

Adapting crop production to climate change and air pollution at different scales

Evgenios Agathokleous^{1,2}, Michael Frei³, Oliver M. Knopf⁴, Onno Muller⁴, Yansen Xu^{1,2}, Thuy Huu Nguyen⁵, Thomas Gaiser⁵, Xiaoyu Liu⁶, Bing Liu⁷, Costas J Saitanis⁸, Bo Shang^{1,2}, Muhammad Shahedul Alam³, Yanru Feng³, Frank Ewert⁵, Zhaozhong Feng^{1,2*}

¹Collaborative Innovation Center on Forecast and Evaluation of Meteorological Disasters (CIC-FEMD), Nanjing University of Information Science & Technology, Nanjing 210044, Jiangsu, People's Republic of China.

² Key Laboratory of Ecosystem Carbon Source and Sink, China Meteorological Administration (ECSS-CMA), School of Applied Meteorology, Nanjing University of Information Science and Technology, Nanjing 210044, Jiangsu, People's Republic of China.

³Department of Agronomy and Crop Physiology, Institute for Agronomy and Plant Breeding, Justus-Liebig University Giessen, 35390, Giessen, Germany

⁴Institute of Bio- and Geoscience 2: plant sciences (IBG-2), Forschungszentrum Jülich GmbH, Jülich, 52425, Germany.

⁵University of Bonn/ INRES, Katzenburgweg 5, 53115 Bonn, Germany.

⁶Institute of Resource, Ecosystem and Environment of Agriculture, Nanjing Agricultural University, 1 Weigang, Nanjing 210095, People's Republic of China.

⁷National Engineering and Technology Center for Information Agriculture, Engineering Research Center of Smart Agriculture, Ministry of Education, Key Laboratory for Crop System Analysis and Decision Making, Ministry of Agriculture, Jiangsu Key Laboratory for Information Agriculture, Jiangsu Collaborative Innovation Center for Modern Crop Production, Nanjing Agricultural University, Nanjing, Jiangsu 210095, PR China.

⁸Lab of Ecology and Environmental Science, Agricultural University of Athens, Iera Odos 75, Athens 11855, Greece.

Editor's summary

Elevated CO₂, global warming, ozone pollution, and drought are major climate-related environmental challenges affecting field crop production. This paper discusses strategies and perspectives for crop production under climate change and air pollution at plant, field, and ecosystems scales.

Abstract: Air pollution and climate change are tightly inter-connected and jointly affect field crop production and agroecosystems health. Although the individual and combined impacts of air pollution and climate change factors are increasingly understood, adaptation of crop production to concurrent air pollution and climate change remains challenging. Here, we review the recent advances on the adaptation of crop production to climate change and air pollution at plant, field, and ecosystems scales. Main approaches at plant level include integration of genetic variation, molecular breeding, and phenotyping. Field level techniques include optimizing cultivation practices, promoting mixed cropping and diversification, applying technologies such as antiozonants, nanotechnology and -robot-assisted farming. Plant and field level techniques would be further facilitated by enhancing soil resilience, incorporating precision agriculture, and modifying the hydrology and microclimate of agricultural landscapes at the ecosystem level. Strategies and perspectives for crop production under climate change and air pollution are discussed.

Challenges of crop production under air pollution and climate changes

Since the introduction of semi-dwarf and disease resistant crop varieties during the green revolution [1], the yield potential of new cultivars in large regions has stagnated due to counteracting effects of climate change [2]. Such effects will likely continue impacting our cropping systems [3] and anthropogenic greenhouse gas emissions continue to rise [4]. Therefore, it is essential that methods of crop production are constantly revised and adapted according to the latest policies and contemporary environmental challenges.

Tackling climate change and air pollution is considered as one of the most urgent global tasks and affects sustainable development goals [5]. Elevated CO₂, global warming, ozone pollution, and drought are major climate-related environmental challenges [6]. Water deficit stresses cause stomata closure, reducing the ozone uptake and its damaging effect on photosynthesis in some cases [7,8]. Elevated CO₂ lowers the concentrations of several nutrients and vitamins, and threatens human dietary needs, such as B vitamins that are reduced by 17-30% in rice [9,10]. However, elevated CO₂ increases photosynthesis and crop growth, and can offset the negative impacts of heat stress [11]. Increasing CO₂ might also alleviate ozone impact due to smaller ozone uptake [12]. The positive effect of CO₂ on crop yields might be reduced depending on local microclimate conditions. Higher CO₂ concentrations in the surrounding air lead to enhanced intracellular CO₂ saturation, decreasing transpiration, and increasing internal water use efficiency [13]. At the regional and global scale, crop production is affected by these climate factors and their interactions with anthropogenic activities. For instance, the complex distribution of different cropping systems and landscape components, as well as their interactions between land and atmosphere strongly affect the climatic condition, i.e. drought [14], warming [15], and ozone concentration [16]. Moreover, the interactions between multiple stressors are highly complex and can lead to dynamic and non-linear impacts, including antagonism, additivity, and synergism [8]. Thus, adaptation strategies under climate change must incorporate positive or negative antagonistic, additive, or synergistic effects of the multiple environmental factors on crop physiological characteristics and crop production. Interactive effects of these stressors on crops might differ across spatial scales. Thus, understanding these interactions (additive, multiplicative, or compensating at different levels) is needed to provide suitable adaptive strategies. However, recommending adaptation options in responses to stressors remains challenging because those abiotic stress factors naturally occur together.

Here, we discuss potential solutions to adapt the crop production to climate change and air pollution at plant, field, and ecosystems scales (Fig. 1). We focus on adaptation to ozone, CO₂, and warming; however, other contemporary stressors are also considered in the discussion where adaptation strategies may offer additional benefits against such stressors.

Adaptation options at plant scale

Breeding approaches

Intraspecific genetic variation in resistance to climate change exists in different crops, which provides the basis for crop breeding [17,18]. Molecular breeding approaches such as marker-assisted selection (MAS) constitute a major leap compared to traditional and time-consuming breeding approaches such as pedigree selection [17,19]. Germplasm collections, such as the fully sequenced plant material available from the 3000 Rice Genomes Project [20] form a powerful resource, as they can be used for phenotypic screening of populations representing the entire genetic diversity of crop species in specific stress environments. Subsequently, the genetic architecture underlying favorable adaptive traits can be readily dissected with the aid of the available genomic sequence information using approaches such as genome-wide association study (GWAS). As a prerequisite, free access to the gene banks curating germplasm collections, as well as sustainable funding and benefit sharing schemes for gene banks need to be ascertained by agreements such as the International Treaty on Plant Genetic Resources for Food and Agriculture (<https://www.fao.org/plant-treaty/en/>). Researchers can then acquire the material to identify adaptive loci for specific environmental stress conditions that will then be introgressed into elite varieties using MAS. Regarding heat stress, ozone stress, and elevated CO₂, quantitative trait loci (QTL) and candidate genes were identified in major crops, which form a prerequisite for MAS breeding [17,18,21]. However, large-scale marker-assisted breeding programs for resistance to ozone and effective utilization of elevated CO₂ are missing. Conversely, heat-tolerance rice breeding is ongoing at the International Rice Research Institute (IRRI) [22], which is expected to add new discoveries in the near future.

Dissecting and integrating genetic variations for adaptation

Recent research programs now offer new insights for ozone. QTL related to ozone resistance were identified in rice [23], maize [24], and wheat [25]. Transcriptomic responses of crops such as common bean, garden pea and soybean to ozone were explored [26,27]. These studies demonstrate that ozone tolerance in crops is a quantitative trait. A classical pedigree selection approach in different field locations might not lead to ozone tolerance due to the unpredictability and heterogeneity of ozone occurrence, and therefore marker-assisted breeding might be the most promising approach to introgress known ozone tolerant loci into established crop varieties.

Carbon source-sink relationships play a significant role in determining the plant ability to utilize elevated CO₂ that increases photosynthesis [28]. QTLs for source- and sink-related traits, including flag leaf traits, yield traits, and photosynthetic pigments under ambient and elevated CO₂ were identified in rice [29–31]. In legumes, physiological traits related to elevated CO₂ were dissected [32]. These known loci can be utilized in breeding programs toward efficient CO₂ utilization. In addition, classical breeding approaches such as pedigree selection take place under gradually rising CO₂ concentration and might thereby unintentionally lead to adaptation and high yields under elevated CO₂ levels.

Improved crop yields due to CO₂ ‘fertilization’ may be limited by CO₂-induced reductions in the maximum carboxylation rate and the maximum rates of electron transport [33]. Therefore, improving biochemical limitations to photosynthesis offers an opportunity to enhance crop productivity. Rubisco activity is also linked to chloroplast CO₂ concentration [34]. Improving mesophyll conductance can enhance CO₂ diffusion from sub-stomatal cavities to chloroplasts, as a target of increasing chloroplast CO₂ concentration and Rubisco activity [35]. Improving the speed of stomatal opening would be beneficial to enhance crop yields under elevated CO₂. Concurrently, variation in ozone sensitivity among plants is related to stomatal conductance [36], and reduction in stomatal conductance and ozone stomatal flux may serve as targets for climate

change adaptation. However, stomata optimize CO₂ uptake while minimizing water loss [37]. Hence, breeding for ozone-tolerant cultivars with lower stomatal conductance may result in suppressed photosynthesis and yield. The trade-off between photosynthesis and stomatal conductance to improve leaf water use efficiency is linked to plant ozone tolerance. Accelerating the response of stomata to light is an efficient approach to improve water use efficiency, which is used to breed drought tolerant cultivars [38,39]. Optimized stomatal behavior to control the trade-off between photosynthesis and ozone flux is also an avoidance of elevated ozone [40]. Ozone tolerance is often related to the concentration of ascorbic acid in the apoplast and leaf tissue in crops such as wheat, soybean, and maize [27,41,42]. Therefore, regulating antioxidant levels could improve ozone tolerance [43]. Increases in leaf cell wall thickness, apoplastic pH, and leaf tissue ascorbic acid concentration also result in greater ozone detoxification by apoplastic ascorbate [44], which may provide an efficient approach to breed ozone-tolerant cultivars. However, manipulation of the leaf apoplast toward a more reduced redox state could have the unintended consequence of altering the plant ability to sense and respond to pathogen infection [45,46]. Hence, a more holistic approach is needed based on biological interactions. Moreover, adaptation options should allocate equal focus between yields and nutritional quality, especially considering the current threat of malnutrition in undeveloped and developing countries.

Crop heat stress tolerance is also complex, and threshold temperatures are species-specific [47]. Multiple loci for heat tolerance were identified in rice at the seedling stage [48,49] and reproductive stage [50–54]. In addition, 35 heat stress-responsive meta-QTLs and 45 candidate genes were identified in rice, facilitating marker-assisted heat-tolerant rice breeding [55]. Heat-responsive loci were also identified in wheat [56–58], maize [59–61], soybean [62], and lentil [63]. These advances facilitate the formulation of targets in developing long-term plans for mitigating current and future global warming impacts [64], especially for tackling the challenge of superior plants under multiple combined stresses.

Breeding crops for a future climate needs to address favorable and unfavorable interactions of CO₂ with increasing ozone and temperature [32]. However, it is challenging to maintain a breeding environment in which all of these environmental factors are stably maintained for a classical selection approach. A solution to this problem could be physiological breeding [65], in which physiological traits that confer advantages in these environmental conditions are introgressed into target varieties. The International Maize and Wheat Improvement Center (CIMMYT) reported the consideration of physiological traits, such as canopy temperature or the normalized difference vegetation index (NDVI), in breeding programs leads to wheat varieties with enhanced performance in heat and drought environments [65]. Such data are also commonly collected at field scale (see also section ‘phenotyping-robot-assisted farming’), further indicating the nexus between plant and field levels. In addition, as stress tolerance is a quantitative trait, many loci should be combined in a process called QTL pyramiding in order to adapt crop varieties to multiple stress conditions [66]. Although this is still an elaborate process, the availability of genome sequences for all major crops has greatly facilitated the fast identification of genetic markers for such MAS schemes [66,67]. One possible challenge that needs to be overcome in QTL pyramiding is ‘linkage drag’ of agronomically unfavorable traits that could be closely associated with favorable adaptive traits. Another challenge could be confounding or even conflicting physiological effects of QTL, for example in the case of stomatal regulation. Precise crop engineering based on mechanistic understanding of adaptive traits may help to overcome these challenges [68].

Adaptation options at field scale

Cultivation techniques for air pollutants and climate changes

There has been limited advance regarding how cultivation techniques may mediate air pollution impacts. Because elevated ozone accelerates leaf senescence and suppresses the photosynthetic rate of flag leaves during later developmental stages [69], early sowing of winter wheat helps to effectively reduce the ozone stomatal flux. On the other hand, the occurrence of ozone peak levels depends on the weather conditions, and sowing timing should be adjusted according to local conditions. Crop management practices such as changing irrigation facilitate improved crop tolerance to ozone. However, such a practice should also account for CO₂ emissions, e.g. decreased water supply may decrease CO₂ emission [70]. Adjustment of water supply would reduce water use and increase water use efficiency, thereby reducing ozone uptake flux. Preliminary findings suggest straw addition may offer benefits to crops under ozone stress [71]. Straw and other organic amendments to soil can also decrease soil CO₂ emissions, potentially offering double benefits [72,73]. Potential benefits in crop production of using crop management practices should be assessed using modeling frameworks and field experiments with multiple factors [74].

Ozone alters soil microbial communities and root activity [75,76], thereby impacting crop N uptake and grain protein accumulation [77]. There are no consistent results on the combined effects of ozone and N on crops [78,79]. N fertilizer aggravates, slows down or does not alter the effects of ozone on crops [78,80]. Ozone can also increase the optimum N application rates, decrease crop N use efficiency, and increase the risk of N-related environmental problems, such as soil N leaching, aquatic eutrophication, and greenhouse gas N₂O emissions [80]. Therefore, the time, frequency, dose and type of fertilization need to be further optimized to improve the N uptake capacity and antioxidant capacity of crops to cope with ozone stress. Fertilization regimes should also consider the influence of CO₂ effects. For example, the negative effects of low soil N on grain quality may be exacerbated by elevated CO₂ [81]. Concurrently, some agronomic measures are applied to increase the N utilization efficiency and reduce environmental pollution, such as combined application of organic and inorganic fertilizers [82]. Further research is needed to investigate whether these agronomic measures are effective in mitigating ozone damage to crops, especially with concurrent elevated CO₂ and warming [83].

Optimization of agricultural land management may act against global warming. For example, under the Representative Concentration Pathway 2.6, current agricultural mitigation practices to increase soil organic carbon sequestration may decrease global temperature by up to 0.26 °C, potentially favoring also soil water holding capacity and fertility [84]. Moreover, irrigation can offset crop production loss due to global warming projected by the 2050s, although >5% expansion of irrigation areas in warm regions is needed to fully counteract losses [85]. These suggest that multi-factorial studies are needed to optimize the cultivation techniques for concurrent adaptation to ozone, CO₂, and warming at field scale.

Diversification of cropping systems for adaptation

Nowadays farming systems often utilize crop rotation (growing different crops on the same field in successive growing season), intercropping (simultaneously cultivating two or more crops on the same field), and overall crop diversification (incorporating additional new crops or cropping systems to existing farmland) [86]. These systems aid in increasing crop diversity, reducing chemical pollution, repressing pests and diseases, enhancing soil health and nutrient and use efficiency, promoting benefits of soil microorganisms, and increasing yields [86]. Agroforestry also promotes diversification, mitigates climate change impacts, and facilitates ecosystem services and economic development [87–89]. Such diversifying practices have a great potential to enhance agricultural sustainability and promote resource savings. For instance, modifications in crop rotations and distributions (switching) can contribute 23–40% toward the targets of agricultural sustainable development in China by 2030 and facilitate global resource savings [90]. Among other benefits, fertilizers (-532 to -10.9%), pesticides (-4.3 to -10.8%) and greenhouse gases (-1.7 to -7.7%) can be decreased while farmers incomes are increased (+2.9 to +7.5%) [90]. Similar to crops, a recent analysis of Canada's National Forest Inventory (NFI) data revealed that enhancing functional diversity of trees increases decadal soil carbon (32%) and nitrogen (50%) in the mineral horizon [91]. Hence, such structural changes in the agroecosystems, including agroforestry, are widely addressed in the agroecological literature and have the potential to serve as a key to adaptation [92–95]. However, the role of diversifying crop systems under air pollution with or without combined climate changes has received little interest and remains elusive.

Diversifying crop cultures from monocultures to multi-species cultures increases annual primary productivity, plant biomass, and seed yield, but can decrease harvest index, a side effect that may be attributed to the breeding of current cultivars for best performance in monocultures [96]. However, from a climate change adaptation perspective, having a lower harvest index is not necessarily negative, if yields are also higher, given that the additional biomass production is incorporated into the soil to increase carbon sequestration, obtaining adaptation benefits from higher soil organic matter (SOM). Hence, greater seed yield gains may be compromised by diversification of cropping systems with crop cultivars bred for monocultures [96], suggesting that breeding programs should be optimized for mixed cropping systems to improve crop yields under future climate change and air pollution. Richness loss of services-providing organisms such as pest predator and pollinator contributes up to 50% of the impacts of landscape simplification on ecosystem services, negatively affecting crop yields [97]. Hence, to sustain crop productivity and agroecosystem health, the diversity of services-providing organisms should also be sustained [98]. The enhancement of crop productivity by increased crop diversity are partly driven by changes in the composition of soil microbial communities, especially increased abundance of plant-growth promoting microorganisms such as Actinobacteria [99]. However, the effects of crop diversity on communities of soil microorganisms depend on soil conditions, such as fertilization and moisture [99]. However, the continuum soil microbes-crop diversity-crop productivity is not well understood [99], calling for more research to optimize the outcomes. Diversification of cropping systems can also decrease the inputs of agrochemicals in the agroecosystems, thus offering a sustainable intervention for environmental health [100]. Hence, conservation of on-farm crop diversification is a promising approach to enhance crop

production across the world, including developing countries [98,101,102] in future climate change and air pollution scenarios.

A system with crop diversification can enhance water infiltration, water storage, and reduce water runoff [103], increasing cooling capacity of the canopy against heat events [104,105]. An intercropping system of potato with legumes facilitates reducing soil temperature and increasing crop water productivity and radiation use efficiency as compared to solo potato [106]. Cooling effects may minimize the negative effects of heat stress while stomata are still open, but this might harm crops due to an increased ozone uptake [7,107]. There is an increasing understanding of solo and interactive effects of elevated CO₂, ozone, and high temperature based on both modeling work [108–114] and experimental studies [115]. However, the tradeoff effects induced by mixture of canopy structure and crop diversification (i.e. in mixed cropping system) on various climate factors, particularly with ozone, and their interactions need future studies.

Emerging technologies and management

Ethylenediurea (EDU) is a synthetic chemical and antiozonant protecting numerous crops [46,116–119]. Potential application of antiozonants within agroecosystems in the future would require the selection of the application method [120], but more toxicological and ecotoxicological studies covering the entire spectrum of low and high dose effects (hormesis) are needed before it can be applied into the agroecosystems. Moreover, cost-benefit analyses are needed to optimize the application interval and concentration and amount relative to the desired yield benefits. A more efficient use of antiozonants would require the concurrent protection against air pollutants and climate changes. For example, pretreatment of rice (*Oryza sativa* L.) plants with calcium acetate led to higher rice yield under combined O₃ and heat stresses [121]. Similarly, preliminary results show that EDU can improve plant functionality under water deficit stress in O₃-polluted ambient air (*Fraxinus ornus*; [122]), adverse ambient O₃ stress combined with particulate matter deposition (*Triticum aestivum*; [123]), and O₃ phytotoxicity under elevated CO₂ (*Oryza sativa*; [124]), hinting to the possibility of antiozonant application under multiple environmental stresses. Recent studies provide evidence for nanomaterials-induced protection against O₃ [125,126], with mechanisms resembling those protecting against other oxidative stresses [118], suggesting that nanomaterials can provide protection against various air pollutants and climate changes that induce oxidative stress in plants. Therefore, future chemical engineering developments have the potential to facilitate the incorporation of greener technologies with a lower potential accumulation of residues in the environment [118].

Biostimulants represent an extremely broad category of biological products that enhance plant productivity by affecting plants directly and indirectly [127,128]. Biostimulants often enhance the quality of harvestable products and their nutritional value [129]. With the tremendous increase in the use of biostimulants in the agricultural sector in the recent years [130], their simultaneous protection against air pollution and climate change is promising. Considerable literature indicates their efficiency in mitigating abiotic stresses and their potential as a low-input tool of cropping management for more sustainable farming [131,132]. These deductions are substantiated by recent studies showing different biostimulants protecting crops

against O₃ damage [133–135], which adds to the widespread evidence for protection against climate changes.

Melatonin is also a highly pleiotropic molecule produced by animals and plants, which affects numerous diverse functions and processes in plants, including the circadian rhythms and its crucial role in day/night cycles, growth, photosynthesis, and rhizogenesis [136,137]. Melatonin improves plants and edible plant products by ca.7-30% against various elements of climate change and pollution [136,137]. The efficient protection of melatonin against different types of stresses suggests its suitability as a plant protectant that can facilitate its incorporation into the agricultural practice. While melatonin is proposed as a biostimulant for crops and post-harvest products, multidisciplinary studies are needed to explore the links of the impact of its use among agricultural, food technology, human nutrition, and environmental health [138].

Priming accelerates and boosts the defense response during subsequent exposures and leads to induced resistance and/or tolerance to both biotic and abiotic stress, including major climate changes such as heat and drought [139–141]. Seed priming can decrease the cost required for treatments on the field and to considerably larger plants, thus being a resource- and labor-efficient approach that poses also minimal environmental risk [140,142]. Priming gives an additional benefit as it can be used to regulate the (a)synchronous occurrence of ontogenic stages that are particularly sensitive to air pollutants to seasonal peaks [143–145]. On-field application of seed priming leads to increased economic returns and decreased cost-benefit ratio, with the greatest outcomes obtained when priming is incorporated with other agronomic practices, such as fertilizer micro-dosing and row spacing [144,145]. The use of nanotechnology, i.e. application of nano-primers, can facilitate an even more eco-friendly and sustainable priming [143], to protect against a broad spectrum of environmental stresses associated with air pollution and climate change.

Phenotyping robots can conduct measurements of plant traits at a high throughput and precision rate and thus have a great potential to support data acquisition for a better understanding of plant reactions in a changing environment. Assessment by phenotyping robots can accelerate the breeding process and allows for a closer study of the effects of environmental factors on complex plant traits [146]. The combination of phenotyping robots with precision farming has a significant potential for more target-oriented inputs and reduced impact on ecosystems, such as by decreasing greenhouse gas emissions by site-specific application of fertilizers or plant protection measures [147]. Moreover, phenotyping can speed up the breeding process and reveal new genotypic differences [148,149]. Therefore, phenotyping robot-assisted farming can aid efforts to adapt to ozone, CO₂, and warming by offering fast and real-time large data, providing a framework for the integration of a more holistic view to mitigate climate change and develop climate-resilient cultivars.

Adaptation options at ecosystem scale

Enhancing soil resilience

Climate change factors, such as elevated CO₂, ozone and warming, negatively influence SOM content and soil biota activity, which may consequently affect soil fertility, crop productivity, and the environment. For example, although elevated CO₂ increases the net primary productivity [150], a lower SOM storage is expected because of the faster decomposition of SOM under elevated CO₂ [151]. Warming enhances native SOM loss via altering soil functional microbial communities [152]. For instance, warming and elevated ozone alone and in combination generate tradeoffs between arbuscular mycorrhizae and the host roots and stimulate organic carbon decomposition in nontilled soybean agroecosystems [153]. As a key indicator of soil resilience, SOM content determines the level of soil fertility to a great extent and thus crop yield [154,155]. Therefore, the loss of SOM resulting from climate change will likely decrease soil resilience and productivity. Increasing soil carbon storage is a promising way of enhancing soil resilience to mitigate the negative effect of climate change on agriculture. Lots of practices have been used to maintain SOM storage in agriculture, such as applying organic fertilizers, conservation tillage, straw mulching, and intercropping [156]. Some of these measures, e.g. incorporating organic ammendments into agricultural soils, have been proved to be efficient in increasing soil fertility and crop productivity via soil carbon sequestration [157].

To achieve adaptation to climate change and air pollution, soil organic matter should be increased via a set of management practices such as cover crops, agroforestry, organic inputs, reduced tillage, varieties, mixed farming [158]. These would directly affect the functioning of agroecosystem, including autonomy, biodiversity, infiltration, soil protection, water retention, nutrient availability, and carbon sequestration [158]. Thereby, impacts from climate change and resource depletion decrease [158]. Altogether, these would lead to increased yield and net primary productivity [158].

Incorporation of remote sensing technologies

Recent developments in remote sensing satellites include the Fluorescence Explorer (FLEX), which will be able to measure chlorophyll *a* fluorescence emitted from plants at high spatial- and temporal resolution from space after its launch in 2024 [159,160]. Passive or sun-induced plant chlorophyll fluorescence (SIF) measurements are sensitive indicators of photosynthesis. Since photosynthesis is the process where CO₂ is transformed into substance, the fluorescence signal serves as an integrative indicator of ecosystem dynamics and the status of vegetation functioning. The launch opens new perspectives and adaptation options since the integrated Fluorescence Imaging Spectrometer (FLORIS) allows for valuable assessments of direct and indirect ecosystem functions integrating effects of elevated CO₂, increased air pollution, and global warming. In order to support the FLEX data and correct, e.g., aerosol influences, Sentinel-3 satellites have been chosen to serve as tandem partners [160]. The auxiliary data provided by Sentinel-3 will be included to improve models of future atmospheric CO₂ concentrations. FLEX high sensitivity will enable generating instantaneous data which has not been aggregated over a longer time period and several overflights. By obtaining precise databases of dynamic crop adaptation, remote sensing can guide precision agriculture for advanced planning, crop yield and health estimation, and agroecosystems management [161]. Nevertheless, high degree of dependence on technology may be compromised in conditions of low availability of energy that

can be expected to increase in a context of resource depletion and increasing conflicts. For example, the ongoing Russo-Ukrainian War affects the food system more through its effects on energy prices than through the decreased crop production in the area [162].

Modified hydrology and microclimate of agricultural landscapes

Under drought stress, plants close stomata to reduce water loss, which is limiting the uptake of ozone by vegetation which consequently increases surface ozone concentrations [163,164]. Ecosystem-atmosphere interactions reduced ozone removal by water-stressed vegetation, which exacerbated ozone air pollution over Europe during 1960-2018 (Lin et al., 2020). Simulation studies showed that irrigation decreases surface ozone by 0–5 ppb in irrigated areas but increases them by 2–7 ppb over the surrounding non-irrigation areas. Irrigation largely affects ozone exposure, but local impacts depend on the seasonality of emissions and climate [111]. Large-scale irrigation has the potential to bring positive effects through flexibility to avoid ozone peaks by shifting cropping calendars under irrigation. However, under irrigation, crops are sensitive to ozone damage because stomata fully open, thus increasing ozone uptake [7]. For heat stress, the effect of irrigation is positive because its cooling effect helps to reduce canopy temperature and thus restrict the negative effects of heat stress [165,166]. However, a decrease in canopy temperature may prolong phenological processes in crops, thus affecting crop yields [167]. Moreover, increased water input through irrigation may result in significant sensitivity of the ecosystem CO₂ respiration during the vegetation period as well as relatively high releases of CH₄ and N₂O [14,168,169], which are major ozone precursors. These impacts might offset the positive effects of irrigation on regulating ozone [170]. However, how irrigation or related management measures affect biological or atmospheric processes, in particular ozone concentration, remains poorly investigated [171]. Freshwater use is considered a planetary process whose boundary can soon be trespassed, which presents the main problem related to irrigation expansion [172]. Moreover, changes in landscape patterns strongly alternate the hydrological processes and, thus, water availability for irrigating cropland [173,174] might indirectly influence the ozone concentration at larger scales.

Modifying the microclimate has also a high potential for improving crop performance in changing environmental conditions. One measure of modifying the microclimate for field crops is the integration of woody species in cropping systems (such as agroforestry), or agricultural landscapes [175]. Trees typically have deeper root systems than annual crops and can increase air humidity and decrease air temperature [176]. Moreover, via hydraulic lift they can provide moisture in more shallow soil layers to annual crops [177]. The combination of lower temperatures and higher air humidity potentially facilitates the adaption to heat stress, enhances ozone degradation, and improves CO₂ assimilation. However, exploiting such synergies would require carefully balanced and locally adapted agroforestry systems that are technically feasible and economically profitable.

Crop growth modeling

Given the large spatial variation in changes of climatic factors and ozone pollution across different global crop production regions [178,179], adapting crop production at regional scale

should be region-specific. Therefore, it is essential to assess the spatial-variation before developing adaptation strategies [180,181]. Crop growth models, which can simulate dynamics of crop-soil system as influenced by climate conditions, soil characteristics and crop management, have been widely used for assessing climate change impacts on crop productivity [182,183]. Some recent estimations of elevated ozone impact on crop production were mostly based on statistical regressions, which usually ignore the cultivar or regions differences in the responses of crop production [184]. Applying the crop growth models at regional scale provides the opportunity to estimate the integrated impacts of climate change and elevated ozone together [113]. Another potential application of crop models is evaluating the possible impacts of different adaptation strategies, focusing on crop management and cultivar shifting.

Crop growth models have been improved substantially to enhance the suitability across different climate change scenarios and different regions, especially through the efforts within the Agricultural Model Intercomparison and Improvement Project (AgMIP) [185]. However, most of the current improvements in crop growth models focused on simulating the impacts of climate variables (e.g. rising temperatures, extreme climate events, CO₂) [186]. Elevated ozone potentially interacts with climate factors (rising temperature, CO₂) on several important growth processes, especially on leaf photosynthesis leaf senescence and grain C/N dynamics [187,188]. Current crop growth models rarely incorporated these interactions, except some empirical factors that were recently introduced into the models to modify the leaf photosynthesis on senescence [189,190]. Modeling the effects of rising ozone and temperature on leaf stomatal, and subsequently on evapotranspiration and canopy temperature should be the priority, as they have important implications for quantifying the accumulation of leaf photosynthesis and water use to elevated CO₂ under future climate. Coupling modeling with field empirical data from different cultivars and their responses is also needed. Furthermore, modeling should also account for dynamic responses and non-linear phenomena to produce more reliable impact estimates [191,192].

Perspectives for crop production under climate change and air pollution

The herein analysis suggests that climate change may not necessarily result in lower yields across all crops and locations. This is partly due to ongoing countermeasures adopted by local governments as well as because of individual actions by farmers in an effort to increase crop yields and thus their income, although further efforts are clearly needed to maximize adaptation and resilience under concurrent climate changes and air pollution [64,90,94,193]. Future local adaptation would require its own specific extent of weather and air pollution predictors, but finer spatiotemporal resolution of integrated weather, air pollution and yield data should be achieved, especially to capture risks during the most critical crop growth stages [193].

The present analysis suggests that no single method or approach would suffice to adapt crop ecosystems to climate change and air pollution. A plethora of methods or approaches should be integrated considering not only crop productivity but also environmental impact, thus requiring cost-benefit analyses in terms of both agricultural and environmental sectors. The

adaptation includes different scales, i.e. plant, field, and ecosystem (Fig. 1). However, a new generation of research programs is needed to move to more multidimensional researches in which different sets of methods and approaches at different levels (plant, field, ecosystem) would be tested as to benefits under combined environmental stressors (Fig. 2). Incorporation of key traditional strategies of agroecology, such as biodiversification, soil management and water harvesting, into agroecosystem management would facilitate increasing resilience while providing economic benefits [194].

Adaptation at food system level may be additionally needed to counteract air pollution and climate change and their associated possible yield losses, such as by decreasing meat consumption or food waste [195,196]. This suggests that aspects other than farming would define the degree of adaptation of agroecosystems such as willingness of the public to adopt changes in lifestyle. For instance, advanced nitrogen management along with shifts in dietary customs could enhance planetary feeding capacity [197]. The complexity in climate change and air pollution lies in the fact that environmental, economic and social dimensions are dynamically linked in both antagonistic and mutually beneficial ways in a system that is highly unpredictable. The systemic approach and combination of disciplinary knowledge in interdisciplinary research enables a multi-perspective view for a closer approximation and better understanding of the phenomena. New approaches are formed in the common exchange process of ideas and perspectives, enabling innovation. However, with underlying differences in research methodology, interdisciplinary collaboration can undoubtedly be challenging, and opinions diverge already at the debate of which knowledge concept is at the forefront [198]. Since the debate over greenhouse gas emissions and the adaptation to climate change, air pollution and other biophysical and social stressors does affect a myriad of stakeholders, the interdisciplinary approach is sometimes found to be insufficient. It has been therefore suggested going beyond an interdisciplinary approach and applying an open transdisciplinary approach, a combination of formal basic knowledge from different disciplines with informal knowledge and application-oriented practical knowledge by linking them in a participatory and adaptive processes [199]. Such inter- and transdisciplinary research is highly needed to adapt crop ecosystems to climate change and air pollution.

Acknowledgements This study was supported by the Sino-German Mobility Programme (M-0105) and the National Natural Science Foundation of China (No. 42130714, 42207123, 42107270).

Competing interests

The authors declare that they have no conflict of interest.

Author contributions: E.A., M.F., and Z.F. designed the study. All authors contributed parts of different sections and, thus, all authors contributed to writing the initial draft. E.A., M.F., and Z.F. edited the initial draft and generated the first draft of the integrated manuscript. All authors revised the paper, read the final manuscript, and approved its submission.

References

- [1] G.S. Khush, Green revolution: the way forward, *Nat. Rev. Genet.* 2 (2001) 815–822. <https://doi.org/10.1038/35093585>.
- [2] N. Brisson, P. Gate, D. Gouache, G. Charmet, F.X. Oury, F. Huard, Why are wheat yields stagnating in Europe? A comprehensive data analysis for France, *F. Crop. Res.* 119 (2010) 201–212. <https://doi.org/10.1016/j.fcr.2010.07.012>.
- [3] United Nations, *World Population Prospects: The 2017 Revision. Economic & Social Affairs*, 1–53, 53., 2017.
- [4] IPCC, *Climate Change 2021: The Physical Science Basis. Contribution of Working Group I to the Sixth Assessment Report of the Intergovernmental Panel on Climate Change* [Masson-Delmotte, V., P. Zhai, A. Pirani, S. L. Connors, C. Péan, S. Berger, N. Caud, Y. Chen, 2021.
- [5] United Nations, *Transforming our world: The 2030 Agenda for Sustainable Development*. 16301(October), 1–35. UN General Assembly. <https://www.refworld.org/docid/57b6e3e44.html>, 2015.
- [6] F. Wang, J.D. Harindintwali, K. Wei, Y. Shan, Z. Mi, M.J. Costello, S. Grunwald, Z. Feng, F. Wang, Y. Guo, X. Wu, P. Kumar, M. Kästner, X. Feng, S. Kang, Z. Liu, Y. Fu, W. Zhao, C. Ouyang, J. Shen, H. Wang, S.X. Chang, D.L. Evans, R. Wang, C. Zhu, L. Xiang, J. Rinklebe, M. Du, L. Huang, Z. Bai, S. Li, R. Lal, M. Elsner, J.-P. Wigneron, F. Florindo, X. Jiang, S.M. Shaheen, X. Zhong, R. Bol, G.M. Vasques, X. Li, S. Pfautsch, M. Wang, X. He, E. Agathokleous, H. Du, H. Yan, F.O. Kengara, F. Brahushi, X.-E. Long, P. Pereira, Y.S. Ok, M.C. Rillig, E. Jeppesen, D. Barceló, X. Yan, N. Jiao, B. Han, A. Schäffer, J.M. Chen, Y. Zhu, H. Cheng, W. Amelung, C. Spötl, J. Zhu, J.M. Tiedje, F. Wang, J.D. Harindintwali, K. Wei, Y. Shan, Z. Mi, M.J. Costello, S. Grunwald, Z. Feng, F. Wang, Y. Guo, X. Wu, P. Kumar, M. Kästner, X. Feng, S. Kang, Z. Liu, Y. Fu, W. Zhao, C. Ouyang, J. Shen, H. Wang, S.X. Chang, D.L. Evans, R. Wang, C. Zhu, L. Xiang, J. Rinklebe, M. Du, L. Huang, Z. Bai, S. Li, R. Lal, M. Elsner, J.-P. Wigneron, F. Florindo, X. Jiang, S.M. Shaheen, X. Zhong, R. Bol, G.M. Vasques, X. Li, S. Pfautsch, M. Wang, X. He, E. Agathokleous, H. Du, H. Yan, F.O. Kengara, F. Brahushi, X.-E. Long, P. Pereira, Y.S. Ok, M.C. Rillig, E. Jeppesen, D. Barceló, X. Yan, N. Jiao, B. Han, A. Schäffer, J.M. Chen, Y. Zhu, H. Cheng, W. Amelung, C. Spötl, J. Zhu, J.M. Tiedje, *Climate change: Strategies for mitigation and adaptation*, *Innov. Geosci.* 1 (2023) 100015. <https://doi.org/10.59717/j.xinn-geo.2023.100015>.

- 556 [7] J. Fuhrer, Agroecosystem responses to combinations of elevated CO₂, ozone, and global
557 climate change, *Agric. Ecosyst. Environ.* 97 (2003) 1–20. [https://doi.org/10.1016/s0167-](https://doi.org/10.1016/s0167-8809(03)00125-7)
558 8809(03)00125-7.
- 559 [8] F. Otu-Larbi, A. Conte, S. Fares, O. Wild, K. Ashworth, Current and future impacts of
560 drought and ozone stress on Northern Hemisphere forests, *Glob. Chang. Biol.* 26 (2020)
561 6218–6234. <https://doi.org/10.1111/gcb.15339>.
- 562 [9] M.R. Smith, S.S. Myers, Global health implications of nutrient changes in rice under high
563 atmospheric carbon dioxide, *GeoHealth*. 3 (2019) 190–200.
564 <https://doi.org/10.1029/2019gh000188>.
- 565 [10] I. Loladze, Hidden shift of the ionome of plants exposed to elevated CO₂ depletes
566 minerals at the base of human nutrition, *Elife*. 3 (2014) e02245.
567 <https://doi.org/10.7554/elif.02245>.
- 568 [11] S.G. Chavan, R.A. Duursma, M. Tausz, O. Ghannoum, Elevated CO₂ alleviates the
569 negative impact of heat stress on wheat physiology but not on grain yield, *J. Exp. Bot.* 70
570 (2019) 6447–6459. <https://doi.org/10.1093/jxb/erz386>.
- 571 [12] A.A. Singh, A. Ghosh, B. Pandey, M. Agrawal, S.B. Agrawal, Unravelling the ozone
572 toxicity in *Zea mays* L. (C₄ plant) under the elevated level of CO₂ fertilization, *Trop. Ecol.*
573 2023. (2023) 1–17. <https://doi.org/10.1007/s42965-023-00298-6>.
- 574 [13] H. Poorter, O. Knopf, I.J. Wright, A.A. Temme, S.W. Hogewoning, A. Graf, L.A.
575 Cernusak, T.L. Pons, A meta-analysis of responses of C₃ plants to atmospheric CO₂:
576 dose–response curves for 85 traits ranging from the molecular to the whole-plant level,
577 *New Phytol.* 233 (2022) 1560–1596. <https://doi.org/10.1111/nph.17802>.
- 578 [14] L. Samaniego, S. Thober, R. Kumar, N. Wanders, O. Rakovec, M. Pan, M. Zink, J.
579 Sheffield, E.F. Wood, A. Marx, Anthropogenic warming exacerbates European soil
580 moisture droughts, *Nat. Clim. Chang.* 8 (2018) 421–426. [https://doi.org/10.1038/s41558-](https://doi.org/10.1038/s41558-018-0138-5)
581 018-0138-5.
- 582 [15] D. Zhou, J. Xiao, S. Frolking, S. Liu, L. Zhang, Y. Cui, G. Zhou, Croplands intensify
583 regional and global warming according to satellite observations, *Remote Sens. Environ.*
584 264 (2021) 112585. <https://doi.org/10.1016/j.rse.2021.112585>.
- 585 [16] M. Lin, L.W. Horowitz, Y. Xie, F. Paulot, S. Malyshev, E. Shevliakova, A. Finco, G.
586 Gerosa, D. Kubistin, K. Pilegaard, Vegetation feedbacks during drought exacerbate ozone
587 air pollution extremes in Europe, *Nat. Clim. Chang.* 10 (2020) 444–451.
588 <https://doi.org/10.1038/s41558-020-0743-y>.
- 589 [17] M. Frei, Breeding of ozone resistant rice: Relevance, approaches and challenges, *Environ.*
590 *Pollut.* 197 (2015) 144–155. <https://doi.org/10.1016/j.envpol.2014.12.011>.
- 591 [18] G. Mills, K. Sharps, D. Simpson, H. Pleijel, M. Frei, K. Burkey, L. Emberson, J. Uddling,
592 M. Broberg, Z. Feng, K. Kobayashi, M. Agrawal, Closing the global ozone yield gap:
593 Quantification and cobenefits for multistress tolerance, *Glob. Chang. Biol.* 24 (2018)
594 4869–4893. <https://doi.org/10.1111/gcb.14381>.
- 595 [19] B.C.Y. Collard, D.J. Mackill, Marker-assisted selection: an approach for precision plant

breeding in the twenty-first century, *Philos. Trans. R. Soc. B Biol. Sci.* 363 (2007) 557–572. <https://doi.org/10.1098/rstb.2007.2170>.

- [20] W. Wang, R. Mauleon, Z. Hu, D. Chebotarov, S. Tai, Z. Wu, M. Li, T. Zheng, R.R. Fuentes, F. Zhang, L. Mansueto, D. Copetti, M. Sanciangco, K.C. Palis, J. Xu, C. Sun, B. Fu, H. Zhang, Y. Gao, X. Zhao, F. Shen, X. Cui, H. Yu, Z. Li, M. Chen, J. Detras, Y. Zhou, X. Zhang, Y. Zhao, D. Kudrna, C. Wang, R. Li, B. Jia, J. Lu, X. He, Z. Dong, J. Xu, Y. Li, M. Wang, J. Shi, J. Li, D. Zhang, S. Lee, W. Hu, A. Poliakov, I. Dubchak, V.J. Ulat, F.N. Borja, J.R. Mendoza, J. Ali, Q. Gao, Y. Niu, Z. Yue, M.E.B. Naredo, J. Talag, X. Wang, J. Li, X. Fang, Y. Yin, J.C. Glaszmann, J. Zhang, J. Li, R.S. Hamilton, R.A. Wing, J. Ruan, G. Zhang, C. Wei, N. Alexandrov, K.L. McNally, Z. Li, H. Leung, Genomic variation in 3,010 diverse accessions of Asian cultivated rice, *Nature*. 557 (2018) 43–49. <https://doi.org/10.1038/s41586-018-0063-9>.
- [21] M. Tester, P. Langridge, Breeding technologies to increase crop production in a changing world, *Science*. 327 (2010) 818–822. <https://doi.org/10.1126/science.1183700>.
- [22] C. Ye, X. Li, E. Redoña, T. Ishimaru, K. Jagadish, C. Ye, X. Li, E. Redoña, T. Ishimaru, K. Jagadish, Genetics and Breeding of Heat Tolerance in Rice, in: J. Ali, S.H. Wani (Eds.), *Rice Improv.*, Springer, Cham, 2021: pp. 203–220. https://doi.org/10.1007/978-3-030-66530-2_7.
- [23] M. Frei, J.P. Tanaka, M. Wissuwa, Genotypic variation in tolerance to elevated ozone in rice: dissection of distinct genetic factors linked to tolerance mechanisms, *J. Exp. Bot.* 59 (2008) 3741–3752. <https://doi.org/10.1093/jxb/ern222>.
- [24] C.A. Sorgini, I. Barrios-Perez, P.J. Brown, E.A. Ainsworth, Examining genetic variation in maize inbreds and mapping oxidative stress response QTL in B73-Mo17 nearly isogenic lines, *Front. Sustain. Food Syst.* 3 (2019) 51. <https://doi.org/10.3389/fsufs.2019.00051>.
- [25] H. Begum, M.S. Alam, Y. Feng, P. Koua, M. Ashrafuzzaman, A. Shrestha, M. Kamruzzaman, S. Dadshani, A. Ballvora, A.A. Naz, M. Frei, Genetic dissection of bread wheat diversity and identification of adaptive loci in response to elevated tropospheric ozone, *Plant. Cell Environ.* 43 (2020) 2650–2665. <https://doi.org/10.1111/pce.13864>.
- [26] N. Waldeck, K. Burkey, T. Carter, D. Dickey, Q. Song, E. Taliencio, RNA-Seq study reveals genetic responses of diverse wild soybean accessions to increased ozone levels, *BMC Genomics*. 18 (2017) 1–10. <https://doi.org/10.1186/s12864-017-3876-2>.
- [27] C.R. Yendrek, R.P. Koester, E.A. Ainsworth, A comparative analysis of transcriptomic, biochemical, and physiological responses to elevated ozone identifies species-specific mechanisms of resilience in legume crops, *J. Exp. Bot.* 66 (2015) 7101–7112. <https://doi.org/10.1093/jxb/erv404>.
- [28] M. Tausz, S. Tausz-Posch, R.M. Norton, G.J. Fitzgerald, M.E. Nicolas, S. Seneweera, Understanding crop physiology to select breeding targets and improve crop management under increasing atmospheric CO₂ concentrations, *Environ. Exp. Bot.* 88 (2013) 71–80. <https://doi.org/10.1016/j.envexpbot.2011.12.005>.
- [29] L.P. Dai, X.L. Lu, W.W. Zou, C.J. Wang, L. Shen, J. Hu, G.H. Zhang, D.Y. Ren, G. Chen,

- Q. Zhang, D.W. Xue, G.J. Dong, Z.Y. Gao, L.B. Guo, L. Zhu, T.M. Mou, Q. Qian, D.L. Zeng, Mapping of QTLs for source and sink associated traits under elevated CO₂ in rice (*Oryza sativa* L.), *Plant Growth Regul.* 90 (2020) 359–367. <https://doi.org/10.1007/s10725-019-00564-5>.
- [30] H. Nakano, S. Yoshinaga, T. Takai, Y. Arai-Sanoh, K. Kondo, T. Yamamoto, H. Sakai, T. Tokida, Y. Usui, H. Nakamura, T. Hasegawa, M. Kondo, Quantitative trait loci for large sink capacity enhance rice grain yield under free-air CO₂ enrichment conditions, *Sci. Rep.* 7 (2017) 1827. <https://doi.org/10.1038/s41598-017-01690-8>.
- [31] G. FAN, Y. DONG, C. WANG, J. WAN, H. XIE, C. XU, J. ZHU, Q. CAI, Analysis of QTLs for flag leaf shape and its response to elevated CO₂ in rice (*Oryza sativa*), *Rice Sci.* 14 (2007) 7–12. [https://doi.org/10.1016/s1672-6308\(07\)60002-x](https://doi.org/10.1016/s1672-6308(07)60002-x).
- [32] P. Palit, H. Kudapa, R. Zougmore, J. Kholova, A. Whitbread, M. Sharma, R.K. Varshney, An integrated research framework combining genomics, systems biology, physiology, modelling and breeding for legume improvement in response to elevated CO₂ under climate change scenario, *Curr. Plant Biol.* 22 (2020) 100149. <https://doi.org/10.1016/j.cpb.2020.100149>.
- [33] E.A. Ainsworth, S.P. Long, 30 years of free-air carbon dioxide enrichment (FACE): What have we learned about future crop productivity and its potential for adaptation?, *Glob. Chang. Biol.* 27 (2021) 27–49. <https://doi.org/10.1111/gcb.15375>.
- [34] J. Flexas, M. Ribas-Carbó, J. Bota, J. Galmés, M. Henkle, S. Martínez-Cañellas, H. Medrano, Decreased Rubisco activity during water stress is not induced by decreased relative water content but related to conditions of low stomatal conductance and chloroplast CO₂ concentration, *New Phytol.* 172 (2006) 73–82. <https://doi.org/10.1111/j.1469-8137.2006.01794.x>.
- [35] J. Flexas, M. Ribas-Carbó, A. Diaz-Espejo, J. Galmés, H. Medrano, Mesophyll conductance to CO₂: current knowledge and future prospects, *Plant. Cell Environ.* 31 (2008) 602–621. <https://doi.org/10.1111/J.1365-3040.2007.01757.X>.
- [36] E. Paoletti, Impact of ozone on Mediterranean forests: A review, *Environ. Pollut.* 144 (2006) 463–474. <https://doi.org/10.1016/j.envpol.2005.12.051>.
- [37] B.E. Medlyn, R.A. Duursma, D. Eamus, D.S. Ellsworth, I.C. Prentice, C.V.M. Barton, K.Y. Crous, P. De Angelis, M. Freeman, L. Wingate, Reconciling the optimal and empirical approaches to modelling stomatal conductance, *Glob. Chang. Biol.* 17 (2011) 2134–2144. <https://doi.org/10.1111/j.1365-2486.2010.02375.x>.
- [38] M. Papanatsiou, J. Petersen, L. Henderson, Y. Wang, J.M. Christie, M.R. Blatt, Optogenetic manipulation of stomatal kinetics improves carbon assimilation, water use, and growth, *Science.* 363 (2019) 1456–1459. <https://doi.org/10.1126/science.aaw0046>.
- [39] M. Qu, J. Essemine, J. Xu, G. Ablat, S. Perveen, H. Wang, K. Chen, Y. Zhao, G. Chen, C. Chu, X. Zhu, Alterations in stomatal response to fluctuating light increase biomass and yield of rice under drought conditions, *Plant J.* 104 (2020) 1334–1347. <https://doi.org/10.1111/tpj.15004>.

- [40] Y. Hoshika, M. Watanabe, N. Inada, T. Koike, Model-based analysis of avoidance of ozone stress by stomatal closure in Siebold's beech (*Fagus crenata*), *Ann. Bot.* 112 (2013) 1149–1158. <https://doi.org/10.1093/aob/mct166>.
- [41] Z. Feng, J. Pang, I. Nouchi, K. Kobayashi, T. Yamakawa, J. Zhu, Apoplastic ascorbate contributes to the differential ozone sensitivity in two varieties of winter wheat under fully open-air field conditions, *Environ. Pollut.* 158 (2010) 3539–3545. <https://doi.org/10.1016/j.envpol.2010.08.019>.
- [42] N.E. Choquette, E.A. Ainsworth, W. Bezodis, A.P. Cavanagh, Ozone tolerant maize hybrids maintain Rubisco content and activity during long-term exposure in the field, *Plant. Cell Environ.* 43 (2020) 3033. <https://doi.org/10.1111/pce.13876>.
- [43] E.A. Ainsworth, Understanding and improving global crop response to ozone pollution, *Plant J.* 90 (2017) 886–897. <https://doi.org/10.1111/tpj.13298>.
- [44] L. Dai, K. Kobayashi, I. Nouchi, Y. Masutomi, Z. Feng, Quantifying determinants of ozone detoxification by apoplastic ascorbate in peach (*Prunus persica*) leaves using a model of ozone transport and reaction, *Glob. Chang. Biol.* (2020) gcb.15049. <https://doi.org/10.1111/gcb.15049>.
- [45] A.M. Mashaheet, K.O. Burkey, C.J. Saitanis, A.S. Abdelrhim, Rafiullah, D.S. Marshall, Differential ozone responses identified among key rust-susceptible wheat genotypes, *Agronomy.* 10 (2020) 1853. <https://doi.org/10.3390/agronomy10121853>.
- [46] E. Agathokleous, M. Kitao, Y. Hoshika, M. Haworth, Y. Tang, T. Koike, Ethylenediurea protects against ozone phytotoxicity not by adding nitrogen or controlling stomata in a stomata-unresponsive hybrid poplar, *Sci. Total Environ.* 875 (2023) 162672. <https://doi.org/10.1016/j.scitotenv.2023.162672>.
- [47] N. Driedonks, I. Rieu, W.H. Vriezen, Breeding for plant heat tolerance at vegetative and reproductive stages, *Plant Reprod.* 29 (2016) 67–79. <https://doi.org/10.1007/s00497-016-0275-9>.
- [48] D. Lei, L. Tan, F. Liu, L. Chen, C. Sun, Identification of heat-sensitive QTL derived from common wild rice (*Oryza rufipogon* Griff.), *Plant Sci.* 201–202 (2013) 121–127. <https://doi.org/10.1016/j.plantsci.2012.12.001>.
- [49] N.L. Kilasi, J. Singh, C.E. Vallejos, C. Ye, S.V.K. Jagadish, P. Kusolwa, B. Rathinasabapathi, Heat stress tolerance in rice (*Oryza sativa* L.): Identification of quantitative trait loci and candidate genes for seedling growth under heat stress, *Front. Plant Sci.* 871 (2018) 1578. <https://doi.org/10.3389/fpls.2018.01578>.
- [50] Z. Chang-lan, X. Ying-hui, W. Chunming, J. Ling, Z. Huqu, W. Jianmin, Mapping QTL for heat-tolerance at grain filling stage in rice, *Rice Sci.* 12 (2005) 33–38. <https://doi.org/10.3389/fgene.2020.621871>.
- [51] T. Lafarge, C. Bueno, J. Frouin, L. Jacquin, B. Courtois, N. Ahmadi, Genome-wide association analysis for heat tolerance at flowering detected a large set of genes involved in adaptation to thermal and other stresses, *PLoS One.* 12 (2017) e0171254. <https://doi.org/10.1371/journal.pone.0171254>.

- 717 [52] S. Ps, A.M. Sv, C. Prakash, R. Mk, R. Tiwari, T. Mohapatra, N.K. Singh, High Resolution
718 Mapping of QTLs for heat tolerance in rice using a 5K SNP array, *Rice*. 10 (2017) 28.
719 <https://doi.org/10.1186/s12284-017-0167-0>.
- 720 [53] C. Malumpong, R. Buadchee, B. Thammasamisorn, P. Mounng-Ngam, B. Wasuri, C.
721 Saensuk, S. Arikrit, A. Vannavichit, S. Cheabu, Backcross breeding for improvement of
722 heat tolerance at reproductive phase in Thai rice (*Oryza sativa* L.) varieties, *J. Agric. Sci.*
723 158 (2020) 496–510. <https://doi.org/10.1017/s0021859620000957>.
- 724 [54] L. Chen, Q. Wang, M. Tang, X. Zhang, Y. Pan, X. Yang, G. Gao, R. Lv, W. Tao, L. Jiang,
725 T. Liang, QTL mapping and identification of candidate genes for heat tolerance at the
726 flowering stage in rice, *Front. Genet.* 11 (2021) 1840.
727 <https://doi.org/10.3389/fgene.2020.621871>.
- 728 [55] Q. Raza, A. Riaz, K. Bashir, M. Sabar, Reproductive tissues-specific meta-QTLs and
729 candidate genes for development of heat-tolerant rice cultivars, *Plant Mol. Biol.* 104 (2020)
730 97–112. <https://doi.org/10.1007/s11103-020-01027-6>.
- 731 [56] L. Li, X. Mao, J. Wang, X. Chang, M. Reynolds, R. Jing, Genetic dissection of drought
732 and heat-responsive agronomic traits in wheat, *Plant. Cell Environ.* 42 (2019) 2540–2553.
733 <https://doi.org/10.1111/pce.13577>.
- 734 [57] S.F. Abou-Elwafa, T. Shehzad, Genetic diversity, GWAS and prediction for drought and
735 terminal heat stress tolerance in bread wheat (*Triticum aestivum* L.), *Genet. Resour. Crop*
736 *Evol.* 68 (2020) 711–728. <https://doi.org/10.1007/s10722-020-01018-y>.
- 737 [58] H.G. Muhi-Din Ahmed, M. Naeem, Y. Zeng, M.A.R. Rashid, A. Ullah, A. Saeed, A.
738 Qadeer, Genome-wide association mapping for high temperature tolerance in wheat
739 through 90k SNP array using physiological and yield traits, *PLoS One*. 17 (2022)
740 e0262569. <https://doi.org/10.1371/journal.pone.0262569>.
- 741 [59] D. Van Inghelandt, F.P. Frey, D. Ries, B. Stich, QTL mapping and genome-wide
742 prediction of heat tolerance in multiple connected populations of temperate maize, *Sci.*
743 *Rep.* 9 (2019) 1–16. <https://doi.org/10.1038/s41598-019-50853-2>.
- 744 [60] N. Longmei, G.K. Gill, P.H. Zaidi, R. Kumar, S.K. Nair, V. Hindu, M.T. Vinayan, Y.
745 Vikal, Genome wide association mapping for heat tolerance in sub-tropical maize, *BMC*
746 *Genomics.* 22 (2021) 1–14. <https://doi.org/10.1186/s12864-021-07463-y>.
- 747 [61] K. Seetharam, P.H. Kuchanur, K.B. Koirala, M.P. Tripathi, A. Patil, V. Sudarsanam, R.R.
748 Das, R. Chaurasia, K. Pandey, H. Vemuri, M.T. Vinayan, S.K. Nair, R. Babu, P.H. Zaidi,
749 Genomic regions associated with heat stress tolerance in tropical maize (*Zea mays* L.), *Sci.*
750 *Rep.* 11 (2021) 1–11. <https://doi.org/10.1038/s41598-021-93061-7>.
- 751 [62] S.K. Bazzar, L.C. Purcell, Identification of quantitative trait loci associated with canopy
752 temperature in soybean, *Sci. Rep.* 10 (2020) 17604. <https://doi.org/10.1038/s41598-020-74614-8>.
- 754 [63] J. Kumar, D. Sen Gupta, I. Djalovic, Breeding, genetics, and genomics for tolerance
755 against terminal heat in lentil: Current status and future directions, *Legum. Sci.* 2 (2020)
756 e38. <https://doi.org/10.1002/leg3.38>.

- [64] Y. Masutomi, T. Takimoto, T. Manabe, Y. Imai, M. Tamura, K. Kobayashi, Breeding targets for heat-tolerant rice varieties in Japan in a warming climate, *Mitig. Adapt. Strateg. Glob. Chang.* 28 (2023) 2. <https://doi.org/10.1007/s11027-022-10027-4>.
- [65] M. Reynolds, P. Langridge, Physiological breeding, *Curr. Opin. Plant Biol.* 31 (2016) 162–171. <https://doi.org/10.1016/j.pbi.2016.04.005>.
- [66] M. Ashikari, M. Matsuoka, Identification, isolation and pyramiding of quantitative trait loci for rice breeding, *Trends Plant Sci.* 11 (2006) 344–350. <https://doi.org/10.1016/j.tplants.2006.05.008>.
- [67] Y. Wang, L. Yang, M. Höller, S. Zaisheng, J. Pariasca-Tanaka, M. Wissuwa, M. Frei, Pyramiding of ozone tolerance QTLs *OzT8* and *OzT9* confers improved tolerance to season-long ozone exposure in rice, *Environ. Exp. Bot.* 104 (2014) 26–33. <https://doi.org/10.1016/j.envexpbot.2014.03.005>.
- [68] J. Bailey-Serres, J.E. Parker, E.A. Ainsworth, G.E.D. Oldroyd, J.I. Schroeder, Genetic strategies for improving crop yields, *Nature.* 575 (2019) 109–118. <https://doi.org/10.1038/s41586-019-1679-0>.
- [69] Z. Feng, J. Pang, K. Kobayashi, J. Zhu, D.R. Ort, Differential responses in two varieties of winter wheat to elevated ozone concentration under fully open-air field conditions, *Glob. Chang. Biol.* (2011) 580–591. <https://doi.org/10.1111/j.1365-2486.2010.02184.x>.
- [70] C. Wei, S. Ren, P. Yang, Y. Wang, X. He, Z. Xu, R. Wei, S. Wang, Y. Chi, M. Zhang, Effects of irrigation methods and salinity on CO₂ emissions from farmland soil during growth and fallow periods, *Sci. Total Environ.* 752 (2021) 141639. <https://doi.org/10.1016/j.scitotenv.2020.141639>.
- [71] B. Mao, Y. Wang, T.H. Zhao, Q. Zhao, Y. San, S.S. Xiao, Response of carbon, nitrogen and phosphorus concentration and stoichiometry of plants and soils during a soybean growth season to O₃ stress and straw return in Northeast China, *Sci. Total Environ.* 822 (2022) 153573. <https://doi.org/10.1016/j.scitotenv.2022.153573>.
- [72] Z. Gou, W. Yin, Q. Chai, Straw and residual film management enhances crop yield and weakens CO₂ emissions in wheat–maize intercropping system, *Sci. Rep.* 11 (2021) 14077. <https://doi.org/10.1038/s41598-021-93497-x>.
- [73] A. Fares, A. Bensley, H. Bayabil, R. Awal, S. Fares, H. Valenzuela, F. Abbas, Carbon dioxide emission in relation with irrigation and organic amendments from a sweet corn field, *J. Environ. Sci. Health. B.* 52 (2017) 387–394. <https://doi.org/10.1080/03601234.2017.1292094>.
- [74] Z. Xu, H. Shimizu, S. Ito, Y. Yagasaki, C. Zou, G. Zhou, Y. Zheng, Effects of elevated CO₂, warming and precipitation change on plant growth, photosynthesis and peroxidation in dominant species from North China grassland, *Planta.* 239 (2013) 421–435. <https://doi.org/10.1007/s00425-013-1987-9>.
- [75] F. Changey, M. Bagard, M. Souleymane, T.Z. Lerch, Cascading effects of elevated ozone on wheat rhizosphere microbial communities depend on temperature and cultivar sensitivity, *Environ. Pollut.* 242 (2018) 113–125.

797 <https://doi.org/10.1016/j.envpol.2018.06.073>.

798 [76] T.J. Kou, W.W. Yu, S.K. Lam, D.L. Chen, Y.P. Hou, Z.Y. Li, Differential root responses
799 in two cultivars of winter wheat (*Triticum aestivum* L.) to elevated ozone concentration
800 under fully open-air field conditions, *J. Agron. Crop Sci.* 204 (2018) 325–332.
801 <https://doi.org/10.1111/jac.12257>.

802 [77] M.C. Broberg, S. Daun, H. Pleijel, Ozone induced loss of seed protein accumulation is
803 larger in soybean than in wheat and rice, *Agronomy*. 10 (2020) 357.
804 <https://doi.org/10.3390/agronomy10030357>.

805 [78] Z. Feng, B. Shang, Z. Li, V. Calatayud, E. Agathokleous, Ozone will remain a threat for
806 plants independently of nitrogen load, *Funct. Ecol.* 33 (2019) 1365–2435.13422.
807 <https://doi.org/10.1111/1365-2435.13422>.

808 [79] M.C. Broberg, J. Uddling, G. Mills, H. Pleijel, Fertilizer efficiency in wheat is reduced by
809 ozone pollution, *Sci. Total Environ.* 607–608 (2017) 876–880.
810 <https://doi.org/10.1016/j.scitotenv.2017.07.069>.

811 [80] J. Peng, Y. Xu, B. Shang, L. Qu, Z. Feng, Impact of ozone pollution on nitrogen
812 fertilization management during maize (*Zea mays* L.) production, *Environ. Pollut.* 266
813 (2020) 115158. <https://doi.org/10.1016/j.envpol.2020.115158>.

814 [81] B.A. Kimball, C.F. Morris, P.J. Pinter, G.W. Wall, D.J. Hunsaker, F.J. Adamsen, R.L.
815 LaMorte, S.W. Leavitt, T.L. Thompson, A.D. Matthias, T.J. Brooks, Elevated CO₂,
816 drought and soil nitrogen effects on wheat grain quality, *New Phytol.* 150 (2001) 295–303.
817 <https://doi.org/10.1046/j.1469-8137.2001.00107.x>.

818 [82] B. Liu, X. Wang, L. Ma, D. Chadwick, X. Chen, Combined applications of organic and
819 synthetic nitrogen fertilizers for improving crop yield and reducing reactive nitrogen
820 losses from China's vegetable systems: A meta-analysis, *Environ. Pollut.* 269 (2021)
821 116143. <https://doi.org/10.1016/j.envpol.2020.116143>.

822 [83] D. Helman, D.J. Bonfil, Six decades of warming and drought in the world's top wheat-
823 producing countries offset the benefits of rising CO₂ to yield, *Sci. Rep.* 12 (2022) 7921.
824 <https://doi.org/10.1038/s41598-022-11423-1>.

825 [84] A. Mayer, Z. Hausfather, A.D. Jones, W.L. Silver, The potential of agricultural land
826 management to contribute to lower global surface temperatures, *Sci. Adv.* 4 (2018)
827 eaaq0932. <https://doi.org/10.1126/sciadv.aaq0932>.

828 [85] P. Zhu, J. Burney, J. Chang, Z. Jin, N.D. Mueller, Q. Xin, J. Xu, L. Yu, D. Makowski, P.
829 Ciais, Warming reduces global agricultural production by decreasing cropping frequency
830 and yields, *Nat. Clim. Chang.* 12 (2022) 1016–1023. <https://doi.org/10.1038/s41558-022-01492-5>.

832 [86] T. Yang, K.H.M. Siddique, K. Liu, Cropping systems in agriculture and their impact on
833 soil health-A review, *Glob. Ecol. Conserv.* 23 (2020) e01118.
834 <https://doi.org/10.1016/j.gecco.2020.e01118>.

835 [87] J. Bettles, D.S. Battisti, S.C. Cook-Patton, T. Kroeger, J.T. Spector, N.H. Wolff, Y.J.
836 Masuda, Agroforestry and non-state actors: A review, *For. Policy Econ.* 130 (2021)

837 102538. <https://doi.org/10.1016/j.forpol.2021.102538>.

838 [88] Y. Telwala, Unlocking the potential of agroforestry as a nature-based solution for
839 localizing sustainable development goals: A case study from a drought-prone region in
840 rural India, *Nature-Based Solut.* 3 (2023) 100045.
841 <https://doi.org/10.1016/j.nbsj.2022.100045>.

842 [89] I. Apuri, K. Peprah, G.T.W. Achana, Climate change adaptation through agroforestry: The
843 case of Kassena Nankana West District, Ghana, *Environ. Dev.* 28 (2018) 32–41.
844 <https://doi.org/10.1016/j.envdev.2018.09.002>.

845 [90] W. Xie, A. Zhu, T. Ali, Z. Zhang, X. Chen, F. Wu, J. Huang, K.F. Davis, Crop switching
846 can enhance environmental sustainability and farmer incomes in China, *Nature.* 616 (2023)
847 300–305. <https://doi.org/10.1038/s41586-023-05799-x>.

848 [91] X. Chen, A.R. Taylor, P.B. Reich, M. Hisano, H.Y.H. Chen, S.X. Chang, Tree diversity
849 increases decadal forest soil carbon and nitrogen accrual, *Nature.* 618 (2023) 94–101.
850 <https://doi.org/10.1038/s41586-023-05941-9>.

851 [92] S.E. Castle, D.C. Miller, N. Merten, P.J. Ordonez, K. Baylis, Evidence for the impacts of
852 agroforestry on ecosystem services and human well-being in high-income countries: a
853 systematic map, *Environ. Evid.* 11 (2022) 1–27. [https://doi.org/10.1186/s13750-022-](https://doi.org/10.1186/s13750-022-00260-4)
854 [00260-4](https://doi.org/10.1186/s13750-022-00260-4).

855 [93] B. Ghale, E. Mitra, H.S. Sodhi, A.K. Verma, S. Kumar, Carbon sequestration potential of
856 agroforestry systems and its potential in climate change mitigation, *Water, Air, Soil Pollut.*
857 233 (2022) 1–27. <https://doi.org/10.1007/s11270-022-05689-4>.

858 [94] K.M. Dittmer, S. Rose, S.S. Snapp, Y. Kebede, S. Brickman, S. Shelton, C. Egler, M.
859 Stier, E. Wollenberg, Agroecology can promote climate change adaptation outcomes
860 without compromising yield in smallholder systems, *Environ. Manage.* 72 (2023) 333–
861 342. <https://doi.org/10.1007/s00267-023-01816-x>.

862 [95] J. Rising, N. Devineni, Crop switching reduces agricultural losses from climate change in
863 the United States by half under RCP 8.5, *Nat. Commun.* 11 (2020) 1–7.
864 <https://doi.org/10.1038/s41467-020-18725-w>.

865 [96] J. Chen, N. Engbersen, L. Stefan, B. Schmid, H. Sun, C. Schöb, Diversity increases yield
866 but reduces harvest index in crop mixtures, *Nat. Plants* 2021 77. 7 (2021) 893–898.
867 <https://doi.org/10.1038/s41477-021-00948-4>.

868 [97] M. Dainese, E.A. Martin, M.A. Aizen, M. Albrecht, I. Bartomeus, R. Bommarco, L.G.
869 Carvalheiro, R. Chaplin-Kramer, V. Gagic, L.A. Garibaldi, J. Ghazoul, H. Grab, M.
870 Jonsson, D.S. Karp, C.M. Kennedy, D. Kleijn, C. Kremen, D.A. Landis, D.K. Letourneau,
871 L. Marini, K. Poveda, R. Rader, H.G. Smith, T. Tscharntke, G.K.S. Andersson, I.
872 Badenhauer, S. Baensch, A.D.M. Bezerra, F.J.J.A. Bianchi, V. Boreux, V. Bretagnolle,
873 B. Caballero-Lopez, P. Cavigliasso, A. Četković, N.P. Chacoff, A. Classen, S. Cusser, F.D.
874 Da Silva E Silva, G. Arjen De Groot, J.H. Dudenhöffer, J. Ekroos, T. Fijen, P. Franck,
875 B.M. Freitas, M.P.D. Garratt, C. Gratton, J. Hipólito, A. Holzschuh, L. Hunt, A.L. Iverson,
876 S. Jha, T. Keasar, T.N. Kim, M. Kishinevsky, B.K. Klatt, A.M. Klein, K.M. Krewenka, S.
877 Krishnan, A.E. Larsen, C. Lavigne, H. Liere, B. Maas, R.E. Mallinger, E.M. Pachon, A.

- 878 Martínez-Salinas, T.D. Meehan, M.G.E. Mitchell, G.A.R. Molina, M. Nesper, L. Nilsson,
879 M.E. O'Rourke, M.K. Peters, M. Plečáš, S.G. Potts, D. de L. Ramos, J.A. Rosenheim, M.
880 Rundlöf, A. Rusch, A. Sáez, J. Scheper, M. Schleuning, J.M. Schmack, A.R. Sciligo, C.
881 Seymour, D.A. Stanley, R. Stewart, J.C. Stout, L. Sutter, M.B. Takada, H. Taki, G.
882 Tamburini, M. Tschumi, B.F. Viana, C. Westphal, B.K. Willcox, S.D. Wratten, A.
883 Yoshioka, C. Zaragoza-Trello, W. Zhang, Y. Zou, I. Steffan-Dewenter, A global synthesis
884 reveals biodiversity-mediated benefits for crop production, *Sci. Adv.* 5 (2019).
885 <https://doi.org/10.1126/sciadv.aax0121>.
- 886 [98] C. Bellora, É. Blanc, J.-M. Bourgeon, E. Strobl, E. Strobl Amse-Greqam, Estimating the
887 impact of crop diversity on agricultural productivity in South Africa, *NBER Chapters*.
888 (2017) 185–215. <https://doi.org/10.3386/w23496>.
- 889 [99] L. Stefan, M. Hartmann, N. Engbersen, J. Six, C. Schöb, Positive Effects of crop diversity
890 on productivity driven by changes in soil microbial composition, *Front. Microbiol.* 12
891 (2021) 808. <https://doi.org/10.3389/fmicb.2021.660749>.
- 892 [100] A.S. Davis, J.D. Hill, C.A. Chase, A.M. Johanns, M. Liebman, Increasing cropping
893 system diversity balances productivity, profitability and environmental health, *PLoS One*.
894 7 (2012) e47149. <https://doi.org/10.1371/journal.pone.0047149>.
- 895 [101] H.P.P. Donfouet, A. Barczak, C. Détang-Dessendre, E. Maigné, Crop production and crop
896 diversity in France: A spatial analysis, *Ecol. Econ.* 134 (2017) 29–39.
897 <https://doi.org/10.1016/j.ecolecon.2016.11.016>.
- 898 [102] C. Li, X. He, S. Zhu, H. Zhou, Y. Wang, Y. Li, J. Yang, J. Fan, J. Yang, G. Wang, Y.
899 Long, J. Xu, Y. Tang, G. Zhao, J. Yang, L. Liu, Y. Sun, Y. Xie, H. Wang, Y. Zhu, Crop
900 diversity for yield increase, *PLoS One*. 4 (2009) e8049.
901 <https://doi.org/10.1371/journal.pone.0008049>.
- 902 [103] D. Beillouin, T. Ben-Ari, E. Malézieux, V. Seufert, D. Makowski, Positive but variable
903 effects of crop diversification on biodiversity and ecosystem services, *Glob. Chang. Biol.*
904 27 (2021) 4697–4710. <https://doi.org/10.1111/gcb.15747>.
- 905 [104] J. Tack, A. Barkley, N. Hendricks -, W. African Sudan Savanna Babacar Faye, H. Webber,
906 S. Siebert, F. Ewert, E. Eyshi Rezaei, H. Kage, R. Graß, Impact of heat stress on crop
907 yield—on the importance of considering canopy temperature, *Environ. Res. Lett.* 9 (2014)
908 044012. <https://doi.org/10.1088/1748-9326/9/4/044012>.
- 909 [105] H. Webber, P. Martre, S. Asseng, B. Kimball, J. White, M. Ottman, G.W. Wall, G. De
910 Sanctis, J. Doltra, R. Grant, B. Kassie, A. Maiorano, J.E. Olesen, D. Ripoche, E.E. Rezaei,
911 M.A. Semenov, P. Stratonovitch, F. Ewert, Canopy temperature for simulation of heat
912 stress in irrigated wheat in a semi-arid environment: A multi-model comparison, *F. Crop*.
913 *Res.* 202 (2017) 21–35. <https://doi.org/10.1016/j.fcr.2015.10.009>.
- 914 [106] S.O. Nyawade, N.N. Karanja, C.K.K. Gachene, H.I. Gitari, E. Schulte-Geldermann, M.L.
915 Parker, Intercropping optimizes soil temperature and increases crop water productivity
916 and radiation use efficiency of rainfed potato, *Am. J. Potato Res.* 96 (2019) 457–471.
917 <https://doi.org/10.1007/s12230-019-09737-4>.
- 918 [107] J.S. Amthor, Effects of atmospheric CO₂ concentration on wheat yield: review of results

919 from experiments using various approaches to control CO₂ concentration, F. Crop. Res. 73
920 (2001) 1–34. [https://doi.org/10.1016/s0378-4290\(01\)00179-4](https://doi.org/10.1016/s0378-4290(01)00179-4).

921 [108] R.A. Díaz, G.O. Magrín, M.I. Travasso, R.O. Rodríguez, Climate change and its impact
922 on the properties of agricultural soils in the Argentinean Rolling Pampas, Clim. Res. 09
923 (1997) 25–30. <https://doi.org/10.3354/cr009025>.

924 [109] F.N. Tubiello, M. Donatelli, C. Rosenzweig, C.O. Stockle, Effects of climate change and
925 elevated CO₂ on cropping systems: model predictions at two Italian locations, Eur. J.
926 Agron. 13 (2000) 179–189. [https://doi.org/10.1016/s1161-0301\(00\)00073-3](https://doi.org/10.1016/s1161-0301(00)00073-3).

927 [110] K. Paustian, E.T. Elliott, G.A. Peterson, K. Killian, Modelling climate, CO₂ and
928 management impacts on soil carbon in semi-arid agroecosystems, Plant Soil. 187 (1995)
929 351–365. <https://doi.org/10.1007/bf00017100>.

930 [111] E. Teixeira, G. Fischer, H. van Velthuis, R. van Dingenen, F. Dentener, G. Mills, C.
931 Walter, F. Ewert, Limited potential of crop management for mitigating surface ozone
932 impacts on global food supply, Atmos. Environ. 45 (2011) 2569–2576.
933 <https://doi.org/10.1016/j.atmosenv.2011.02.002>.

934 [112] F. Ewert, M. Van Oijen, J.R. Porter, Simulation of growth and development processes of
935 spring wheat in response to CO₂ and ozone for different sites and years in Europe using
936 mechanistic crop simulation models, Eur. J. Agron. 10 (1999) 231–247.
937 [https://doi.org/10.1016/s1161-0301\(99\)00013-1](https://doi.org/10.1016/s1161-0301(99)00013-1).

938 [113] F. Tao, Z. Feng, H. Tang, Y. Chen, K. Kobayashi, Effects of climate change, CO₂ and O₃
939 on wheat productivity in Eastern China, singly and in combination, Atmos. Environ. 153
940 (2017) 182–193. <https://doi.org/10.1016/j.atmosenv.2017.01.032>.

941 [114] A.P.K. Tai, M. Sadiq, J.Y.S. Pang, D.H.Y. Yung, Z. Feng, Impacts of surface ozone
942 pollution on global crop yields: comparing different ozone exposure metrics and
943 incorporating co-effects of CO₂, Front. Sustain. Food Syst. 5 (2021) 63.
944 <https://doi.org/10.3389/fsufs.2021.534616>.

945 [115] F.A. Dijkstra, S.A. Prior, G.B. Runion, H.A. Torbert, H. Tian, C. Lu, R.T. Venterea,
946 Effects of elevated carbon dioxide and increased temperature on methane and nitrous
947 oxide fluxes: evidence from field experiments, Front. Ecol. Environ. 10 (2012) 520–527.
948 <https://doi.org/10.1890/120059>.

949 [116] E. Agathokleous, M. Kitao, C. Shi, N. Masui, S. Abu-ElEla, K. Hikino, F. Satoh, T. Koike,
950 Ethylenediurea (EDU) spray effects on willows (*Salix sachalinensis* F. Schmid) grown in
951 ambient or ozone-enriched air: implications for renewable biomass production, J. For. Res.
952 33 (2022) 397–422. <https://doi.org/10.1007/s11676-021-01400-1>.

953 [117] E. Agathokleous, M. Kitao, X. Wang, Q. Mao, H. Harayama, W.J. Manning, T. Koike,
954 Ethylenediurea (EDU) effects on Japanese larch: an one growing season experiment with
955 simulated regenerating communities and a four growing season application to individual
956 saplings, J. For. Res. 32 (2021) 2047–2057. <https://doi.org/10.1007/s11676-020-01223-6>.

957 [118] C.J. Saitanis, E. Agathokleous, Exogenous application of chemicals for protecting plants
958 against ambient ozone pollution: What should come next?, Curr. Opin. Environ. Sci. Heal.

- 19 (2021) 100215. <https://doi.org/10.1016/j.coesh.2020.10.003>.
- [119] S. Tiwari, Ethylenediurea as a potential tool in evaluating ozone phytotoxicity: a review study on physiological, biochemical and morphological responses of plants, *Environ. Sci. Pollut. Res.* 24 (2017) 14019–14039. <https://doi.org/10.1007/s11356-017-8859-y>.
- [120] E. Agathokleous, Perspectives for elucidating the ethylenediurea (EDU) mode of action for protection against O₃ phytotoxicity, *Ecotoxicol. Environ. Saf.* 142 (2017) 530–537. <https://doi.org/10.1016/j.ecoenv.2017.04.057>.
- [121] K. Lakaew, S. Akeprathumchai, P. Thiravetyan, Foliar spraying of calcium acetate alleviates yield loss in rice (*Oryza sativa* L.) by induced anti-oxidative defence system under ozone and heat stresses, *Ann. Appl. Biol.* 178 (2021) 414–426. <https://doi.org/10.1111/aab.12653>.
- [122] E. Salvatori, L. Fusaro, F. Manes, Effects of the antiozonant ethylenediurea (EDU) on *Fraxinus ornus* L.: The role of drought, *Forests*. 8 (2017) 320. <https://doi.org/10.3390/f8090320>.
- [123] U. Mina, K. Smiti, P. Yadav, Thermotolerant wheat cultivar (*Triticum aestivum* L. var. WR544) response to ozone, EDU, and particulate matter interactive exposure, *Environ. Monit. Assess.* 2021 1936. 193 (2021) 1–16. <https://doi.org/10.1007/s10661-021-09079-x>.
- [124] S. Surabhi, V. Pande, V. Pandey, Ethylenediurea (EDU) mediated protection from ambient ozone-induced oxidative stress in wheat (*Triticum aestivum* L.) under a high CO₂ environment, *Atmos. Pollut. Res.* 13 (2022) 101503. <https://doi.org/10.1016/j.apr.2022.101503>.
- [125] R. Kannaujia, P. Singh, V. Prasad, V. Pandey, Evaluating impacts of biogenic silver nanoparticles and ethylenediurea on wheat (*Triticum aestivum* L.) against ozone-induced damages, *Environ. Res.* 203 (2022) 111857. <https://doi.org/10.1016/j.envres.2021.111857>.
- [126] V. Picchi, S. Gobbi, M. Fattizzo, M. Zefelippo, F. Faoro, Chitosan nanoparticles loaded with N-acetyl cysteine to mitigate ozone and other possible oxidative stresses in durum wheat, *Plants*. 10 (2021) 691. <https://doi.org/10.3390/plants10040691>.
- [127] O.I. Yakhin, A.A. Lubyantsev, I.A. Yakhin, P.H. Brown, Biostimulants in plant science: A global perspective, *Front. Plant Sci.* 7 (2017) 2049. <https://doi.org/10.3389/fpls.2016.02049>.
- [128] P.L. Godínez-Mendoza, A.K. Rico-Chávez, N.I. Ferrusquía-Jimenez, I.A. Carbajal-Valenzuela, A.I. Villagómez-Aranda, I. Torres-Pacheco, R.G. Guevara-González, Plant hormesis: Revising of the concepts of biostimulation, elicitation and their application in a sustainable agricultural production, *Sci. Total Environ.* 894 (2023) 164883. <https://doi.org/10.1016/j.scitotenv.2023.164883>.
- [129] J. Majkowska-Gadomska, A. Dobrowolski, K.K. Jadwisieńczyk, Z. Kaliniewicz, A. Francke, Effect of biostimulants on the growth, yield and nutritional value of *Capsicum annuum* grown in an unheated plastic tunnel, *Sci. Reports* 2021 111. 11 (2021) 1–14. <https://doi.org/10.1038/s41598-021-01834-x>.
- [130] P. Calvo, L. Nelson, J.W. Kloepper, Agricultural uses of plant biostimulants, *Plant Soil*.

- 999 383 (2014) 3–41. <https://doi.org/10.1007/s11104-014-2131-8>.
- 1000 [131] L. Nepthali, L.A. Piater, I.A. Dubery, V. Patterson, J. Huyser, K. Burgess, F. Tugizimana,
1001 Biostimulants for plant growth and mitigation of abiotic stresses: A metabolomics
1002 perspective, *Metabolites*. 10 (2020) 505. <https://doi.org/10.3390/metabo10120505>.
- 1003 [132] M.H. Shahrajabian, C. Chaski, N. Polyzos, S.A. Petropoulos, Biostimulants application: A
1004 low input cropping management tool for sustainable farming of vegetables, *Biomolecules*.
1005 11 (2021) 698. <https://doi.org/10.3390/biom11050698>.
- 1006 [133] S. Macias-Benitez, S. Navarro-Torre, P. Caballero, L. Martín, E. Revilla, A. Castaño, J.
1007 Parrado, Biostimulant capacity of an enzymatic extract from rice bran against ozone-
1008 induced damage in *Capsicum annum*, *Front. Plant Sci.* 12 (2021) 2573.
1009 <https://doi.org/10.3389/fpls.2021.749422>.
- 1010 [134] A. Vannini, R. Fedeli, M. Guarnieri, S. Loppi, Foliar application of wood distillate
1011 alleviates ozone-induced damage in lettuce (*Lactuca sativa* L.), *Toxics*. 10 (2022) 178.
1012 <https://doi.org/10.3390/toxics10040178>.
- 1013 [135] L. Liu, X. Zhao, Y. Huang, L. Ke, R. Wang, G. Qi, Protecting tobacco plants from O₃
1014 injury by *Bacillus velezensis* with production of acetoin, *Physiol. Plant.* 170 (2020) 158–
1015 171. <https://doi.org/10.1111/ppl.13120>.
- 1016 [136] E. Agathokleous, B. Zhou, J. Xu, A. Ioannou, Z. Feng, C.J. Saitanis, M. Frei, E.J.
1017 Calabrese, V. Fotopoulos, Exogenous application of melatonin to plants, algae, and
1018 harvested products to sustain agricultural productivity and enhance nutritional and
1019 nutraceutical value: A meta-analysis, *Environ. Res.* 200 (2021) 111746.
1020 <https://doi.org/10.1016/j.envres.2021.111746>.
- 1021 [137] M.B. Arnao, J. Hernández-Ruiz, Functions of melatonin in plants: a review, *J. Pineal Res.*
1022 59 (2015) 133–150. <https://doi.org/10.1111/jpi.12253>.
- 1023 [138] M.B. Arnao, J. Hernández-Ruiz, Melatonin as a plant biostimulant in crops and during
1024 post-harvest: a new approach is needed, *J. Sci. Food Agric.* 101 (2021) 5297–5304.
1025 <https://doi.org/10.1002/jsfa.11318>.
- 1026 [139] B. Mauch-Mani, I. Baccelli, E. Luna, V. Flors, Defense priming: An adaptive part of
1027 induced resistance, *Annu. Rev. Plant Biol.* 68 (2017) 485–512.
1028 <https://doi.org/10.1146/annurev-arplant-042916-041132>.
- 1029 [140] S.M. Westman, K.J. Kloth, J. Hanson, A.B. Ohlsson, B.R. Albrechtsen, Defence priming in
1030 *Arabidopsis* – a meta-analysis, *Sci. Rep.* 9 (2019) 13309. <https://doi.org/10.1038/s41598-019-49811-9>.
- 1031
- 1032 [141] A. Christou, E. Agathokleous, V. Fotopoulos, Safeguarding food security: Hormesis-
1033 based plant priming to the rescue, *Curr. Opin. Environ. Sci. Heal.* 28 (2022) 100374.
1034 <https://doi.org/10.1016/j.coesh.2022.100374>.
- 1035 [142] A. Savvides, S. Ali, M. Tester, V. Fotopoulos, Chemical priming of plants against
1036 multiple abiotic stresses: Mission possible, *Trends Plant Sci.* 21 (2016) 329–340.
1037 <https://doi.org/10.1016/j.tplants.2015.11.003>.

- 1038 [143] P. Acharya, G.K. Jayaprakash, K.M. Crosby, J.L. Jifon, B.S. Patil, Nanoparticle-
 1039 mediated seed priming improves germination, growth, yield, and quality of watermelons
 1040 (*Citrullus lanatus*) at multi-locations in Texas, *Sci. Rep.* 10 (2020) 5037.
 1041 <https://doi.org/10.1038/s41598-020-61696-7>.
- 1042 [144] M. Farooq, M. Hussain, M.M. Habib, M.S. Khan, I. Ahmad, S. Farooq, K.H.M. Siddique,
 1043 Influence of seed priming techniques on grain yield and economic returns of bread wheat
 1044 planted at different spacings, *Crop Pasture Sci.* 71 (2020) 725–738.
 1045 <https://doi.org/10.1071/cp20065>.
- 1046 [145] G. Sime, J.B. Aune, On-farm seed priming and fertilizer micro-dosing: Agronomic and
 1047 economic responses of maize in semi-arid Ethiopia, *Food Energy Secur.* 9 (2020) e190.
 1048 <https://doi.org/10.1002/fes3.190>.
- 1049 [146] A. Atefi, Y. Ge, S. Pitla, J. Schnable, Robotic technologies for high-throughput plant
 1050 phenotyping: Contemporary reviews and future perspectives, *Front. Plant Sci.* 12 (2021)
 1051 1082. <https://doi.org/10.3389/fpls.2021.611940>.
- 1052 [147] A. Balafoutis, B. Beck, S. Fountas, J. Vangeyte, T. Van Der Wal, I. Soto, M. Gómez-
 1053 Barbero, A. Barnes, V. Eory, Precision agriculture technologies positively contributing to
 1054 GHG emissions mitigation, farm productivity and economics, *Sustainability.* 9 (2017)
 1055 1339. <https://doi.org/10.3390/su9081339>.
- 1056 [148] M. Morisse, D.M. Wells, E.J. Millet, M. Lillemo, S. Fahrner, F. Cellini, P. Lootens, O.
 1057 Muller, J.M. Herrera, A.R. Bentley, M. Janni, A European perspective on opportunities
 1058 and demands for field-based crop phenotyping, *F. Crop. Res.* 276 (2022) 108371.
 1059 <https://doi.org/10.1016/j.fcr.2021.108371>.
- 1060 [149] M. Watt, F. Fiorani, B. Usadel, U. Rascher, O. Muller, U. Schurr, Phenotyping: New
 1061 windows into the plant for breeders, *Annu. Rev. Plant Biol.* 71 (2020) 689–712.
 1062 <https://doi.org/10.1146/annurev-arplant-042916-041124>.
- 1063 [150] S. Liu, C. Ji, C. Wang, J. Chen, Y. Jin, Z. Zou, S. Li, S. Niu, J. Zou, Climatic role of
 1064 terrestrial ecosystem under elevated CO₂ : a bottom-up greenhouse gases budget, *Ecol.*
 1065 *Lett.* 21 (2018) 1108–1118. <https://doi.org/10.1111/ele.13078>.
- 1066 [151] K.J. Van Groenigen, X. Qi, C.W. Osenberg, Y. Luo, B.A. Hungate, Faster decomposition
 1067 under increased atmospheric CO₂ limits soil carbon storage, *Science.* 344 (2014) 508–509.
 1068 <https://doi.org/10.1126/science.1249534>.
- 1069 [152] L. Cheng, N. Zhang, M. Yuan, J. Xiao, Y. Qin, Y. Deng, Q. Tu, K. Xue, J.D. Van
 1070 Nostrand, L. Wu, Z. He, X. Zhou, M.B. Leigh, K.T. Konstantinidis, E.A.G. Schuur, Y.
 1071 Luo, J.M. Tiedje, J. Zhou, Warming enhances old organic carbon decomposition through
 1072 altering functional microbial communities, *ISME J.* 11 (2017) 1825.
 1073 <https://doi.org/10.1038/ismej.2017.48>.
- 1074 [153] Y. Qiu, L. Guo, X. Xu, L. Zhang, K. Zhang, M. Chen, Y. Zhao, K.O. Burkey, H.D. Shew,
 1075 R.W. Zobel, Y. Zhang, S. Hu, Warming and elevated ozone induce tradeoffs between fine
 1076 roots and mycorrhizal fungi and stimulate organic carbon decomposition, *Sci. Adv.* 7
 1077 (2021) abe9256. <https://doi.org/10.1126/sciadv.abe9256>.

- 1078 [154] M.W.I. Schmidt, M.S. Torn, S. Abiven, T. Dittmar, G. Guggenberger, I.A. Janssens, M.
1079 Kleber, I. Kögel-Knabner, J. Lehmann, D.A.C. Manning, P. Nannipieri, D.P. Rasse, S.
1080 Weiner, S.E. Trumbore, Persistence of soil organic matter as an ecosystem property,
1081 *Nature*. 478 (2011) 49–56. <https://doi.org/10.1038/nature10386>.
- 1082 [155] E.E. Oldfield, M.A. Bradford, S.A. Wood, Global meta-analysis of the relationship
1083 between soil organic matter and crop yields, *SOIL*. 5 (2019) 15–32.
1084 <https://doi.org/10.5194/soil-5-15-2019>.
- 1085 [156] R. Crystal-Ornelas, R. Thapa, K.L. Tully, Soil organic carbon is affected by organic
1086 amendments, conservation tillage, and cover cropping in organic farming systems: A
1087 meta-analysis, *Agric. Ecosyst. Environ.* 312 (2021) 107356.
1088 <https://doi.org/10.1016/j.agee.2021.107356>.
- 1089 [157] M. Diacono, F. Montemurro, Long-term effects of organic amendments on soil fertility. A
1090 review, *Agron. Sustain. Dev.* 30 (2010) 401–422. <https://doi.org/10.1051/agro/2009040>.
- 1091 [158] E. Aguilera, C. Díaz-Gaona, R. García-Laureano, C. Reyes-Palomo, G.I. Guzmán, L.
1092 Ortolani, M. Sánchez-Rodríguez, V. Rodríguez-Estévez, Agroecology for adaptation to
1093 climate change and resource depletion in the Mediterranean region. A review, *Agric. Syst.*
1094 181 (2020) 102809. <https://doi.org/10.1016/j.agsy.2020.102809>.
- 1095 [159] A. Damm, S. Cogliati, R. Colombo, L. Fritsche, A. Genangeli, L. Genesio, J. Hanus, A.
1096 Peressotti, P. Rademske, U. Rascher, D. Schuettemeyer, B. Siegmann, J. Sturm, F.
1097 Miglietta, Response times of remote sensing measured sun-induced chlorophyll
1098 fluorescence, surface temperature and vegetation indices to evolving soil water limitation
1099 in a crop canopy, *Remote Sens. Environ.* 273 (2022) 112957.
1100 <https://doi.org/10.1016/j.rse.2022.112957>.
- 1101 [160] M. Drusch, J. Moreno, U. Del Bello, R. Franco, Y. Goulas, A. Huth, S. Kraft, E.M.
1102 Middleton, F. Miglietta, G. Mohammed, L. Nedbal, U. Rascher, D. Schuettemeyer, W.
1103 Verhoef, The FLuorescence EXplorer Mission Concept-ESA's Earth Explorer 8, *IEEE*
1104 *Trans. Geosci. Remote Sens.* 55 (2017) 1273–1284.
1105 <https://doi.org/10.1109/tgrs.2016.2621820>.
- 1106 [161] S. Kumar, R.S. Meena, S. Sheoran, C.K. Jangir, M.K. Jhariya, A. Banerjee, A. Raj,
1107 Remote sensing for agriculture and resource management, in: M.K. Jhariya, R.S. Meena,
1108 A. Banerjee, S.N. Meena (Eds.), *Nat. Resour. Conserv. Adv. Sustain.*, Elsevier, 2022: pp.
1109 91–135. <https://doi.org/10.1016/b978-0-12-822976-7.00012-0>.
- 1110 [162] P. Alexander, A. Arneth, R. Henry, J. Maire, S. Rabin, M.D.A. Rounsevell, High energy
1111 and fertilizer prices are more damaging than food export curtailment from Ukraine and
1112 Russia for food prices, health and the environment, *Nat. Food*. 4 (2022) 84–95.
1113 <https://doi.org/10.1038/s43016-022-00659-9>.
- 1114 [163] D. Fowler, K. Pilegaard, M.A. Sutton, P. Ambus, M. Raivonen, J. Duyzer, D. Simpson, H.
1115 Fagerli, S. Fuzzi, J.K. Schjoerring, C. Granier, A. Neftel, I.S.A. Isaksen, P. Laj, M.
1116 Maione, P.S. Monks, J. Burkhardt, U. Daemmgen, J. Neirynck, E. Personne, R. Wichink-
1117 Kruit, K. Butterbach-Bahl, C. Flechard, J.P. Tuovinen, M. Coyle, G. Gerosa, B. Loubet, N.
1118 Altimir, L. Gruenhage, C. Ammann, S. Cieslik, E. Paoletti, T.N. Mikkelsen, H. Ro-

- 1119 Poulsen, P. Cellier, J.N. Cape, L. Horváth, F. Loreto, Ü. Niinemets, P.I. Palmer, J. Rinne,
1120 P. Misztal, E. Nemitz, D. Nilsson, S. Pryor, M.W. Gallagher, T. Vesala, U. Skiba, N.
1121 Brüggemann, S. Zechmeister-Boltenstern, J. Williams, C. O'Dowd, M.C. Facchini, G. de
1122 Leeuw, A. Flossman, N. Chaumerliac, J.W. Erisman, Atmospheric composition change:
1123 Ecosystems–atmosphere interactions, *Atmos. Environ.* 43 (2009) 5193–5267.
1124 <https://doi.org/10.1016/j.atmosenv.2009.07.068>.
- 1125 [164] G. Gerosa, A. Finco, S. Mereu, M. Vitale, F. Manes, A.B. Denti, Comparison of seasonal
1126 variations of ozone exposure and fluxes in a Mediterranean Holm oak forest between the
1127 exceptionally dry 2003 and the following year, *Environ. Pollut.* 157 (2009) 1737–1744.
1128 <https://doi.org/10.1016/j.envpol.2007.11.025>.
- 1129 [165] D.B. Lobell, C.J. Bonfils, L.M. Kueppers, M.A. Snyder, Irrigation cooling effect on
1130 temperature and heat index extremes, *Geophys. Res. Lett.* 35 (2008) 9705.
1131 <https://doi.org/10.1029/2008gl034145>.
- 1132 [166] E. Vogel, M.G. Donat, L. V. Alexander, M. Meinshausen, D.K. Ray, D. Karoly, N.
1133 Meinshausen, K. Frieler, The effects of climate extremes on global agricultural yields,
1134 *Environ. Res. Lett.* 14 (2019) 054010. <https://doi.org/10.1088/1748-9326/ab154b>.
- 1135 [167] E. Eyshi Rezaei, H. Webber, T. Gaiser, J. Naab, F. Ewert, Heat stress in cereals:
1136 Mechanisms and modelling, *Eur. J. Agron.* 64 (2015) 98–113.
1137 <https://doi.org/10.1016/j.eja.2014.10.003>.
- 1138 [168] B.M. McGill, S.K. Hamilton, | Neville Millar, | G Philip Robertson, W.K. Kellogg, W.K.
1139 Kellogg Biological, K.B. Survey, The greenhouse gas cost of agricultural intensification
1140 with groundwater irrigation in a Midwest U.S. row cropping system, *Glob. Chang. Biol.*
1141 24 (2018) 5948–5960. <https://doi.org/10.1111/gcb.14472>.
- 1142 [169] M.L. Cayuela, E. Aguilera, A. Sanz-Cobena, D.C. Adams, D. Abalos, L. Barton, R. Ryals,
1143 W.L. Silver, M.A. Alfaro, V.A. Pappa, P. Smith, J. Garnier, G. Billen, L. Bouwman, A.
1144 Bondeau, L. Lassaletta, Direct nitrous oxide emissions in Mediterranean climate cropping
1145 systems: Emission factors based on a meta-analysis of available measurement data, *Agric.*
1146 *Ecosyst. Environ.* 238 (2017) 25–35. <https://doi.org/10.1016/j.agee.2016.10.006>.
- 1147 [170] J. Li, A. Mahalov, P. Hyde, Impacts of agricultural irrigation on ozone concentrations in
1148 the Central Valley of California and in the contiguous United States based on WRF-Chem
1149 simulations, *Agric. For. Meteorol.* 221 (2016) 34–49.
1150 <https://doi.org/10.1016/j.agrformet.2016.02.004>.
- 1151 [171] R.S. Massad, J. Lathière, S. Strada, M. Perrin, E. Personne, M. Stéfanon, P. Stella, S.
1152 Szopa, N. De Noblet-Ducoudré, Reviews and syntheses: Influences of landscape structure
1153 and land uses on local to regional climate and air quality, *Biogeosciences.* 16 (2019)
1154 2369–2408. <https://doi.org/10.5194/bg-16-2369-2019>.
- 1155 [172] T. Gleeson, L. Wang-Erlandsson, S.C. Zipper, M. Porkka, F. Jaramillo, D. Gerten, I.
1156 Fetzer, S.E. Cornell, L. Piemontese, L.J. Gordon, J. Rockström, T. Oki, M. Sivapalan, Y.
1157 Wada, K.A. Brauman, M. Flörke, M.F.P. Bierkens, B. Lehner, P. Keys, M. Kummu, T.
1158 Wagener, S. Dadson, T.J. Troy, W. Steffen, M. Falkenmark, J.S. Famiglietti, The water
1159 planetary boundary: Interrogation and revision, *One Earth.* 2 (2020) 223–234.

1160 <https://doi.org/10.1016/j.oneear.2020.02.009>.

1161 [173] R.P. Neupane, S. Kumar, Estimating the effects of potential climate and land use changes
1162 on hydrologic processes of a large agriculture dominated watershed, *J. Hydrol.* 529 (2015)
1163 418–429. <https://doi.org/10.1016/j.jhydrol.2015.07.050>.

1164 [174] M.N. Chisola, M. van der Laan, K.L. Bristow, A landscape hydrology approach to inform
1165 sustainable water resource management under a changing environment. A case study for
1166 the Kaleya River Catchment, Zambia, *J. Hydrol. Reg. Stud.* 32 (2020) 100762.
1167 <https://doi.org/10.1016/j.ejrh.2020.100762>.

1168 [175] M.G. Arenas-Corraliza, M.L. López-Díaz, G. Moreno, Winter cereal production in a
1169 Mediterranean silvoarable walnut system in the face of climate change, *Agric. Ecosyst.*
1170 *Environ.* 264 (2018) 111–118. <https://doi.org/10.1016/j.agee.2018.05.024>.

1171 [176] M. Kanzler, C. Böhm, J. Mirck, D. Schmitt, M. Veste, Microclimate effects on
1172 evaporation and winter wheat (*Triticum aestivum* L.) yield within a temperate agroforestry
1173 system, *Agrofor. Syst.* 93 (2018) 1821–1841. <https://doi.org/10.1007/s10457-018-0289-4>.

1174 [177] H.H. Liste, J.C. White, Plant hydraulic lift of soil water – implications for crop production
1175 and land restoration, *Plant Soil.* 313 (2008) 1–17. [https://doi.org/10.1007/s11104-008-](https://doi.org/10.1007/s11104-008-9696-z)
1176 [9696-z](https://doi.org/10.1007/s11104-008-9696-z).

1177 [178] IPCC, Climate Change 2014: Synthesis Report; Chapter Observed Changes and their
1178 Causes, 2014. https://www.ipcc.ch/site/assets/uploads/2018/02/ar5_syr_final_spm.pdf.

1179 [179] R. Bezner Kerr, T. Hasegawa, R. Lasco, I. Bhatt, D. Deryng, A. Farrell, H. Gurney-Smith,
1180 H. Ju, S. Lluch-Cota, F. Meza, G. Nelson, H. Neufeldt, P. Thornton, Chapter 5: Food,
1181 Fibre and Other Ecosystem Products, in: H.-O. Pörtner, D.C. Roberts, M. Tignor, E.S.
1182 Poloczanska, K. Mintenbeck, A. Alegría, M. Craig, S. Langsdorf, S. Löschke, V. Möller,
1183 A. Okem, B. Rama (Eds.), *Clim. Chang. 2022 Impacts, Adapt. Vulnerability. Contribution*
1184 *Work. Gr. II to Sixth Assess. Rep. Intergov. Panel Clim. Chang., Intergovernmental Panel*
1185 *on Climate Change (IPCC), 2022*. <https://www.ipcc.ch/report/ar6/wg2/>.

1186 [180] G. Mills, K. Sharps, D. Simpson, H. Pleijel, M. Broberg, J. Uddling, F. Jaramillo, W.J.
1187 Davies, F. Dentener, M. Van den Berg, M. Agrawal, S.B. Agrawal, E.A. Ainsworth, P.
1188 Büker, L. Emberson, Z. Feng, H. Harmens, F. Hayes, K. Kobayashi, E. Paoletti, R. Van
1189 Dingenen, Ozone pollution will compromise efforts to increase global wheat production,
1190 *Glob. Chang. Biol.* 24 (2018) 3560–3574. <https://doi.org/10.1111/gcb.14157>.

1191 [181] L.D. Schiferl, C.L. Heald, Particulate matter air pollution may offset ozone damage to
1192 global crop production, *Atmos. Chem. Phys.* 18 (2018) 5953–5966.
1193 <https://doi.org/10.5194/acp-18-5953-2018>.

1194 [182] K. Chenu, J.R. Porter, P. Martre, B. Basso, S.C. Chapman, F. Ewert, M. Bindi, S. Asseng,
1195 Contribution of crop models to adaptation in wheat, *Trends Plant Sci.* 22 (2017) 472–490.
1196 <https://doi.org/10.1016/j.tplants.2017.02.003>.

1197 [183] S. Asseng, F. Ewert, P. Martre, R.P. Rötter, D.B. Lobell, D. Cammarano, B.A. Kimball,
1198 M.J. Ottman, G.W. Wall, J.W. White, M.P. Reynolds, P.D. Alderman, P.V.V. Prasad, P.K.
1199 Aggarwal, J. Anothai, B. Basso, C. Biernath, A.J. Challinor, G. De Sanctis, J. Doltra, E.

1200 Fereres, M. Garcia-Vila, S. Gayler, G. Hoogenboom, L.A. Hunt, R.C. Izaurralde, M.
1201 Jabloun, C.D. Jones, K.C. Kersebaum, A.K. Koehler, C. Müller, S. Naresh Kumar, C.
1202 Nendel, G. O’leary, J.E. Olesen, T. Palosuo, E. Priesack, E. Eyshi Rezaei, A.C. Ruane,
1203 M.A. Semenov, I. Shcherbak, C. Stöckle, P. Stratonovitch, T. Streck, I. Supit, F. Tao, P.J.
1204 Thorburn, K. Waha, E. Wang, D. Wallach, J. Wolf, Z. Zhao, Y. Zhu, Rising temperatures
1205 reduce global wheat production, *Nat. Clim. Chang.* 5 (2014) 143–147.
1206 <https://doi.org/10.1038/nclimate2470>.

1207 [184] H. Tang, M. Takigawa, G. Liu, J. Zhu, K. Kobayashi, A projection of ozone-induced
1208 wheat production loss in China and India for the years 2000 and 2020 with exposure-
1209 based and flux-based approaches, *Glob. Chang. Biol.* 19 (2013) 2739–2752.
1210 <https://doi.org/10.1111/gcb.12252>.

1211 [185] C. Rosenzweig, J.W. Jones, J.L. Hatfield, A.C. Ruane, K.J. Boote, P. Thorburn, J.M.
1212 Antle, G.C. Nelson, C. Porter, S. Janssen, S. Asseng, B. Basso, F. Ewert, D. Wallach, G.
1213 Baigorria, J.M. Winter, The Agricultural Model Intercomparison and Improvement
1214 Project (AgMIP): Protocols and pilot studies, *Agric. For. Meteorol.* 170 (2013) 166–182.
1215 <https://doi.org/10.1016/j.agrformet.2012.09.011>.

1216 [186] E. Wang, P. Martre, Z. Zhao, F. Ewert, A. Maiorano, R.P. Rötter, B.A. Kimball, M.J.
1217 Ottman, G.W. Wall, J.W. White, M.P. Reynolds, P.D. Alderman, P.K. Aggarwal, J.
1218 Anothai, B. Basso, C. Biernath, D. Cammarano, A.J. Challinor, G. De Sanctis, J. Doltra, E.
1219 Fereres, M. Garcia-Vila, S. Gayler, G. Hoogenboom, L.A. Hunt, R.C. Izaurralde, M.
1220 Jabloun, C.D. Jones, K.C. Kersebaum, A.K. Koehler, L. Liu, C. Müller, S. Naresh Kumar,
1221 C. Nendel, G. O’Leary, J.E. Olesen, T. Palosuo, E. Priesack, E. Eyshi Rezaei, D. Ripoche,
1222 A.C. Ruane, M.A. Semenov, I. Shcherbak, C. Stöckle, P. Stratonovitch, T. Streck, I. Supit,
1223 F. Tao, P. Thorburn, K. Waha, D. Wallach, Z. Wang, J. Wolf, Y. Zhu, S. Asseng, The
1224 uncertainty of crop yield projections is reduced by improved temperature response
1225 functions, *Nat. Plants.* 3 (2017) 17102. <https://doi.org/10.1038/nplants.2017.102>.

1226 [187] M.C. Broberg, Z. Feng, Y. Xin, H. Pleijel, Ozone effects on wheat grain quality- A
1227 summary, *Environ. Pollut.* 197 (2015) 203–213.
1228 <https://doi.org/10.1016/j.envpol.2014.12.009>.

1229 [188] L.D. Emberson, H. Pleijel, E.A. Ainsworth, M. van den Berg, W. Ren, S. Osborne, G.
1230 Mills, D. Pandey, F. Dentener, P. Büker, F. Ewert, R. Koeble, R. Van Dingenen, Ozone
1231 effects on crops and consideration in crop models, *Eur. J. Agron.* 100 (2018) 19–34.
1232 <https://doi.org/10.1016/j.eja.2018.06.002>.

1233 [189] I. Droutsas, A.J. Challinor, S.R. Arnold, T.N. Mikkelsen, E.M.Ø. Hansen, A new model of
1234 ozone stress in wheat including grain yield loss and plant acclimation to the pollutant, *Eur.*
1235 *J. Agron.* 120 (2020) 126125. <https://doi.org/10.1016/j.eja.2020.126125>.

1236 [190] J.R. Guarin, B. Kassie, A.M. Mashaeet, K. Burkey, S. Asseng, Modeling the effects of
1237 tropospheric ozone on wheat growth and yield, *Eur. J. Agron.* 105 (2019) 13–23.
1238 <https://doi.org/10.1016/j.eja.2019.02.004>.

1239 [191] E. Agathokleous, R.G. Belz, V. Calatayud, A. De Marco, Y. Hoshika, M. Kitao, C.J.
1240 Saitanis, P. Sicard, E. Paoletti, E.J. Calabrese, Predicting the effect of ozone on vegetation
1241 via linear non-threshold (LNT), threshold and hormetic dose-response models, *Sci. Total*

- Environ. 649 (2019) 61–74. <https://doi.org/10.1016/j.scitotenv.2018.08.264>.
- [192] R.L. Heath, A.S. Lefohn, R.C. Musselman, Temporal processes that contribute to nonlinearity in vegetation responses to ozone exposure and dose, *Atmos. Environ.* 43 (2009) 2919–2928. <https://doi.org/10.1016/j.atmosenv.2009.03.011>.
- [193] S. Mohammadi, K. Rydgren, V. Bakkestuen, M.A.K. Gillespie, Impacts of recent climate change on crop yield can depend on local conditions in climatically diverse regions of Norway, *Sci. Rep.* 13 (2023) 3633. <https://doi.org/10.1038/s41598-023-30813-7>.
- [194] M.A. Altieri, C.I. Nicholls, The adaptation and mitigation potential of traditional agriculture in a changing climate, *Clim. Change.* 140 (2017) 33–45. <https://doi.org/10.1007/s10584-013-0909-y>.
- [195] Q. Schiermeier, Eat less meat: UN climate-change report calls for change to human diet, *Nature.* 572 (2019) 291–292. <https://doi.org/10.1038/d41586-019-02409-7>.
- [196] X. Liu, A.P.K. Tai, Y. Chen, L. Zhang, G. Shaddick, X. Yan, H.M. Lam, Dietary shifts can reduce premature deaths related to particulate matter pollution in China, *Nat. Food.* 2 (2021) 997–1004. <https://doi.org/10.1038/s43016-021-00430-6>.
- [197] L. Xia, X. Yan, Maximizing Earth’s feeding capacity, *Nat. Food.* 4 (2023) 353–354. <https://doi.org/10.1038/s43016-023-00736-7>.
- [198] E.L.F. Schipper, N.K. Dubash, Y. Mulugetta, Climate change research and the search for solutions: rethinking interdisciplinarity, *Clim. Change.* 168 (2021) 18. <https://doi.org/10.1007/s10584-021-03237-3>.
- [199] D.J. Lang, A. Wiek, M. Bergmann, M. Stauffacher, P. Martens, P. Moll, M. Swilling, C.J. Thomas, Transdisciplinary research in sustainability science: Practice, principles, and challenges, *Sustain. Sci.* 7 (2012) 25–43. <https://doi.org/10.1007/s11625-011-0149-x>.

Figures and captions

Fig. 1. Adaptation of crop production to air pollution and climate change at plant, field, and ecosystem levels. At plant scale, integrating genetic variation, QTL, molecular breeding, and phenotyping can offer a perspective for improving crop performance under such environmental challenges. Then, a number of techniques can be applied on the field, including adjusting cultivation practices (e.g. fertilization, irrigation), diversifying cultivations and promoting mixed cropping systems, applying antiozonants (if toxicologically tested) and biostimulants, utilizing priming technology, and expanding phenotyping-robot-assisted farming. At the ecosystem level, enhancing soil resilience, incorporating remote sensing technology (precision agriculture), and modifying the hydrology and microclimate of agricultural landscapes can further facilitate crop adaptation to climate change and air pollution. Photo sources: Background photo: ©Mokhammad Edliadi (flickr.com; cifor.org); Puzzle pieces: ©Pixabay.com; istockphoto.com.

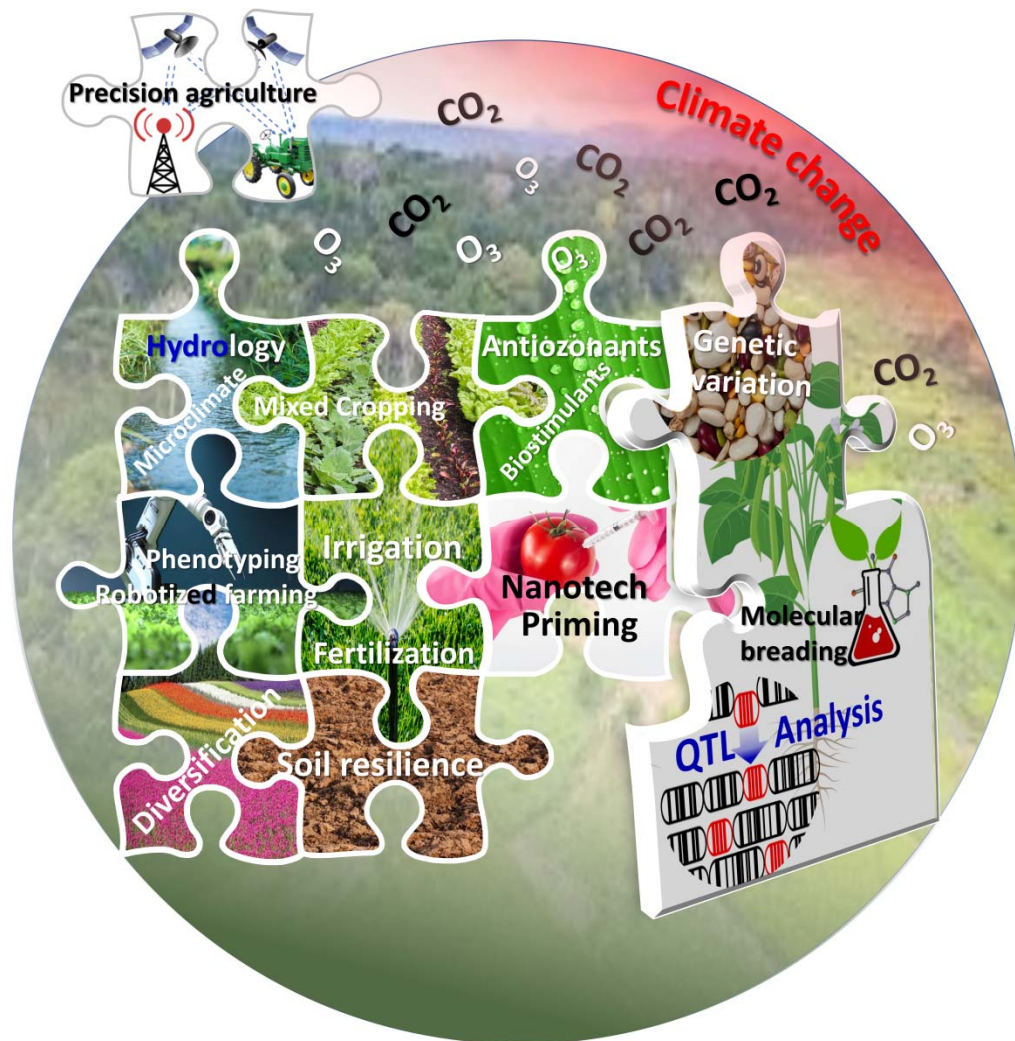


Fig. 2. Overcoming current limitations in adapting agroecosystems to climate change and air pollution at different scales. To overcome current limitations, needs include moving toward more multi-dimensional experiments, employing a transdisciplinary approach, and addressing combined effects of climate change and air pollution in models. Needs also include coupling modeling with empirical data, incorporating pleiotropic responses and non-linear phenomena in modeling and adaptation programs, applying growth models at regional scale, and targeting region-specific adaptation.

