

Sensing the Dynamic Response of Photosynthesis to Abiotic Stressors with Forced Oscillating Light

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Introduction and Rationale

- ❖ In nature, solar irradiance is characterized by **variable fluctuations** in time and intensity.
- ❖ Photosynthetic organisms evolved a range of **adaptive mechanisms** to cope with these **dynamic light environments**.
- ❖ The response of photosynthesis is typically studied in **steady-state** conditions, such as in **constant light** and with **dark - light** or **low light - high light** transitions.

- ❖ To systematically investigate photosynthesis in **non-steady-state** conditions, a frequency-domain chlorophyll fluorescence (**ChIF**)^{[1],[2]} measurement protocol was established. Actinic light was modulated as a **harmonic function** at **selected frequencies**, generating **oscillating (sinusoidal) light**.
- ❖ This method was implemented to measure the photosynthesis response in plants exposed to **High Temperature (HT)** or **High Light (HL)**.
- ❖ **Advantage:** This method allows to detect fingerprints of dynamic acclimation in **light-adapted** plants (**no need for dark-adaptation**).

Materials and Methods

Harmonic modulation

Arabidopsis thaliana (Columbia-0)

Fast Fourier Transform (FFT)

$$ReX[k] = \sum_{i=0}^{N-1} x[i] \cos(2\pi k i / N)$$
$$ImX[k] = - \sum_{i=0}^{N-1} x[i] \sin(2\pi k i / N)$$

E_k determination

FFT components

Frequency scan

DREAM Macroscope

- ❖ **Control:** irradiance $\approx 80\text{--}90 \mu\text{E m}^{-2} \text{s}^{-1}$; air temperature $\approx 22\text{--}24^\circ\text{C}$ (light) and $\approx 16\text{--}17^\circ\text{C}$ (dark); leaf temperature $\approx 22\text{--}24^\circ\text{C}$ during measurements.
- ❖ **HT:** irradiance $\approx 100\text{--}110 \mu\text{E m}^{-2} \text{s}^{-1}$; air temperature $\approx 41\text{--}42^\circ\text{C}$ (light) and $\approx 25\text{--}26^\circ\text{C}$ (dark), with intermediate steps at $\approx 36\text{--}38^\circ\text{C}$; leaf temperature $\approx 22\text{--}24^\circ\text{C}$ during measurements.
- ❖ **HL:** irradiance at $\approx 500\text{--}550 \mu\text{E m}^{-2} \text{s}^{-1}$, with intermediate steps at $\approx 250\text{--}260 \mu\text{E m}^{-2} \text{s}^{-1}$; air temperature $\approx 22\text{--}24^\circ\text{C}$ (light) and $\approx 16\text{--}17^\circ\text{C}$ (dark); leaf temperature $\approx 22\text{--}24^\circ\text{C}$ during measurements.
- ❖ Photoperiod was **12h/12h light/dark**.
- ❖ **Mature plants** were exposed to **HT** or **HL** for **three consecutive days** and measured every day at **+4 hours** in the light period.

- ❖ A novel **macroscopic fluorescence imaging system** was implemented (Dr. Ian Coghill, CNRS, Paris, France, DREAM).
- ❖ $\lambda_{\text{excitation}} = 645 \text{ nm}$; $\lambda_{\text{detection}} = 690 \text{ nm}$ (50 nm bandwidth).
- ❖ A specific illumination protocol was applied, based on the **light saturation coefficient E_k** ^[3]. $U_0 \approx 2.5 E_k$ and $U_v \approx 0.25 E_k$.
- ❖ **Frequency-scan** from $\approx 0.008 \text{ Hz}$ ($T = 128 \text{ s}$) to **4 Hz** ($T = 0.25 \text{ s}$).

Results and Conclusion

ChIF forced oscillations (time)

ChIF in the frequency domain (FFT)

ChIF in the time domain

High Temperature

High Light

Conclusion

The frequency-domain ChIF analysis can detect **frequency-specific fingerprints of stress response** in **light-adapted** plants without **dark adaptation**.

Outlook

- ❖ **Machine learning** algorithms to categorize stress responses in a **fast and reliable** manner (collaboration with **SONY CSL, DREAM**).
- ❖ Application of the method to **crop plants under relevant growth environments**.
- ❖ Exploration of **various stress scenarios**, such as nutrient deficiency, drought, biotic stress, and combination of multiple stress factors.

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[2]: Niu, Y. et al. (2023). Plants cope with fluctuating light by frequency-dependent nonphotochemical quenching and cyclic electron transport. *New Phytologist*, 239: 1869–1886. doi: 10.1111/nph.19083.
[3]: Consalvey, M. et al. (2005). PAM fluorescence: a beginners guide for benthic diatomists. *Diatom Research*, 20(1):1–22. https://doi.org/10.1080/0269249X.2005.9705619.