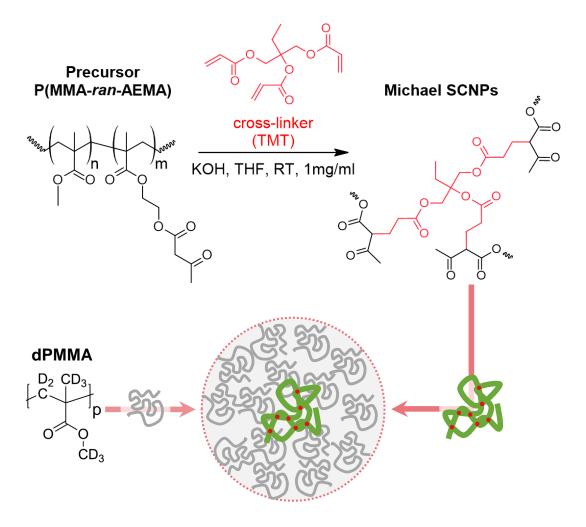
commonly invoked to analyze the structure ¹⁶ and dynamics ^{17,18} of polymer blends and, even though it works best for concentrated systems and melts, it has also been used to elucidate the structural and thermodynamical quantities of polymer solutions ^{19,20} and gels. ²¹ Here, we use a three-component RPA approach to analyze the SANS results on hydrogenated SCNPs in solutions of deuterated linear crowders and deuterated solvent. For this, we use PMMA-based SCNPs with 30% functional groups cross-linked with a trifunctional crosslinker. The crowder is deuterated PMMA in solution of deuterated dimethylformamide (DMF). We compare the results of considering the RPA with the ones obtained under the above mentioned simplification.

Materials and Methods

Sample preparation

We investigate SCNPs and the corresponding linear precursors (Prec) as reference. The precursors consist of random copolymers of methyl methacrylate (MMA) and (2-acetoacetoxy)ethyl methacrylate (AEMA), namely P(MMA_{0.69}-ran-AEMA_{0.31}), synthesized through reversible addition-fragmentation chain-transfer (RAFT) polymerization.²² The SCNPs were obtained through Michael addition of the trifunctional cross-linking agent trimethylolpropane triacrylate (TMT, 33 mol % to AEMA) (Sigma-Aldrich, technical grade) to β -ketoester functional groups of the precursors in a procedure described earlier²² (see Scheme 1). Two different molecular weights were investigated. Molecular weights and polydispersities of the samples (as determined by SEC/MALLS, see Supporting Information (SI) S.1) as well as other physico-chemical parameters are displayed in Table 1.

The neutron scattering experiments were carried out on solutions with deuterated N,N-dimethylformamide (dDMF, 99.5 atom %, Acros Organics) as solvent. Crowded solutions were obtained by adding deuterated linear PMMA chains of two different molecular weights (dPMMA, Polymer Source, see Table 1). After synthesis and purification, stock solutions



Scheme 1: Synthesis of random poly(methyl methacrylate-ran-(2-acetoacetoxy)ethyl methacrylate) single-chain nanoparticles. In the samples studied in this work, n=0.69 and m=0.31. Schematic illustration of the SCNPs under crowding with dPMMA chains in dDMF is included at the bottom.

of precursors and SCNPs were prepared, and appropriate quantities of dPMMA were added immediately to reach the desired total concentration: in the case of low- $M_{\rm w}$ probes, $c_{\rm Tot} = c_{\rm probe} + c_{\rm crowder} = 20 \, {\rm mg/mL} + 180 \, {\rm mg/mL} = 200 \, {\rm mg/mL}$, and in the high- $M_{\rm w}$ probes $c_{\rm Tot} = c_{\rm probe} + c_{\rm crowder} = 5 \, {\rm mg/mL} + 195 \, {\rm mg/mL} = 200 \, {\rm mg/mL}$. The concentration of the probes is below the overlap concentration, estimated as $c^* = M_{\rm w} / \left[(2R_{\rm g})^3 \, N_{\rm A} \right]$ for the two cases (see Table 1). In the crowded samples, the total concentration of polymer is above the overlap concentration of the probe. The neutron scattering length density ρ of dDMF is $6.36 \times 10^{10} \, {\rm cm}^{-2}$, close to that of dPMMA (see Table 1).

Table 1: Molecular characteristics of precursors, SCNPs, crowders, and solvent: Molecular weight $(M_{\rm w})$ and polydispersty index $(M_{\rm w}/M_{\rm n})$, mass density (d), scattering length density (ρ) , radius of gyration (R_g) , scaling exponent (ν) , and overlap concentration (c^*) .

		$M_{\rm w}{}^a$ (kg/mol)	$M_{ m w}/M_{ m n}{}^a$	d (g/cm ³)	$\rho (10^{10} \text{ cm}^{-2})$	$R_{\rm g}^{\ b}$ (nm)	$ u^b$	c^{*c} (mg/mL)
-		(1101)		(8/ СШ)	(10 cm)	(11111)		(1116/11112)
Probes	Lo-Prec	33.1	1.116	1.21	1.27	5.3	0.481	46
	Lo-SCNPs	33.9	1.043			4.3	0.389	90
	Hi-Prec	247	1.345			16.2	0.556	12
	Hi-SCNPs	239.2	1.31			14.5	0.484	16
Crowders	Lo-dPMMA	9.6	1.109	1.27	6.97	3.4	0.59	51
	Hi-dPMMA	99.1	1.093			10.9	0.59	16
Solvent	dDMF			1.03	6.36			

 $[^]a$ from SEC/MALLS in THF

SANS Measurements

SANS measurements were performed on the instrument KWS-2 at the Forschungs – Neutronenquelle Heinz Maier-Leibnitz (MLZ) in Garching, Germany. ²³ Measurements were carried out at room temperature using a neutron wavelength $\lambda = 5$ Å. For the low-molecular weight probes, two sample-detector distances were used (2 and 8 m) with 8 m collimation. For the bigger macromolecules, additional measurements were conducted at a distance of 19.9 m with 20 m collimation. Data from different detector positions were merged applying the same calibration factor for all the samples. Samples were contained in 2 mm thick quartz cuvettes (QS, Hellma). The sensitivity of the detector elements was accounted for by comparing to the scattering of a 1.0 mm sample of water, and a Plexiglas measurement was used for absolute scaling. Sample thickness, transmission, detector dead time, and electronic background were considered, and the background due to the scattering of the cell filled with deuterated solvent was subtracted from the sample measurements with H-labeled macromolecules for the RPA analysis. Finally, the azimuthally averaged scattered intensities were obtained as a function of the wave-vector magnitude, $Q = 4\pi \sin(\theta/2)/\lambda$, where θ is the scattering angle.

 $[^]b$ from SANS in dilute solutions c $c^* = M_{\rm w} / \left[\left(2 R_{\rm g} \right)^3 {\rm N_A} \right]$

In the data analyses the additional incoherent background arising mainly from the hydrogens in the polymers was fitted as a constant $I_{\rm inc}$ at high-Q and subtracted from the data such that $I(Q) = I_{\rm exp}(Q) - I_{\rm inc}$, where $I_{\rm exp}$ is the experimentally obtained data (see SI for further details). We note that the influence of the background in our case is not that critical due to the high coherent scattering intensity of the samples compared to the incoherent background and, not less important, due to the wide Q-range of the slope between $\pi/R_{\rm g}$ and the point where the curve flattens. This is even less important for the high-molecular weight probes where I(0) is higher and $\pi/R_{\rm g}$ is lower. The error bars for $Q < 0.3 \text{ Å}^{-1}$ are smaller than the size of the points (see Fig. S4).

SANS Analysis

For a binary system, such as a polymer in solution, the total measured differential coherent scattering cross section per unit sample volume, I(Q), depends on the contrast between the two components as

$$I(Q) = I_0 P(Q) S_{\rm I}(Q), \qquad (1)$$

where I_0 is the forward scattering $(Q \to 0 \text{ value})$, P(Q) is the form factor, and $S_{\rm I}(Q)$ is the structure factor accounting for intermolecular interactions between particles. The forward scattering can be written as $I_0 = \phi \Delta \rho^2 V$, where ϕ is the volume fraction of scatterers, $\Delta \rho$ is the scattering contrast given by the difference in scattering length density ρ between the components, $V = M_{\rm w}/d{\rm N_A}$ is the volume of the scatterer; with $M_{\rm w}$ being the weight average molecular weight, d the mass density and ${\rm N_A}$ the Avogadro number. Note that eq. 1 assumes monodisperse objects. Under high dilution conditions, interactions between different macromolecules are negligible, and the associated structure factor can be considered close to unity, $S_{\rm I}(Q) \approx 1$. Thus, the Q-dependence of the measured curve is determined just by the form factor of the particles in solution, P(Q).

For linear polymers and SCNPs in dilute solutions, 24 to take into account molecular

conformation including excluded volume effects a generalized Gaussian coil form factor $P(Q, R_{\rm g}, \nu)^{25}$ should be used:

$$P(Q, R_{\rm g}, \nu) = \frac{1}{\nu u^{1/2\nu}} \gamma\left(\frac{1}{2\nu}, u\right) - \frac{1}{\nu u^{1/\nu}} \gamma\left(\frac{1}{\nu}, u\right)$$
(2a)

with

$$u = Q^2 R_g^2 (2\nu + 1) (2\nu + 2) / 6,$$
 (2b)

$$\gamma(a,b) = \int_0^b t^{a-1} e^{-t} dt,$$
 (2c)

where $R_{\rm g}$ is the radius of gyration and ν is the scaling exponent. Fully swollen linear chains have a ν exponent of 3/5 (good solvent), Gaussian chains of 1/2 (linear chains in θ -solvent) while collapsed macromolecules of 1/3 (bad solvent). This form factor is normalized to 1 at low Q. The Debye function is recovered from Eq. 2 for $\nu = 0.5$. When the solvent-polymer interactions are strong, or in the semidilute and concentrated regime where intermolecular interactions are non-negligible, the scattered intensity is affected and formalisms such as the RPA must be used to account for such effects.

Random Phase Approximation for a Three-Component System

The Random Phase Approximation (RPA) theory has been used to describe the SANS data. In the following, we present the scattering function of a polymer solution consisting of a mixture of protonated and deuterated polymers in this framework. For incompressible mixtures, ^{26–28} the macroscopic scattering cross section is given by

$$I(Q) = \Delta \rho^{\mathrm{T}} \mathbf{S}(Q) \, \Delta \rho, \tag{3}$$

where, for a three-component system, $\mathbf{S}(Q)$ is a 2 × 2-matrix and $\Delta \rho$ is a 2-column vector for the scattering length density differences relative to the background (3rd component) and $\Delta \rho^{\mathrm{T}}$ is its transpose. The inverse structure matrix for this system can be written as

$$\mathbf{S}^{-1}(Q) = \mathbf{S_0}^{-1}(Q) + \mathbf{U}(Q). \tag{4}$$

 $\mathbf{S_0}(Q)$ is a matrix of non-interacting (bare) structure factors, which for homopolymer solutions is diagonal. The excluded volume interactions are contained in matrix \mathbf{U} , which can be expressed in terms of the bare structure factor for the background component $(S_{33}^0(Q))$ and the Flory-Huggins interaction parameters (χ_{ij}) :

$$U_{ii} = \frac{1}{S_{33}^{0}(Q)} - 2\frac{\chi_{i3}}{v_{0}} \quad \text{for } i = 1, 2$$

$$U_{ij} = \frac{1}{S_{33}^{0}(Q)} + \frac{\chi_{ij}}{v_{0}} - \frac{\chi_{i3}}{v_{0}} - \frac{\chi_{j3}}{v_{0}} \quad \text{for } i, j = 1, 2$$

$$(5)$$

where v_0 is a reference volume. Thus, for a three-component system consisting of two polymers and a solvent, being component 1 the probe polymer, component 2 the crowder, and component 3 the solvent, the scattered intensity is given as the summation of squares of scattering length density differences between polymer chains and solvent molecules, multiplied by the structure factors:

$$I(Q) = \Delta \rho_1^2 S_{11}(Q) + \Delta \rho_2^2 S_{22}(Q) + 2\Delta \rho_1 \Delta \rho_2 S_{12}(Q).$$
(6)

According to Eq. 4, the fully interacting system structure factors can be written as

$$S_{11}(Q) = \frac{S_{11}^{0}(Q)(1 + U_{22}S_{22}^{0}(Q))}{(1 + U_{11}S_{11}^{0}(Q))(1 + U_{22}S_{22}^{0}(Q)) - U_{12}^{2}S_{11}^{0}(Q)S_{22}^{0}(Q)},$$
(7a)

$$S_{22}(Q) = \frac{S_{22}^{0}(Q)(1 + U_{11}S_{11}^{0}(Q))}{(1 + U_{11}S_{11}^{0}(Q))(1 + U_{22}S_{22}^{0}(Q)) - U_{12}^{2}S_{11}^{0}(Q)S_{22}^{0}(Q)},$$
(7b)

$$S_{12}(Q) = \frac{-S_{11}^{0}(Q)U_{12}S_{22}^{0}(Q)}{(1 + U_{11}S_{11}^{0}(Q))(1 + U_{22}S_{22}^{0}(Q)) - U_{12}^{2}S_{11}^{0}(Q)S_{22}^{0}(Q)}.$$
 (7c)

The single-chain form factors for homopolymers can be expressed as

$$S_{ii}^{0}(Q) = N_i \phi_i v_i P_i(Q), \qquad (8)$$

where N_i is the degree of polymerization for the component i, ϕ_i its volume fraction, v_i the molar volume of a segment of the chain, and $P_i(Q)$ its form factor. For the solvent, $N_3 = 1$, $P_3(Q) = 1$, and thus, the bare structure factor of component 3 can be written as $S_{33}^0 = v_3(1 - \phi_p)$, with $\phi_p = \phi_1 + \phi_2$ the total volume fraction of polymer in the solution.

Equation 7 can be simplified under the assumptions that the probe and the crowder are compatible ($\chi_{12} = 0$). In our systems, this assumption is made because by experimental design the precursors and SCNPs have very similar chemical composition to the crowder. In addition, the interaction between both polymers and the solvent is the same ($\chi_{13} = \chi_{23} = \chi$), thus

$$U_{11} = U_{22} = U_{12} = \frac{1}{v_3} \left(\frac{1}{1 - \phi_p} - 2\chi \right), \tag{9}$$

where we have chosen v_3 as the reference volume.

In principle, the RPA was derived for polymers that exhibit Gaussian statistics, where P(Q) is described by the Debye scattering function, i. e., the conformation linear polymers adopt in θ -solvent conditions and in bulk. However, in polymers in other solvent conditions or

macromolecular conformation leading to a different scaling exponent, a generalized Gaussian coil form factor (Eq. 2) has to be used. This form factor yielded reasonable results for the analysis of SANS experiments in polymer solutions using the RPA approach. ^{19,20}

Equation 6 could be further simplified under contrast-matching conditions between the crowder and the solvent ($\Delta \rho_2 = 0$), leading to

$$I(Q) = \Delta \rho_1^2 S_{11}(Q)$$
. (10)

We want to emphasize that contrast-matching is different to zero-average contrast (ZAC) conditions, fulfilled for a system where the two polymers have the same degree of polymerization $(N_1 = N_2)$, equal form factor $(P_1(Q) = P_2(Q))$, and the same thermodynamic properties with respect to the solvent, i.e., the quality of the solvent is the same for both polymers $(\chi_{13} = \chi_{23} = \chi)$, the following condition is satisfied

$$\Delta \rho_1 \phi_1 + \Delta \rho_2 \phi_2 = 0, \tag{11}$$

and the interaction between both polymers is zero ($\chi_{12}=0$). Under ZAC conditions the equation $I(Q)=(\rho_1-\rho_2)^2\frac{\phi_1\phi_2}{\phi_p^2}P(Q)$ is obtained, ²⁹ and the single-chain form factor, which contains information on the intramolecular correlations, is accessible.

Lastly, if the Flory-Huggins interaction parameter fulfills the condition

$$\chi_{\theta} = \frac{1}{2\left(1 - \phi_{\mathbf{p}}\right)},\tag{12}$$

the excluded volume interactions are zero $(U_{11} = U_{22} = U_{12} = 0)$ and the system is under θ solvent conditions. Under these circumstances, the fully interacting system structure factors $S_{11}(Q)$ and $S_{22}(Q)$ are equal to the corresponding single-chain form factors (Eq. 8), while

 $S_{12}(Q) = 0$ (see Eq. 7). Under that condition Eq. 6 can be written as

$$I(Q) = \Delta \rho_1^2 S_{11}^0(Q) + \Delta \rho_2^2 S_{22}^0(Q).$$
(13)

If the crowder is contrast-matched with the solvent, $\Delta \rho_2 = 0$ and Eq. 1 is recovered. Equation 12 marks the threshold between good solvent and poor solvent conditions: if χ is lower than $\frac{1}{2(1-\phi_p)}$, the excluded volume interaction parameter U is positive (see Eq. 9) and the system is in good solvent conditions. On the contrary, if $\chi > \frac{1}{2(1-\phi_p)}$, U is negative and the system is in poor solvent conditions.

Results and discussion

To study the effect of the crowding environment on the structure of both precursors and their SCNPs, we used SANS, where the scattering intensity of the protonated precursors and SCNPs is highlighted due to the low contrast between the deuterated crowders and the deuterated solvent (Table 1). We considered two different molecular weights for the protonated precursors; low- $M_{\rm w}=33.1$ kDa and high- $M_{\rm w}=247$ kDa. For both macromolecules, we kept their concentration in all the solutions below their own overlap concentration c^* , i.e., 20 mg/mL for low- $M_{\rm w}$ probes and 5 mg/mL for high- $M_{\rm w}$ probes. In the crowded samples, the total concentration was kept to 200 mg/mL. Crowded conditions were induced using crowders with two molecular weights.

To illustrate the need of using the RPA approach in these cases, we consider the SANS curves of the linear precursors in dilute solutions and in the presence of crowders. Figure 1 shows the SANS results obtained for the low- and high-molecular weight precursors in the dilute regime and under crowding conditions. In the case of the concentrated solutions, the contribution of the crowders to the scattered intensity has been taken into account by subtracting the signal from solutions of the deuterated crowder in deuterated solvent, as it was considered in the previous work¹⁴ (see SI for details on background subtraction).

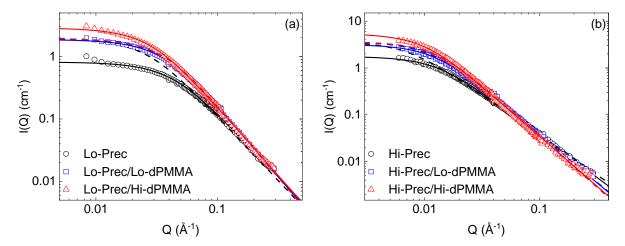


Figure 1: Scattered intensities I(Q) of (a) low-molecular weight and (b) high-molecular weight precursors in dilute (black circles) and under crowding conditions with low-molecular weight crowders (blue squares) and high-molecular weight crowders (red triangles). Lines are fits using Eq. 1 with a generalized random coil form factor (Eq. 2) with forward scattering I_0 for Q = 0 as a fitting parameter (solid lines), and with the forward scattering fixed to that defined by the scattering length density and the molecular parameters, as in Eq. 1 (dashed lines).

In all cases, the shape of the scattering curves corresponds to that of a polymer coil with a Guinier region at low-Q and a $Q^{-1/\nu}$ power law at intermediate-Q related to the chain fractal dimension. In both systems, upon crowding there is an increase in the slope, which is indicative of more compact objects. In addition, it can be seen that the forward scattering intensity increases in crowding conditions. Generally, an increase in forward scattering in binary polymer/solvent systems can be due either to molecular weight increase, caused by chemical intermolecular crosslinking of unreacted functional groups, or due to aggregation, caused by attractive physical interactions between the polymers or bad solvent conditions.

Recently, our group studied aggregation effects in PS-based SCNPs under crowding in molecular weight symmetric and asymmetric systems. ¹⁵ There, it was observed that low-molecular weight crowders added to high- $M_{\rm w}$ SCNPs solutions produce depletion interactions, destabilizing the suspension and causing chemical aggregation mediated by unbounded reactive groups. This is not the case for the samples investigated in this work, since the same intensity increase at low-Q values can be observed when adding crowders to the solutions of

both, precursors and SCNPs (see Figures S2 and S3), irrespective of the presence of reactive groups, and it happens in both symmetric and asymmetric polymer mixtures. Therefore, the observed changes in forward scattering don't appear to be related to aggregation.

Furthermore, according to Eq. 1, in absence of interactions, i.e., $S_{\rm I}(Q) \simeq 1$, the value of the forward intensity should be given by the molecular volume as well as its concentration and contrast (see Table 1). However, when using Eq. 1 to describe the data (Figure 1, solid lines), in the case of dilute samples without crowder, the forward scattering value I_0 given by the fitting is lower than that determined by the molecular parameters of the studied systems, $\Delta \rho^2 \phi V$ (see Figure 1, dashed lines). The fitting parameters are listed in Table S2. This mismatch in the intensity at low-Q values in the dilute samples without crowding can be attributed to the presence of repulsive interactions between the probe chains. In fact, it was observed in a previous study 30 that, at the concentration of 20 mg/mL used for the low molecular weight precursor and SCNP system, the SCNPs in solution display non-negligible intermolecular interactions leading to a structure factor which deviates from unity. When adding crowders to the solutions, these repulsive interactions are screened due to the presence of the dPMMA chains, leading to an increase in the forward intensity value. We note that the high molecular weight crowder leads to a higher intensity increase. In addition, the forward scattering does not correlate with the molecular weight asymmetry in the polymer mixture, ruling out depletion interactions. Therefore, polymer-solvent interactions must be considered in the overall structure factor.

The scattered intensity can be described following the RPA approach for three components (probe, crowder, and solvent) using Eqs. 6, 7 and 8. In this case, note that the background subtracted corresponds to the deuterated solvent (without crowders) and the incoherent contribution (see S.I., Section S. 4). Assuming $\chi_{12} = 0$ and $\chi_{13} = \chi_{23} = \chi$ (see Eq. 9) there are a total of 5 variable parameters: the radii of gyration $(R_{g,1}, R_{g,2})$ and the scaling exponents (ν_1, ν_2) of both polymers, and the Flory-Huggins interaction parameter (χ) . The rest of the parameters can be fixed according to the sample composition and molec-

Table 2: Fixed parameters for RPA curve fitting using Eqs. 6, 7 and 8: Degree of polymerization (N_i) , volume fraction (ϕ_i) , and molar volume (v_i) . $\phi_p = \phi_1 + \phi_2$ is the total volume fraction of polymer in the solution.

	F	Probe	Cro	Solvent	
Degree of polymerization	Low- $M_{ m w}$	$N_1 = 244$	Lo-dPMMA	$N_2 = 90$	$N_3 = 1$
	$\mathrm{Hi}\text{-}M_{\mathrm{w}}$	$N_1 = 1821$	Hi-dPMMA	$N_2 = 920$	
Polymer volume fraction	Low- $M_{ m w}$	$\phi_1 = 0.017$		$\phi_2 = 0.141$	$\phi_3 = 1 - \phi_{\rm p}$
	$\mathrm{Hi}\text{-}M_{\mathrm{w}}$	$\phi_1 = 0.004$		$\phi_2 = 0.154$	= 0.842
Molar volume		$v_1=187~\mathrm{\AA}^3$		$v_2=141~\mathrm{\AA^3}$	$v_3=129~\mathrm{\AA^3}$

ular parameters listed in Table 2. Here, we consider the reference unit for our precursors or SCNPs what we call "effective" monomer, whose properties are the result of averaging over the copolymer components. Thus, the molar mass m_0 of the effective monomer is obtained as $0.69 \times m_{0,\text{MMA}} + 0.31 \times m_{0,\text{AEMA}} = 135.6 \text{ g/mol}$. From this value, we obtained the degree of polymerization and the molar volume of the probes.

As the single-chain form factors in Eq. 7 are strongly correlated, it is hard to treat the radii of gyration and the scaling exponents as independent fitting parameters.³¹ Thus, the values of $R_{\rm g}$ and ν of the crowders were estimated and fixed during the fitting procedure, leaving only three free fitting parameters in the model. The estimation of $R_{\rm g}$ and ν for the crowders was made in the following way: first, from SAXS and SANS measurements, these parameters were determined in dilute conditions; from them, the values corresponding to high concentration were estimated according to computer simulations performed on linear polymers in the semi-dilute regime¹⁰ (see SI). These values, along with the final values of the fit parameters for SCNPs and precursors, are listed in Table 3 for a better comparison.

Figure 2 displays the scattered intensities obtained from SANS measurements for the low- $M_{\rm w}$ system along with the description obtained using RPA formalism described above. The equivalent graphs for the high-molecular weight precursor and SCNPs are shown in Figure 3. The fits describe the experimental results equally well as letting the forward scattering free and neglecting any other contributions, yet here the molecular weights, concentrations and scattering length densities are fixed and the only free parameters are $R_{\rm g}$, ν and χ , which

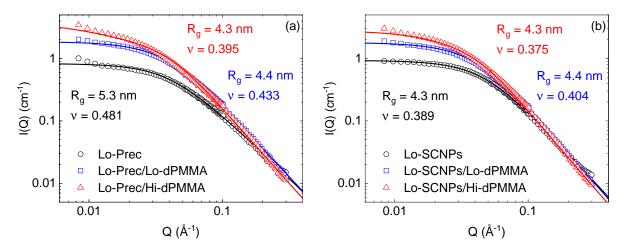


Figure 2: Scattered intensities I(Q) of low-molecular weight (a) precursors and (b) SCNPs in dilute ($c_{\text{Tot}} = 20 \text{ mg/mL}$, black circles) and under crowding conditions ($c_{\text{Tot}} = c_{\text{probe}} + c_{\text{crowder}} = 20 \text{ mg/mL} + 180 \text{ mg/mL}$) with low-molecular weight crowder (blue squares), and high-molecular weight crowder (red triangles). Solid lines are RPA fits with parameters given in the legends and in Table 3.

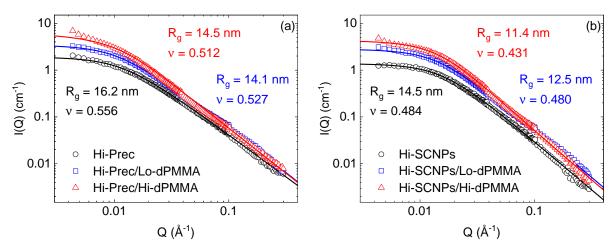


Figure 3: Scattered intensities I(Q) of high-molecular weight (a) precursors and (b) SCNPs in dilute ($c_{\text{Tot}} = 5 \text{ mg/mL}$, blue circles) and under crowding conditions ($c_{\text{Tot}} = c_{\text{probe}} + c_{\text{crowder}} = 5 \text{ mg/ml} + 195 \text{ mg/mL}$) with low-molecular weight crowder (blue squares), and high-molecular weight crowder (red triangles). Solid lines are RPA fits with parameters given in the legends and in Table 3.

are listed in Table 3.

To visualize the contribution of each of the terms in Eq. 7 to the total scattered intensity described by the three-component RPA formalism (Eq. 6), we have selected the case of

the Lo-SCNPs in crowding conditions with low- and high-molecular weight dPMMA. The contribution of each one of the components in Eq. 7 multiplied by the corresponding contrast factor is represented in Fig. 4 for the Lo-SCNPs with the Lo-dPMMA crowder, and in Fig. 5 for the Lo-SCNPs with the Hi-dPMMA crowder. If the interaction parameter χ is lower than $\chi_{\theta} = \frac{1}{2(1-\phi_{\rm p})}$, the excluded volume interaction parameter U is positive and the system is in good solvent conditions. Also, S_{12} is negative and the third (cross) term in Eq. 6 is positive ($\Delta \rho_1 < 1$). This is the case represented in Fig. 4, where $\chi = 0.450 < \chi_{\theta}$. The cross-term $\Delta \rho_1 \Delta \rho_2 S_{12}$ contributes more than the second term in Eq. 6, which reflects the contrast between the crowder and the solvent. On the contrary, when $\chi > \frac{1}{2(1-\phi_{\rm p})}$, the interaction parameter U is negative, meaning that the self-attractions become important and the polymer becomes less soluble. In that case, $\Delta \rho_1 \Delta \rho_2 S_{12}$ is negative. That is the case represented in Fig. 5, where $\chi = 0.595 > \chi_{\theta}$, and the term $\Delta \rho_2^2 S_{22}$ contributes substantially to the overall curve.

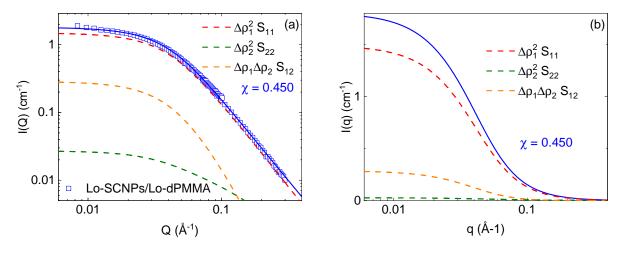


Figure 4: Scattered intensities I(Q) of low-molecular weight SCNPs in crowding with LodPMMA with RPA fits (solid lines). Each contribution on Eq. 6 is represented in a different color (dashed lines, see legend). Data in (a) are represented in logarithmic scale. In (b), the fitting function together with the components is shown in linear scale.

Thus, in our systems, even though the scattering length density of the crowder is very close to that of the solvent and $\Delta \rho_1$ is much larger than $\Delta \rho_2$, the contributions of the crowder scattering as well as the cross-term are non-negligible, and they are more relevant

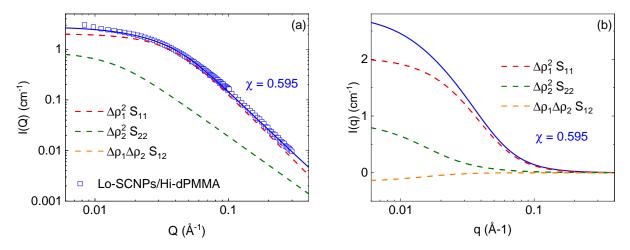


Figure 5: Scattered intensities I(Q) of low-molecular weight SCNPs in crowding with HidPMMA with RPA fits (solid lines). Each contribution on Eq. 6 is represented in a different color (dashed lines, see legend). Data in (a) are represented in logarithmic scale. In (b), the fitting function together with the components is shown in linear scale.

Table 3: Fitting parameters for RPA curve fitting using Eqs. 6, 7 and 8.

		Precursors			SCNPs			$\operatorname{Crowders}^*$	
		$R_{\rm g}~({\rm nm}~\pm 0.1)$	$\nu~(\pm 0.005)$	$\chi~(\pm 0.002)$	$R_{\rm g}~({\rm nm}~\pm 0.1)$	$\nu~(\pm 0.005)$	$\chi~(\pm 0.002)$	$R_{\rm g}~({\rm nm})$	ν
$\text{Low-}M_{\text{w}}$	Dilute	5.3	0.481	0.391	4.3	0.389	0.405		
	with Lo-dPMMA	4.4	0.433	0.545	4.4	0.404	0.450	3.1	0.550
	with Hi-dPMMA	4.3	0.395	0.596	4.3	0.375	0.595	9.3	0.530
$\mathrm{Hi}\text{-}M_{\mathrm{w}}$	Dilute	16.2	0.556	0.462	14.5	0.484	0.428		
	with Lo-dPMMA	14.1	0.527	0.590	12.5	0.480	0.546	3.1	0.550
	with Hi-dPMMA	14.5	0.512	0.597	11.4	0.431	0.594	9.3	0.530

^{*} fixed according to SAXS/SANS measurements and computer simulations. ¹⁰

in the samples crowded with high-molecular weight dPMMA. We note that neglecting the S_{22} and the cross-term contributions would not describe the observed intensities.

Analyzing the probe form factor parameters ($R_{\rm g}$ and ν) obtained from the RPA fits we make several observations. First, for both molecular weights, the SCNP formation via intramolecular cross-linking leads to a reduction of the radius of gyration as well as the scaling exponent. The relative size reduction upon SCNP formation is more pronounced in the case of the higher-molecular weight system, as expected from theory and observed experimentally.^{32,33} On the other hand, for the crowder concentration here investigated, the presence of the crowder produces a significant size reduction on the high-molecular weight probes, while has only a very small effect on the low-molecular weight probes. These findings are qualitatively in agreement with the conclusions from the previous study, ¹⁴ where the overlap concentration of the SCNP was identified as the cross-over concentration above which the macromolecule starts shrinking.

The interaction parameter χ varies with sample composition (see Table 3). In principle, the Flory-Huggins parameter is related to the solvent quality. Lower values than the threshold value given in Eq. 12 indicate good solvent conditions and higher values, poor solvency. In the dilute regime ($\phi_p \ll 1$), the threshold is at $\chi_{\theta} = 0.5$. Our results show that in the dilute regime precursors and SCNPs are in good solvent conditions. However, for the samples in crowded conditions the value of the interaction parameter is close to the threshold value (at the concentration here investigated, $\chi_{\theta} \approx 0.594$). In particular, when crowding is induced with Hi-dPMMA, $\chi > \chi_{\theta}$. These results suggest that for PMMA in semi-dilute solutions DMF is a worse solvent. We note that the values found here are close to the ones reported for PMMA in DMF semi-dilute solutions $\chi_{\text{PMMA/DMF}} \sim 0.56$.³⁴

Conclusions

The conformation of PMMA-based SCNPs and their corresponding precursors in dilute and under crowding conditions with linear PMMA chains has been investigated by SANS varying the molecular weight of the probes and the crowders. In spite of using deuterated crowders in deuterated solvent, the forward scattering intensity increases in crowding conditions, with high molecular weight crowder leading to a higher increase. Thus, the scattered intensity was analyzed in terms of a RPA model for the three components (probe, crowder and solvent) to consider the polymer-solvent interactions and cross-correlations.

The form factor parameters $R_{\rm g}$ and ν of the precursors and the SCNPs, as well as the Flory-Huggins interaction parameter between the PMMA and the solvent, were obtained in dilute and in crowded conditions. The presence of the crowder produces a size reduction

on the probes. For the crowder concentration here investigated, the size reduction is more pronounced in the high-molecular weight probes. Applying the three-component RPA model leads to a more accurate estimation of the form factor parameters since it considers all the contributions to the scattering. Corrections of about 30% in the value of $R_{\rm g}$ and about 10% in the scaling exponent ν are obtained with respect to the values estimated neglecting interactions and cross-correlations. We note that, even considering these corrections, the trends reported in previous works on similar systems are reproduced. The RPA model allows determining the Flory-Huggins interaction parameter, that varies with the sample composition indicating that in the dilute regime DMF is a good solvent for PMMA while in crowded conditions the polymer becomes less soluble.

Studies on SCNP structure in crowded media as that here presented can be considered as a basis for understanding the structural conformation of important and ubiquitous bio-macromolecules as they are intrinsically disorder proteins and unfolded coils in dense environments—the natural state *in vivo* conditions.

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Supporting Information Available

Experimental details of the Size-exclusion or Gel permeation chromatography (SEC/GPC) measurements. SAXS/SANS characterization of the PMMA/dPMMA crowders. Analysis of the SANS data in terms of generalized Gaussian coil. Details on background subtraction.

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