



Quantitative Estimation of Leaf Heat Transfer Coefficients by Active Thermography at Varying Boundary Layer Conditions

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Quantifying heat and mass exchanges processes of plant leaves is crucial for detailed understanding of dynamic plant-environment interactions. The two main components of these processes, convective heat transfer, and transpiration, are inevitably coupled as both processes are restricted by the leaf boundary layer. To measure leaf heat capacity and leaf heat transfer coefficient, we thoroughly tested and applied an active thermography method that uses a transient heat pulse to compute τ , the time constant of leaf cooling after release of the pulse. We validated our approach in the laboratory on intact leaves of spring barley (*Hordeum vulgare*) and common bean (*Phaseolus vulgaris*), and measured τ -changes at different boundary layer conditions. By modeling the leaf heat transfer coefficient with dimensionless numbers, we could demonstrate that τ improves our ability to close the energy budget of plant leaves and that modeling of transpiration requires considerations of convection. Applying our approach to thermal images we obtained spatio-temporal maps of τ , providing observations of local differences in thermal responsiveness of leaf surfaces. We propose that active thermography is an informative methodology to measure leaf heat transfer and derive spatial maps of thermal responsiveness of leaves contributing to improve models of leaf heat transfer processes.

Keywords: active thermography, time constant of cooling, CWSI, leaf transpiration, heat capacity, plant phenotyping

INTRODUCTION

Plants continuously interact with their environment by heat and mass exchange and play an important role in the earth's hydrological and carbon cycle (Foley et al., 2003). The most important physiological process resulting in gas and mass exchange with the atmosphere is photosynthesis accompanied by transpiration. Thus, heat and mass exchange between plants and their environment significantly affects plant productivity, water use, and water use efficiency (Shibuya et al., 2006; Schymanski and Or, 2015). Particularly, water use efficiency is of interest in agriculture for genetic improvement and selection of high-yielding crop varieties for water-limited agriculture (Blum,

2009; Munns et al., 2010; Passioura and Angus, 2010). Consequently, studying plants heat and mass exchange is key to understand the dynamics of plant-environment-interactions.

An important component of plant heat exchange is the convective heat transfer, the exchange of heat between the leaf surface and the surrounding atmospheric conditions. Heat penetrates the boundary layer of a leaf that is characterized by gradients of temperature, gas concentrations, and air velocities determining the boundary layer conductance (Raschke, 1960; Schuepp, 1993; Schreuder et al., 2001; Vogel, 2009). Because transpiration, i.e., heat loss via water vapor, affects leaf temperature (T_L) (Gates, 1968), and T_L also affects convective heat transfer (Dixon and Grace, 1983), convection and transpiration are inevitably coupled to each other. Additionally, transpiration is constrained by boundary layer conductance. Wind also affects transpiration rates by removing the water vapor within the boundary layer leading to a higher leaf-to-air water vapor pressure deficit and ultimately may induce stomatal closure (Grace, 1974; Dixon and Grace, 1983; Bunce, 1985).

Because transpiration, i.e., heat loss via water vapor, affects leaf temperature (T_L) (Gates, 1968), which also affects convective heat transfer (Dixon and Grace, 1983), convection and transpiration are closely coupled. Additionally, stomatal response to environmental conditions such as humidity or CO₂ concentration further affects these exchange rates (Grace, 1974; Dixon and Grace, 1983; Bunce, 1985). The boundary layer conductance cannot be measured directly and it is often approximated by heat transfer coefficients that relate heat flux densities per unit leaf area to the difference between T_L and ambient air temperature ($T_L - T_a$) (Raschke, 1960; Schuepp, 1993). Heat transfer coefficients depend largely on air speed (forced convection) or temperature differences (free convection), which can be further approximated with dimensionless numbers (Defraeye et al., 2013). Usually both free and forced convection occur under a wide range of conditions determining the heat exchange processes to a different extent depending on the prevailing conditions (Dixon and Grace, 1983; Monteith and Unsworth, 2008; Nobel, 2009).

Heat and mass exchange processes are also affected by incoming radiation and thermal conductivity. While radiation is an important factor influencing photosynthesis and transpiration (Roelfsema and Hedrich, 2005; Pieruschka et al., 2010; Mott and Peak, 2011), variation of thermal conductivity is often neglected in quantitative analyses because thermal conductivity of plant leaves is generally low (Jayalakshmy and Philip, 2010). The low thermal conductivity of plant leaves is a result of a relatively high content of water resulting in a high specific heat capacity. Water is the main component of leaves and considerably affects the leaf heat capacity. Therefore, the leaf water content per unit area (LWC), the leaf succulence, affects the dynamics of T_L response to heat absorption, which, in turn, affects the dynamics of convection, and transpiration (Gates, 1968; Dixon and Grace, 1983; Bailey and Meneses, 1993). The thermal responsiveness of a leaf, which describes to what extent and how fast a leaf heats up or cools down, depends on leaf heat capacity and leaf heat transfer coefficient (h_{leaf}), which both can be estimated by the time constant (τ) of T_L dynamics. The time constant is characterized by the dynamic response of T_L to

different environmental factors with τ as the product of leaf heat capacity and the inverse of the heat transfer coefficient (Jones, 1992; Monteith and Unsworth, 2008; Nobel, 2009).

The use of τ as a measure of the thermal responsiveness of leaves and for hleaf modeling was proposed in early studies in the 60's using the so-called "cooling curve technique" (Linacre, 1972; Pearman et al., 1972). According to this method, the temperature of leaves or artificial leaf models, is transiently increased by a short (seconds) heat pulse and the following cooling curve is recorded. This temperature cooling kinetic provides τ , which is the slope of the exponential decay curve. For example, Linacre, 1972 used wet blotting paper to estimate the heat transfer of transpiring leaves and Pearman et al. (1972) used copper discs to estimate leaf heat transfer in a canopy under field conditions. However, artificial leaves have a different thermal conductivity compared to leaf tissue. For example, the thermal conductivity of copper ($400 \text{ W m}^{-1} \text{ K}^{-1}$) is substantially greater than that of water ($0.6 \text{ W m}^{-1} \text{ K}^{-1}$) or leaves (0.2 to $0.5 \text{ W m}^{-1} \text{ K}^{-1}$) (Jayalakshmy and Philip, 2010). Large thermal conductivity also allows lateral heat conduction, which is very low in leaf tissue. Finally, the heat capacity of different materials used as artificial references is also not comparable to that of leaves leading to incorrect estimation of τ -values for real leaves.

Studies using the cooling curve technique on intact leaves were mainly performed using thermocouples or radiometers for T_L measurements (Parlange and Waggoner, 1971; Saldin and Barthakur, 1971; Pearman et al., 1972). Assessment of T_L by using thermocouples is not fully non-invasive, because thermocouples are attached to the leaf surface (Kumar and Barthakur, 1971) and therefore may affect the leaf heat transfer. Radiometers may be preferable, because these sensors are not attached to the leaf and therefore do not disturb the leaf heat transfer. However, radiometers do not provide spatial information of T_L . Recently, non-invasive thermal imaging has become a powerful alternative to point measurements. To our knowledge, there is only one study in which τ was derived by using thermography on plant leaves. Similar to the cooling curve technique Kümmerlen et al. (1999) used the "active thermography" approach and measured intact leaves enclosed in a gas exchange cuvette. These authors were able to derive LWC from τ -measurements and hleaf from gas exchange measurements in *Ricinus communis* plants.

To model leaf heat transfer processes in leaves the implementation of τ is very relevant (Leigh et al., 2012). We have revisited this type of measurements using an active thermography protocol applying short infra-red radiation pulses and tested the thermal responsiveness of contrasting leaf types (i.e., leaf structure, vascular tissue) of spring barley (*Hordeum vulgare*) and common bean (*Phaseolus vulgaris*). We modeled hleaf response to wind and varying irradiance by using τ and dimensionless numbers and hypothesized that τ decreases in response to wind with an increase in h_{leaf} . Additionally, based on pixel-wise computation of τ from time series of thermal images, we provide spatial maps of leaf thermal responsiveness. Those spatial maps allow to separate areas in which thermal responsiveness is mainly driven by leaf heat capacity from areas

where thermal responsiveness is mainly driven by heat transfer processes. We suggest that the active thermography approach can be a powerful tool for modeling leaf heat transfer under well-defined environmental conditions in the laboratory.

MATERIAL AND METHODS

Theoretical Background and Model Description

According to the commonly used description of the steady state leaf energy balance model, all absorbed heat that originates from absorption of solar and thermal radiation (Φ_{in}), is dissipated by heat flux densities ($W\ m^{-2}$), which are namely the long-wave radiative heat flux density (LW), the convective heat flux density (H), and the evapotranspiration (λE) (Linacre, 1972; Jones, 1992; Monteith and Unsworth, 2008; Nobel, 2009) for abbreviations see **Table 1**.

TABLE 1 | Abbreviation list.

Abbreviation	Description	Unit
$C\ A^{-1}leaf$	leaf heat capacity per unit area	$J\ m^{-2}\ K^{-1}$
c_p	specific heat capacity at constant pressure	$J\ kg^{-1}\ K^{-1}$
d	characteristic leaf geometry	m
g_H	boundary layer conductance to convective heat	$m\ s^{-1}$
g_{LW}	boundary layer conductance to radiative heat	$m\ s^{-1}$
Gr	Grashof number	
g_s	stomatal conductance	$m\ s^{-1}$ or $mol\ m^{-2}\ s^{-1}$
H	convective heat flux density	$W\ m^{-2}$
h_H	convective heat transfer coefficient	$W\ m^{-2}\ K^{-1}$
h_{leaf}	leaf heat transfer coefficient	$W\ m^{-2}\ K^{-1}$
h_{LW}	long-wave radiative heat transfer coefficient	$W\ m^{-2}\ K^{-1}$
$h_{\lambda E}$	evapotranspirative heat transfer coefficient	$W\ m^{-2}\ K^{-1}$
k	thermal conductivity of air	$W\ m^{-1}\ K^{-1}$
LW	long-wave radiative heat flux density	$W\ m^{-2}$
LWC	leaf succulence/leaf water content per unit area	$mg\ cm^{-2}$
Nu	Nusselt number	
r_{aW}	boundary layer resistance to water vapor	$s\ m^{-1}$
Re	Reynolds number	
r_H	boundary layer resistance to convective heat	$s\ m^{-1}$
r_s	stomatal resistance	$s\ m^{-1}$ or $m^2\ s\ mol^{-1}$
r_W	leaf resistance to water vapor ($r_{aW} + r_s$)	$s\ m^{-1}$
s	slope relating saturation vapor pressure to temperature	$Pa\ K^{-1}$
$t_{0.5}$	time which is required to reach 50% of the initial value in an exponential decay	s
T_a	ambient air temperature	$^{\circ}C$ or K
T_L	steady state leaf temperature	$^{\circ}C$ or K
T'_L	non-steady state leaf temperature	$^{\circ}C$ or K
$T_L - T_a$	difference leaf temperature to ambient air temperature	$^{\circ}C$ or K
$u_{0.5}$	wind speed at which τ has decreased to 50% of its initial value	$m\ s^{-1}$
γ	psychrometer constant	$Pa\ K^{-1}$
ϵ	emissivity	
λE	evapotranspiration	$W\ m^{-2}$
ρ_a	density of air at constant pressure	$kg\ m^{-3}$
τ	time constant	s
Φ_{in}	incoming radiation (solar radiation + thermal radiation)	$W\ m^{-2}$

$$0 = \Phi_{in} - LW - H - \lambda E \quad (1)$$

If a leaf is not in an equilibrium, for instance following a short heat pulse, T_L transiently changes and afterwards approaches its former steady state value. To what extent a leaf heats up and how rapidly the heat is released, depends to a large extent on the leaf heat capacity per unit area ($C\ A^{-1}_{leaf}$), which is the energy required per leaf area to heat up the leaf by one degree. The difference between the leaf energy balance in equilibrium and in non-equilibrium is described as follows [for a derivation of the non-equilibrium leaf energy balance, see for example Appendix 9 in Jones (1992)]:

$$\frac{C}{A_{leaf}} \frac{\Delta T_L}{\Delta t} = \rho_a c_p (T'_L - T_L) \left[g_{LW} + g_H + \left(\frac{s}{\gamma r_W} \right) \right] \quad (2)$$

With $\Delta T_L \Delta t^{-1}$ as the T_L change over time, ρ_a is the density of air, c_p the specific heat capacity of air, T'_L the leaf temperature in non-steady state, T_L the steady-state temperature, g_{LW} the conductance to long-wave radiative heat, g_H the conductance to convective heat, s the slope relating saturation vapor pressure to air temperature (Penman, 1948), γ the psychrometer constant in $Pa\ K^{-1}$ which changes with temperature (e.g., Table A.3 in Monteith and Unsworth, 2008), and r_W the resistance to water vapor, which is the sum of the boundary layer resistance to water vapor (r_{aW}) and stomatal resistance (r_s). The formulation for r_{aW} accounts for amphistomatous leaves, such as barley and bean. For hypostomatous species this formulation differs slightly (see, e.g., Jones, 1992).

Because, the product of ρ_a , c_p and g is known as the heat transfer coefficient (h), Eq. 2 can be written as:

$$\frac{C}{A_{leaf}} \frac{\Delta T_L}{\Delta t} = (T'_L - T_L) h_{leaf} \quad (3)$$

Where h_{leaf} is the total leaf heat transfer coefficient and is the sum of the heat transfer coefficient for long-wave radiative heat (h_{LW}), the heat transfer coefficient for convective heat (h_H), and the heat transfer coefficient for evapotranspirative heat ($h_{\lambda E}$).

$$h_{leaf} = h_{LW} + h_H + h_{\lambda E} \quad (4a)$$

$$h_{LW} = \rho_a c_p g_{LW} \quad (4b)$$

$$h_H = \rho_a c_p g_H \quad (4c)$$

$$h_{\lambda E} = \rho_a c_p \left(\frac{s}{\gamma r_W} \right) \quad (4d)$$

The g_{LW} is given by $4\epsilon\sigma T_a^3 \rho_a^{-1} c_p^{-1}$ with ϵ being the emissivity and σ the Stefan-Boltzmann constant. As stated above, r_W is the sum of r_{aW} and r_s , where r_{aW} is assumed to be approximately r_H , the resistance to convective heat, as the reciprocal of g_H (Monteith and Unsworth, 2008).

To solve Eq. 3, a first-order differential equation is used, which is in a form of Newtons law of cooling (e.g., see Chapter 15 and Eq. 15.10 in Monteith and Unsworth, 2008):

$$dT_L(t) = \frac{1}{\frac{C}{A_{leaf}} h_{leaf}} (T'_L - T_L) \quad (5)$$

Note, that the application of Newton's law of cooling assumes constant ambient conditions.

In a further step, Eq. 5 can be solved with the following exponential function (**Figure S1**):

$$dT_L^* = T_L - (T_L - T_L')e^{-\frac{t}{\tau}} \quad (6)$$

Where T_L^* is any T_L during leaf cooling and τ the time constant, which is according Eq. 5 the product of leaf heat capacity per unit leaf area and the inverse of the leaf heat transfer coefficient:

$$\tau = \frac{C}{A_{\text{leaf}}} \frac{1}{h_{\text{leaf}}} = \frac{C}{A_{\text{leaf}}} \frac{1}{\rho_a c_p \left(g_{LW} + g_H + \left(\frac{s}{\gamma r_w} \right) \right)} \quad (7)$$

Using dimensionless numbers, g_H can be calculated with the following equation (e.g., Dixon and Grace, 1983; Bailey and Meneses, 1993):

$$g_H = \frac{Nu \, k}{\rho_a c_p d} \quad (6)$$

Nu is the Nusselt number, k the thermal conductivity of air, and d refers to the mean leaf diameter in m.

Nu depends on the prevailing conditions, in particular whether free or forced convection is dominant. Under free convection, where no wind occurs and heat transfer is mainly due to heat upwelling from the leaf surface, Nu depends on a further dimensionless number, the Grashof number (Gr):

$$Nu = aGr^b \quad (7)$$

The numerical constants a and b describe the geometry of a leaf (Schuepp, 1993).

Under conditions where wind occurs, a further dimensionless number has to be considered, the Reynold's number (Re), which describes forced convection. At low wind conditions, mixed convection is most likely (Schuepp, 1993) and Nu has to be calculated with Gr and Re .

$$Nu = a(Gr + 1.4Re^2)^b \quad (8)$$

We determined a and b experimentally for both barley and bean leaves with respect to the prevailing wind conditions. Detailed descriptions of the model we used and the respective values are given in Supplementary material (Eq. S1 to Eq. S8, **Table S1**, and **Figures S1** to **S4**).

Plant Material

All plants were grown in the greenhouse facilities at the IBG-2, Forschungszentrum Jülich in spring 2015. We grew spring barley (*Hordeum vulgare* var. Victoriana) and common bean (*Phaseolus vulgaris* var. Shiny). Barley plants were germinated in 12 x 12 x 15 cm pots and bean plants in 15 x 15 x 18 cm pots. Pots were filled with a potting substrate, enriched with 1 g L⁻¹ NPK fertilizer and with 2 g L⁻¹ of a long-time acting fertilizer (Einheitserde Typ ED73). Plants were placed on moist, water-retentive cloths.

Plants grew in a day-night cycle of 16 h day and 8 h night with air humidity around 55% ± 13%. Mean T_a was 20.8°C ± 2.6°C, and the highest measured T_a during this period was 30.7°C,

whereas the minimum T_a was 16.8°C. On sunny days, light intensities in the greenhouse at midday reached on maximum about 1,300 μmol m⁻²s⁻¹, while the minimum illumination intensity was about 85 μmol m⁻²s⁻¹, including artificial light, in the late afternoon. For the measurements, bean plants were about 2 weeks old and barley plants were about 6 weeks old.

For all measurements, plants were moved from the greenhouse into the laboratory, where they were dark-adapted over night for about 14 h prior to the measurements.

Thermal Imaging

For all measurements we used a VarioCAM ® hr head (InfraTec, Germany). This camera is equipped with a microbolometer focal plane array that captures and integrates thermal infrared radiation within the spectral range between 7.5 μm and 14 μm. The field of view (FOV) is 30° x 23°, with a geometric resolution of 640 by 480 pixels, a measuring accuracy of ± 1 K, with a thermal sensitivity <30 mK. Images were recorded with the IRBIS ® 3 software (InfraTec, Germany) that allows real-time tracking and correction of temperatures by setting parameters, such as ϵ and background radiation. Background radiation was measured by a sheet of crumpled aluminum foil and emissivity (ϵ) was set to 0.95 (Nobel, 2009). Even though the radiator beam was mainly targeting the leaf itself during the measurements, the background temperature showed slight variations in the range of 0.3–0.4 K, as measured by aluminum foil placed in the center of the beam. According to the Stefan-Boltzmann Law this corresponds to a variable incoming background energy of approx. 2 W m² and we assume that these small variations had only a minor effect on the time constant of leaf cooling. The IRBIS software allows the export of single pixel and integrated pixel area data, as well as pixel data as a text file.

To adapt T_a , measured by thermocouples (Type K, Newport Omega, Germany) to temperatures measured by thermography, we developed a correction routine for T_a . For this purpose, we built a “radiation-trap” which absorbs and emits all incoming heat, providing a black-body-like reference. We used a 2x2x15 cm box made of black cardboard, which was thermally insulated by means of a 0.5 cm thick Styrofoam layer to minimize temperature fluctuations. The inner side of the box was covered with aluminum foil, which was crumpled and painted with black emissivity paint ($\epsilon = 0.95$) (TETENAL Europe GmbH, Germany), so that the overall emissivity of the inner box was 1. The upper side of the box, always facing the camera, had a 1-cm-diameter hole. Within the box a thermocouple was placed, which is identically constructed to the one that measures T_a outside the box. To adapt the measured T_a to temperatures obtained by thermography, we calculated the difference between the thermocouple within the box and the temperature of the hole measured with the thermal camera. In preliminary experiments, we compared $T_L - T_a$ obtained by thermography with $T_L - T_a$ obtained by thermocouples and found that we could use the box as a correction factor for T_a .

To actively and transiently warm up leaves with short heat pulses, we used two commercial near-infrared (NIR) heating units (Heizmeister 1000 IP65, Infraloc, Germany) equipped

with a “long life light-tube (Helen Goldröhre”, Infrallogic, Germany), which emits radiation with a maximum power of $1,000 \text{ W m}^{-2}$ in the range between 750 and 2,000 nm. This spectral range is suitable for actively heating leaves because significant water absorption bands located at 1,450 and 1,950 nm within this spectral range (Gausman and Allen, 1973; Asner, 1998; Seelig et al., 2008; Seelig et al., 2009). The heating units were connected to unit (Eurolite ® ESX-4 DMX, Eurolite, Germany) that was software controlled (DMX-Configurator, DMX4ALL GmbH, Germany) allowing the setting of intensity, duration, and interval of the NIR heat pulses. We applied a 1.15 s heat pulse to the leaves at approximately half of the maximum power.

Experimental Set-Up

All instruments were mounted on a metal profile construction (Figures S6 and S7). The camera head was mounted at a height of 1 m at a 90° angle to the ground. The two NIR heaters were placed at the camera's height with a distance of about 30 cm to the camera and facing to the ground at an angle between 60° and 70° . Additionally, two white LED panels (SL 3500-W-G, Photon System Instruments, Czech Republic) were installed at the camera's height at an angle of about 45° to the ground. A small ventilator, capable to produce low wind-speeds between 0.2 and 1.6 m s^{-1} , was installed on a vertical metal bar, in order to produce laminar wind streams from the leaf tip to the leaf base. Wind-speeds were measured by a thermo-anemometer with hotwire (VT 110, KIMO Instruments, France). Leaves were fixed by lab-stand clamps horizontally to the ground, i.e. perpendicular to the camera. T_a was measured with a thermocouple that was attached to the lab-stand and protected against direct irradiation with aluminum foil. T_a and the temperature within the radiation trap were recorded every second with a data logger (HH506RA, Newport Omega, Germany).

Measurements

Leaf Water Content

For the measurements with barley, leaves of different developmental stages were randomly chosen. Measurements with bean, were performed with the primary leaf pair and plants were not older than 2 weeks, because leaves at different developmental stages showed different leaf shapes, which in theory impacts the leaf heat transfer. Intact leaves were dark-adapted and measured at three different wind-speeds, 0.0 m s^{-1} , 0.5 m s^{-1} , and 1.0 m s^{-1} . For each wind-speed step, 15 barley leaves and 8 bean leaves each of different plants were measured. After T_L cooling curves were measured and leaves had cooled to T_L prior to the heat pulse, stomatal conductance (g_s) was measured with a gas exchange device (Licor-6400, LICOR, Nebraska, USA). Leaves were harvested and analyzed for leaf area and fresh weight afterwards. For dry weight determination, harvested leaves were dried in an oven at 80°C for 48 h until a constant weight was reached. We calculated LWC, the absolute leaf water content per unit area, as the difference between fresh weight and dry weight per unit leaf area.

Wind Treatments

To induce changes in the boundary layer thickness and conductance, single leaves were exposed to increasing wind speeds, which were produced by the small ventilator integrated in the set-up. Reference measurements for gH modeling were performed with dark-adapted leaves at eight wind-speed steps, 0.0 , 0.2 , 0.4 , 0.6 , 0.8 , 1.0 , 1.2 , and 1.4 m s^{-1} on a separate set of plants. For the actual experiment, a second set of plants was used. Dark-adapted leaves were exposed to increasing wind at speeds of 0.0 , 0.4 , 0.8 , and 1.2 m s^{-1} and measured at each step. Afterwards, the leaves were light-adapted to a light intensity (photosynthetically active photon flux density) of about $1000 \mu\text{mol m}^{-2} \text{ s}^{-1}$, using white LED panels, until T_L and stomatal conductance (g_s) reached steady state values. Usually this was the case after 30 to 40 min after light exposure. Leaves were measured again at the four wind speeds indicated above. Again, g_s was measured using the Licor-6400 after each leaf cooling curve. Typically, g_s stabilized and reached stable values within 1 min.

Data Processing and Analyses

τ -Measurements

We evaluated measured cooling curves using two procedures. In a first procedure, we used the mean T_L values, which were obtained by defining the whole leaf as an area of interest (IRBIS® 3 software, InfraTec, Germany. The software automatically integrates all temperature pixels and provides the mean T_L value. Measured cooling curves were then fitted with Eq. 6 to obtain τ from the fitting (Origin 8.5, OriginLab, USA).

In a second procedure, we mapped τ spatially by calculating τ for each single pixel in the image. For this purpose, we developed an automated analysis routine for the MATLAB Environment. A typical data set consists of n images, containing a data matrix with the temperatures T^t at the measured time t . The fitting function $T_L(t)_{ij} = T_L(t_\infty)_{ij} - dT_{L_{ij}} e^{-(t/\tau)}$ was computed for each pixel at the position ij and is in the form of Eq. 3. The optimization of the curve fitting was done by minimizing the sum of squared residuals using the downhill simplex approach (Nelder and Mead, 1965).

A graphical user interface (GUI) supports the processing of a single image series, or of several image series. The required input data are an Excel-file containing the time points of each recorded image and the corresponding images as a text file (ASCII), which were exported from the IRBIS® 3 software before. For the data import of several image series, a list (Excel-file) referring to the respective file path is required. Images and time data, which are located in the respective file-path, are then automatically loaded. Additionally, the GUI provides functions for postprocessing of τ -matrices. Minimum- and maximum-thresholds for τ -values can be used. These were generally set between 0 s and 250 s. Additionally, the r -value of the exponential regression can be used as a further filter-parameter, which we have set to $r = 0.9487$, which corresponds to a r^2 -value of 0.9. Resulting filtered as well as non-filtered τ -matrices are provided as Excel-tables for further manual post-processing. The thresholds used in the postprocessing procedure may result in empty pixels on the

imaged leaf, which were filled by using the median-value of the surrounding pixels.

Statistical Analyses

Statistical analyses were performed using SigmaPlot (Systat Software, Inc., USA) and included analyses of variance (ANOVA), Pearson correlation analyses for linear relationships, and Spearman correlation analyses for nonlinear relationships. Before each ANOVA, the data were tested for normal-distribution. In cases where no normal-distribution was present, an ANOVA on the ranks was performed. In each case, the Tukey post-hoc test was used for pairwise comparisons.

RESULTS

Relationship Between LWC and τ of Dark-Adapted Leaves

We tested the relationship between τ , obtained by active thermography, and LWC by measuring dark-adapted leaves of both, spring barley and common bean. These relationships were assessed at three different wind-speeds, 0.0 m s^{-1} , 0.5 m s^{-1} , and 1.0 m s^{-1} (Figure 1). In all cases, correlations between τ and LWC were significant for both barley and bean, at all wind speed steps ($p < 0.05$). Generally, the linear regression to quantify the relationships revealed slopes that increased with increasing wind speed.

Wind- and Light-Induced Changes in Leaf Heat Transfer Parameters

To quantify the effect of a changing boundary layer on τ , we compared wind curves of dark-adapted leaves to wind-curves of light-adapted leaves (Figure 2). For both barley and bean we observed significant changes ($p < 0.05$) in τ for dark- and light-adapted leaves in response to increasing wind speed (Figures 2A, B). We characterized the τ -response with an exponential regression and the derived wind speed at which τ has decreased to 50% of its initial value ($u_{0.5}$). For dark-adapted leaves we obtained $u_{0.5}$ -values of 0.26 m s^{-1} and 0.23 m s^{-1} for barley and bean, respectively. For light-adapted leaves, the decrease was characterized by $u_{0.5}$ -values of 0.52 m s^{-1} for barley and 0.33 m s^{-1} for bean. Additionally, τ -values of light-adapted leaves were significantly lower compared to dark-adapted leaves ($p < 0.05$ for barley and $p < 0.001$ for bean). Significant differences in τ between barley and bean were found at wind speeds of 0.0 m s^{-1} and 0.8 m s^{-1} ($p < 0.05$). Absolute τ -values were always higher for bean leaves compared to barley leaves irrespectively of the wind speed and adaptation state. At zero wind, the mean τ was more than 10 s higher for bean compared to barley.

We could not observe a comparable pattern in the response of $T_L - T_a$ to both wind and light (Figures 2C, D). For barley leaves, $T_L - T_a$ seemed to remain relatively stable throughout the measurements ($p > 0.05$), while light-adapted leaves were generally cooler compared to dark-adapted leaves. Significant differences between dark- and light-adapted leaves, however, were only found at wind speeds of 0.0 m s^{-1} and 0.4 m s^{-1} ($p <$

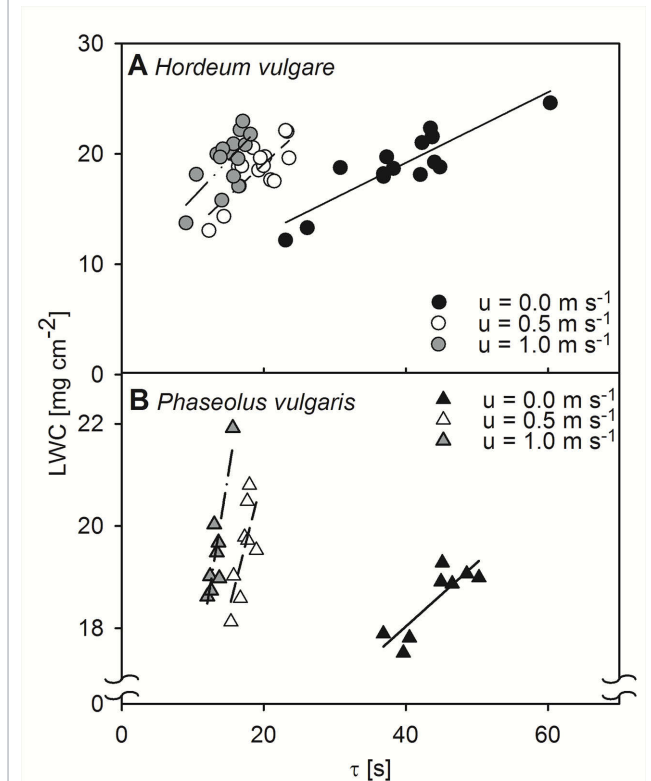


FIGURE 1 | Relationship between leaf water content per unit area (LWC) and time constant (τ) of dark-adapted leaves. Relationships were assessed at different wind speeds of 0.0 m s^{-1} (closed symbols), 0.5 m s^{-1} (open symbols), and 1.0 m s^{-1} (grey symbols). **(A)** Measurement of single leaves of spring barley (*Hordeum vulgare*). Linear regression for measurements at a wind speed of 0.0 m s^{-1} (solid line): $y = 0.32x + 6.40$, $r^2 = 0.79$, $p < 0.001$, for measurements at wind speeds of 0.5 m s^{-1} (dashed line): $y = 0.59x + 7.29$, $r^2 = 0.63$, $p < 0.001$, and for measurements at wind speeds of 1.0 m s^{-1} (dash-dot-dotted line): $y = 0.68x + 9.18$, $r^2 = 0.48$, $p < 0.05$. **(B)** Measurements of single leaves of common bean (*Phaseolus vulgaris*). Linear regression for measurements at a wind speed of 0.0 m s^{-1} (solid line): $y = 0.12x + 13.04$, $r^2 = 0.72$, $p < 0.01$, for measurements at wind speeds of 0.5 m s^{-1} (dashed line): $y = 0.54x + 10.17$, $r^2 = 0.51$, $p < 0.05$, and for measurements at wind speeds of 1.0 m s^{-1} (dash-dot-dotted line): $y = 0.86x + 8.14$, $r^2 = 0.79$, $p < 0.01$. Each point represents an individual leaf that was measured by the active thermography approach and afterwards destructively analyzed for LWC.

0.05). For dark-adapted bean leaves we observed an exponential decrease in $T_L - T_a$ in response to increasing wind speed, and an exponential increase in response to increasing wind speed for light-adapted leaves (Figure 2D). At zero wind, light-adapted leaves were cooler compared to dark-adapted leaves. At a wind speed of 0.4 m s^{-1} , $T_L - T_a$ values were similar and when wind speeds exceeded 0.4 m s^{-1} light-adapted leaves were warmer compared to dark-adapted leaves. Except at wind speeds of 0.4 m s^{-1} the differences between dark- and light-adapted leaves were significant ($p < 0.05$).

For both barley and bean, g_s increased significantly ($p < 0.05$) in response to light (Figures 2E, F). Although g_s slightly decreased on average in response to increasing wind speed, we

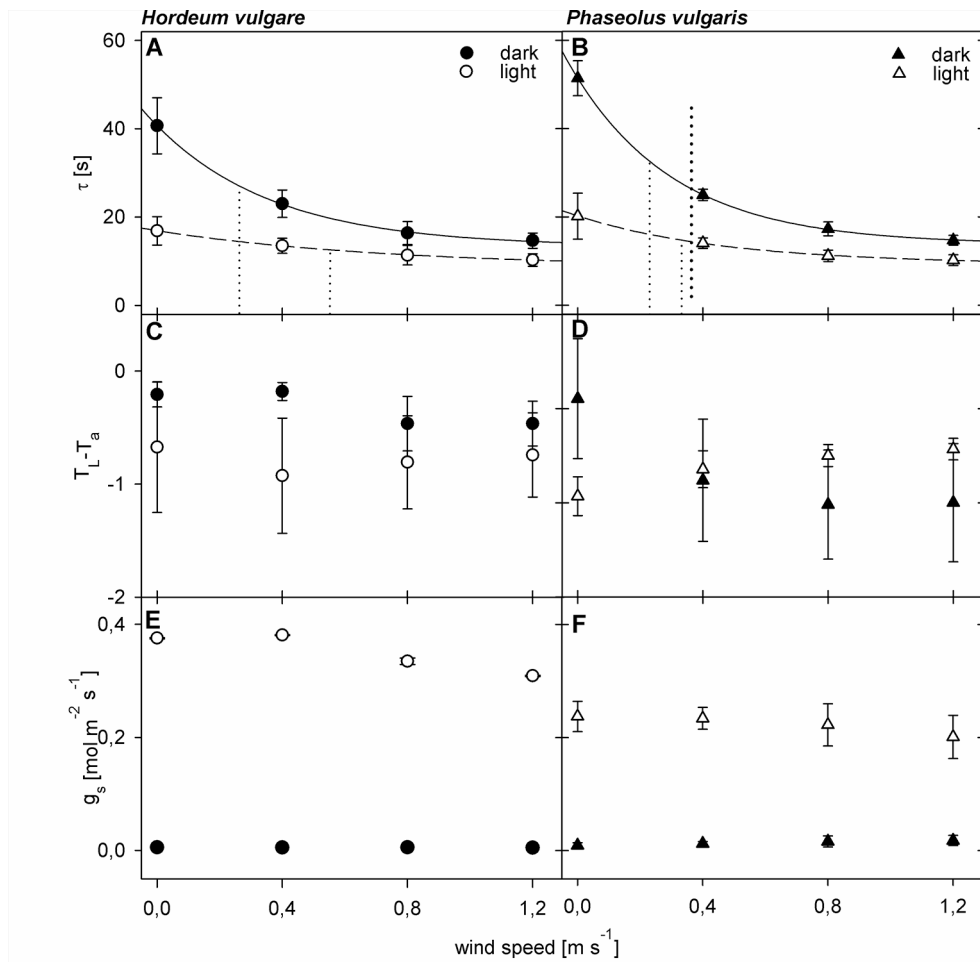


FIGURE 2 | Wind- and light-induced changes in leaf heat transfer parameters of spring barley (*Hordeum vulgare*) and common bean (*Phaseolus vulgaris*). Measurements of dark-adapted leaves are indicated by closed symbols, and measurements of light-adapted leaves are indicated by open symbols. **(A, B)** Wind- and light induced changes in time constant (τ). Dotted lines represent the wind speed at which τ has decreased to 50% of its initial value ($u_{0.5}$). **(C, D)** Wind- and light-induced changes in difference between leaf temperature and ambient air temperature ($T_L - T_a$). **(E, F)** Wind- and light-induced changes in stomatal conductance (g_s). Error bars indicate standard deviation. For barley plants $n = 9$ individual leaves, for bean $n = 10$ individual leaves.

did not find any significant changes, neither for barley, nor for bean.

Wind- and Light Induced Spatial Variations in $T_L - T_a$

To evaluate changes in leaf heat transfer in response to wind and light illumination we mapped $T_L - T_a$ spatially for representative leaves of bean and barley, respectively (**Figure 3**). Generally, dark-adapted leaves had a more homogeneous distribution of $T_L - T_a$ over the leaf surface compared to light-adapted leaves (**Figure S8**). We observed that $T_L - T_a$ for dark-adapted leaves appeared to become more homogeneous with increasing wind speed from 0.0 to 1.2 m s⁻¹. For light-adapted bean leaves, areas in between major veins generally appeared cooler compared to areas with a comparably higher density of major veins. Particularly, at zero wind, leaf areas where lower order veins

are located and areas which are nearer to the leaf edges were cooler than the center of the leaf. With increasing wind speed, $T_L - T_a$ increased and areas that appeared cooler earlier got warmer, particularly at the leaf tip, which was in the direction of the wind-leading edge. For barley leaves, the observed patterns were not as clear as those observed for bean leaves (**Figure 3B**). For example, when leaves were light-adapted at wind speeds of 0.0 m s⁻¹ and 0.8 m s⁻¹, the mid-vein was only partly visible because it was warmer compared to the leaf lamina.

Wind- and Light Induced Spatial Variations in τ

Spatial maps of τ provide information on the thermal responsiveness of a leaf, because this quantity is related to both, water distribution in the leaves and h_{leaf} . As we observed for the mean values, the images illustrate that with increasing

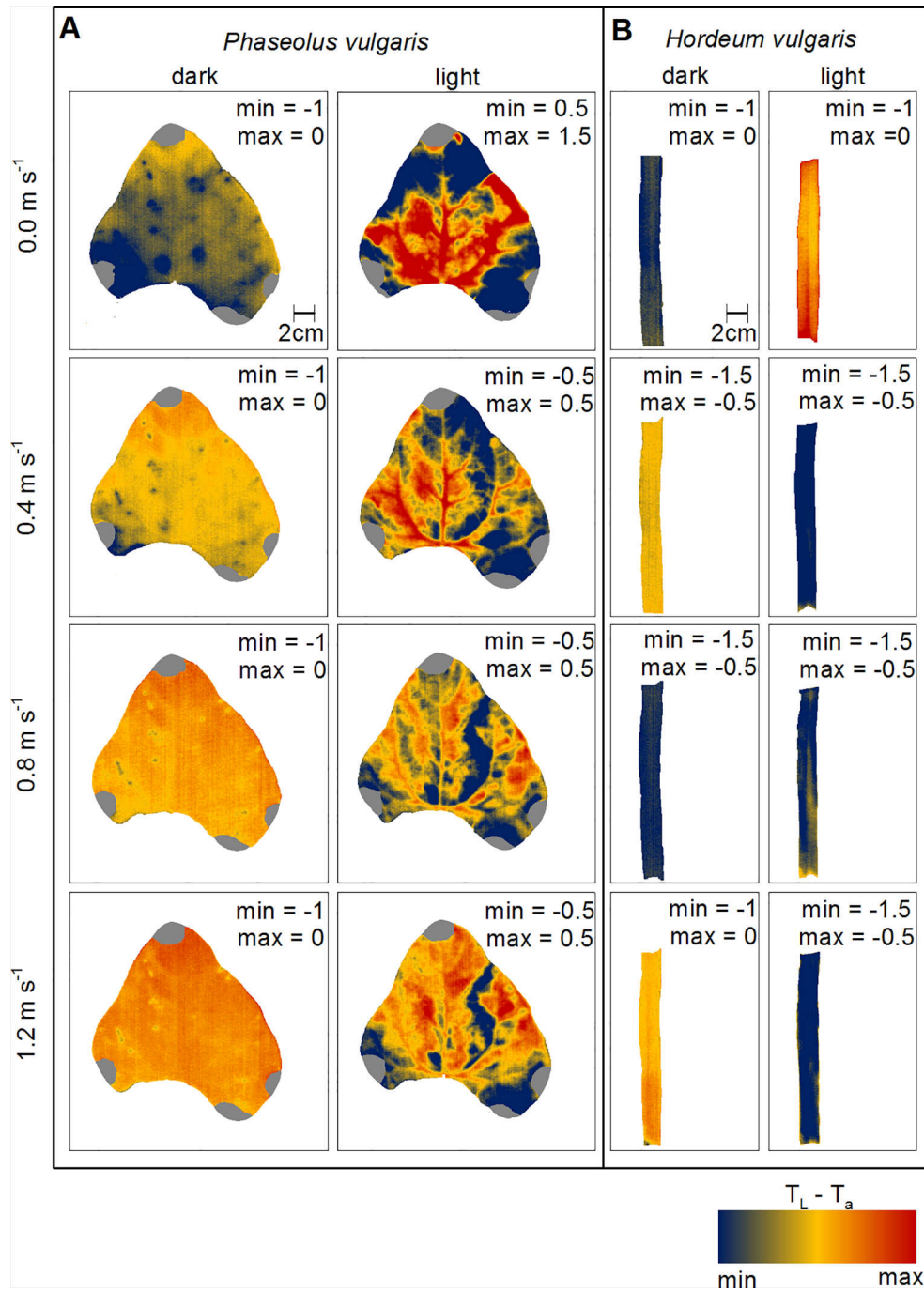


FIGURE 3 | Spatial mapping of wind- and light-induced changes in leaf temperature to ambient air temperature difference ($T_L - T_a$). **(A)** Representative leaf of common bean (*Phaseolus vulgaris*) and **(B)** representative leaf of spring barley (*Hordeum vulgare*). Dark-adapted leaves are presented in the first column from the left and light-adapted leaves in the second column from the left, respectively. Each row represents measurements at different wind speeds of 0.0 m s⁻¹, 0.4 m s⁻¹, 0.8 m s⁻¹, and 1.2 m s⁻¹. $T_L - T_a$ are color-coded as indicated by the false-colorscale at the bottom. Minimum (min) and maximum (max) values of $T_L - T_a$ are given in each panel.

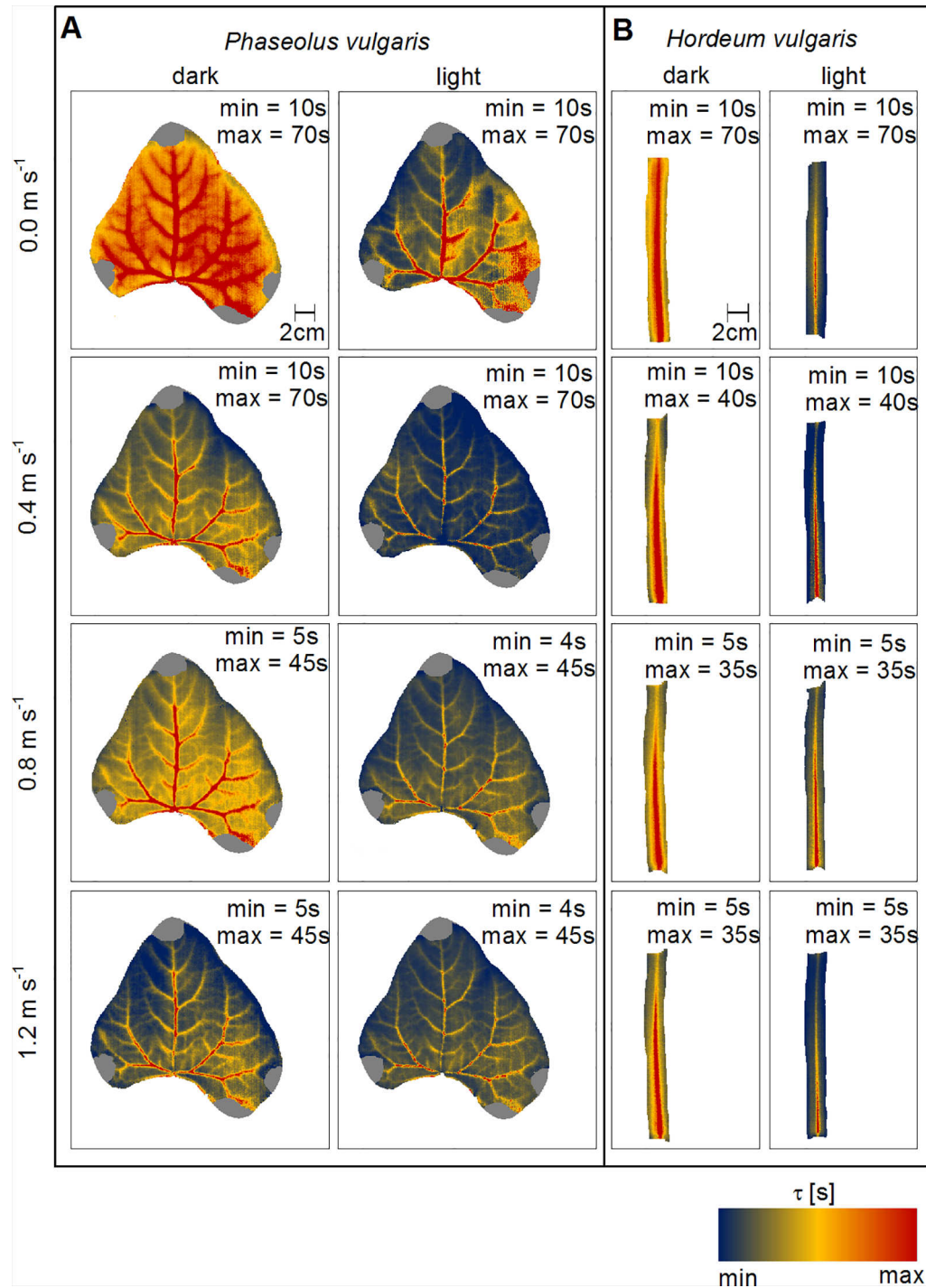


FIGURE 4 | Spatial mapping of wind- and light-induced changes in time constant (τ). **(A)** Representative leaf of common bean (*Phaseolus vulgaris*) and **(B)** representative leaf of spring barley (*Hordeum vulgare*). Dark-adapted leaves are presented in the first column from the left and light-adapted leaves in the second column from the left, respectively. Each row represents measurements at different wind speeds of 0.0 m s⁻¹, 0.4 m s⁻¹, 0.8 m s⁻¹, and 1.2 m s⁻¹. $T_L - T_a$ are color-coded as indicated by the false-color scale at the bottom. Minimum (min) and maximum (max) values of τ are given in each panel.

wind speed and with light-illumination τ decreased (Figure 4). The most prominent structures were the leaf vascular tissues, which was reflected by comparably higher τ -values irrespective of wind speed and illumination state. While we were able to detect second and third order veins on the bean leaf (Figure 4A), we could only map the main-vein on the barley leaf (Figure 4B). In dark-adapted leaves, the highest τ -values were associated with larger order veins near the leaf base. Additionally, a gradient from high to low τ -values could be observed from the leaf base to the leaf tip and towards the leaf edges (Figure S9). Generally, in response to light τ decreased. However, the most prominent leaf structures were the veins, as indicated by higher τ -values compared to the leaf blades, where smaller order veins are located. Still, major, second and third order veins were visible in bean leaves.

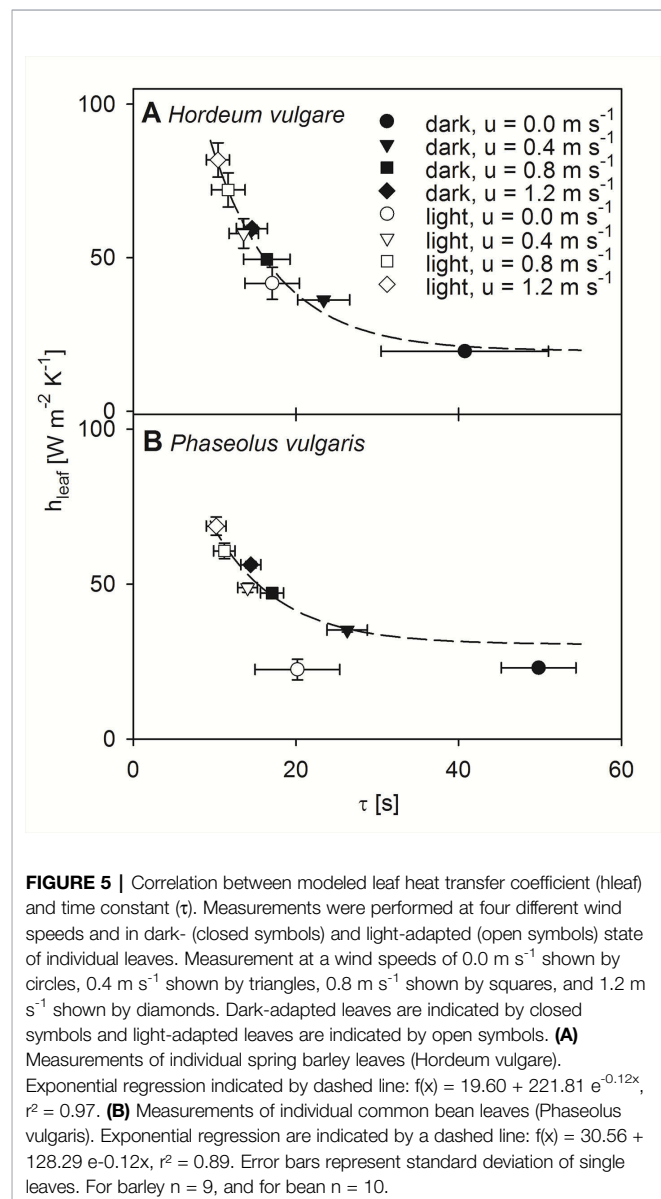
We analyzed the image pixels for nonlinear correlations using the Spearman correlation analysis and found a significant correlation between $T_L - T_a$ and τ for barley and bean at all experimental levels ($p < 0.05$).

Correlation Between Modeled h_{leaf} and τ

Using the dimensionless numbers approach we calculated h_{leaf} for each experimental level (Figure 5). Note here that, because the modeled data for bean leaves at free convection revealed some weaknesses most likely due to a weak linear relationship between τ and LWC (Figure 1B), these data were consequently excluded from further statistical analyses. For both barley and bean, we found a highly significant correlation between the modeled h_{leaf} and the measured τ ($p < 0.001$). Generally, h_{leaf} of light-adapted leaves were higher compared to h_{leaf} of dark-adapted leaves. Relationships between h_{leaf} and τ were characterized by exponential regressions, which revealed the time that is required to reach 50% of the initial value ($t_{0.5}$). For both plant species $t_{0.5}$ -values of 5.6 s were obtained, indicating a similar response of τ to h_{leaf} . However, absolute h_{leaf} -values were higher for barley leaves, compared to bean leaves. While barley leaves reached on average maximum values of $81.8 (\pm 5.6) \text{ W m}^{-2} \text{ K}^{-1}$ (Figure 5A), bean leaves reached on average maximum values of $68.7 (\pm 2.9) \text{ W m}^{-2} \text{ K}^{-1}$ (Figure 5B).

Contribution of h_{LW} , h_{H} , and $h_{\lambda\text{E}}$ to the Overall h_{leaf}

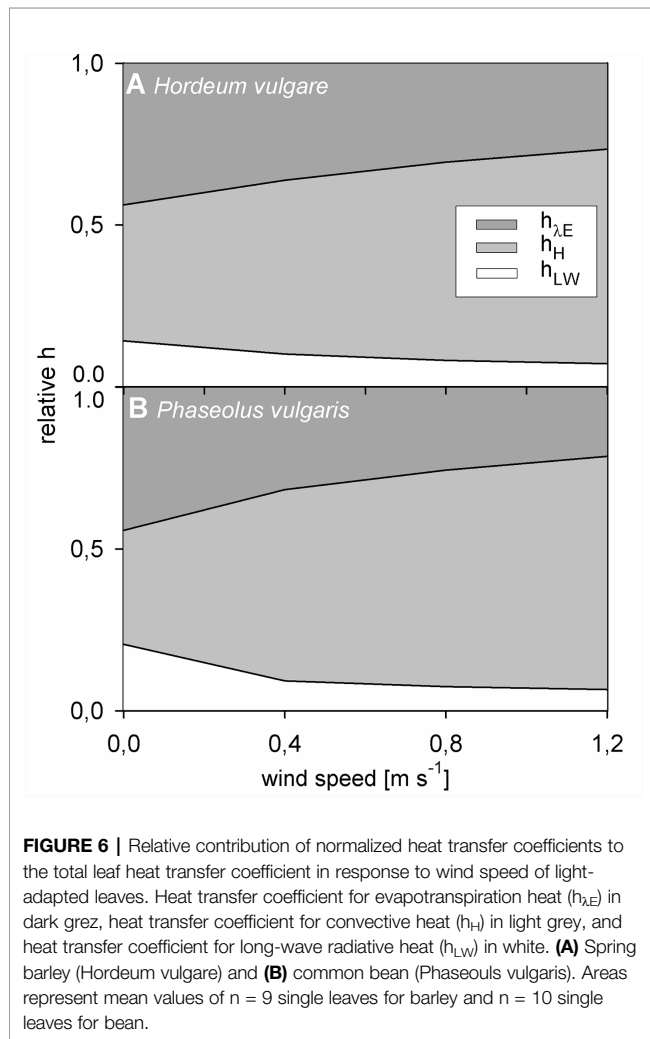
Finally, we evaluated the impact of each heat transfer coefficient on the overall h_{leaf} on light-adapted leaves. For this evaluation, the portion of the respective heat transfer coefficient in the entire h_{leaf} was calculated (e.g., $h_{\text{H}} h_{\text{leaf}}^{-1}$). While h_{LW} did not correlate with h_{leaf} , for both barley and bean, h_{H} and $h_{\lambda\text{E}}$ showed a strong and significant correlation with h_{leaf} ($p < 0.001$). The relative contribution of each heat transfer coefficient to the overall h_{leaf} significantly changed with wind speed ($p < 0.001$) (Figure 6). While the relative contribution of h_{H} increased with increasing wind speed, the relative contribution of h_{LW} and $h_{\lambda\text{E}}$ decreased with increasing wind speed. At zero wind $h_{\lambda\text{E}}$ had the highest impact on h_{leaf} accounting for about 45% of the overall h_{leaf} . However, albeit lower, at 1.2 m s^{-1} the relative contribution of $h_{\lambda\text{E}}$ was still significant with a contribution of 26% and 21% for barley and bean, respectively.



DISCUSSION

In this paper, we introduce the active thermography approach as a powerful method to evaluate leaf heat transfer processes of intact leaves. We thoroughly tested the active thermography experimental protocol in the laboratory on spring barley and common bean leaves, by comparing measured τ to modeled h_{leaf} . We found strong relationships between τ and h_{leaf} , which were valid for dark- and light-adapted leaves at varied wind speeds. Additionally, spatial $T_L - T_a$ - and τ -maps revealed the impact of local differences in thermal responsiveness, related to $C A^{-1}_{\text{leaf}}$ and h_{leaf} differences, on T_L .

To evaluate the measured responses of τ with respect to changes in h_{leaf} , we modeled h_{leaf} using dimensionless numbers. We evaluated the modeled data by comparing $C A^{-1}_{\text{leaf}}$ derived from dimensionless numbers and τ introducing $C A^{-1}_{\text{leaf}}$



derived from the linear relationships found between τ and LWC (Figures 1, S2, and S5).

These models yielded comparable values with some differences at zero wind, particularly for bean. If no wind is present, free convection dominates, which mainly depends on leaf area and surface structure (compare Eq. S3). Because the variation in leaf area was much higher for bean leaves ($\pm 46 \text{ cm}^2$) than for barley leaves ($\pm 5.5 \text{ cm}^2$), we assume a high variability of hleaf leading to errors in the linear relationship between τ and LWC. Both measurements and the modeling of heat transfer in wind-free conditions assuming free convection are difficult, because leaf area and rough surface elements affect the heat transfer more strongly than during forced convection (Kumar and Barthakur, 1971). Particularly for bean leaves, we could observe great spatial heterogeneity of $T_L - T_a$ over single leaves similar to prior simulations (Roth-Nebelsick, 2001) and that may result in thermal instabilities of the boundary layer (Defraeye et al., 2013). In addition to leaf area, surface roughness, caused by trichomes and vascular tissue, affects hleaf (Parkhurst, 1976; Schreuder et al., 2001). In contrast to bean, barley has a relatively

flat surface with a regularly arranged and parallel vein system (Dannenhoffer et al., 1990; Ueno et al., 2006). Bean has an uneven surface with plenty of thick veins that are dichotomously branched, which is likely to disturb the air movements across the leaf surface and influence heat transfer. Furthermore, heterogeneous $T_L - T_a$ may be a result of heterogeneous vein density and distribution of stomata over the leaf surface, which both affect hleaf. It was observed that the leaf conductance to water vapor was up to 18% higher at the leaf tips compared to the leaf base, which was explained by a higher vein density in this region (Nardini et al., 2008). Particularly in free convection conditions and at low wind speeds, leaf tips appeared to be cooler compared to the leaf base (Figure 3), which might be attributable to comparable higher leaf conductance to water vapor. Heterogeneity in the density and distribution of stomata over the leaf surface (Fanourakis et al., 2015), may affect $T_L - T_a$ in a similar way.

Our observation that measured τ and modeled hleaf decrease in response to increased wind-speed, which affects the boundary layer thickness, is in agreement with previous findings (Raschke, 1960; Vogel, 2009). Generally, barley leaves showed higher hleaf and lower τ -values compared to bean leaves. Smaller and narrower leaves have a thinner and more homogeneous boundary layer (Gates, 1965; Sinclair, 1970; Roth-Nebelsick, 2001). However, the response to wind was quite similar for barley and bean leaves, as indicated by $u_{0.5}$ -values (Figure 2). If wind is present, the leaf boundary layer thickness will be reduced and heat will be increasingly transported away from the leaf surface with the air movement, which increases hleaf and thus decreases τ (Schuepp, 1993; Vogel, 2009; Defraeye et al., 2013). The assumption that the boundary layer thickness is progressively reduced (Kitano and Eguchi, 1990) is supported by spatial $T_L - T_a$ and τ -maps of bean leaves. In our experiments the wind-leading edge (leaf tip) in response to wind got warmer and τ decreased compared to the wind-averted leaf edge. For barley this effect was not visible, which might be related to generally more homogeneous boundary layer and smoother leaf surface that offers less resistance to the air stream (Gates, 1965; Sinclair, 1970; Dannenhoffer et al., 1990).

We have observed a decrease in τ of about 24 s and 34 s upon illumination for barley and bean, respectively (Figures 2A, B). Based on the established linear relationships (Figure 1), these changes would correspond to a water loss of 20% to 30%, so that the decrease in τ cannot be explained by a decrease in LWC. However, light induces stomatal opening, which increases hleaf and particularly $h_{\lambda E}$ (Mott et al., 1997; Roelfsema and Hedrich, 2005; Shimazaki et al., 2007; Pieruschka et al., 2010). In our experiments, we found that $h_{\lambda E}$ contributes about 45% to the overall hleaf for light-adapted leaves under wind free conditions. Thus, τ is strongly affected by leaf conductance to water vapor, although the relative contribution of $h_{\lambda E}$ to the overall hleaf decreased with increasing wind speed. At higher wind speeds, stomatal resistance has larger effect on water vapor fluxes than the convective resistance (Cannon et al., 1979; Defraeye et al., 2013). Consequently, $h_{\lambda E}$, which depends on both convective and stomatal resistance, increases less strongly

in response to increased wind compared with h_H , which depends on convective resistance alone. Thus, under these conditions the relative contribution of $h_{\lambda E}$ to the overall hleaf decreases whereas the relative contribution of h_H increases. Nevertheless, because both barley and bean leaves were transpiring at high rates (barley: $0.35 (\pm 0.03) \text{ mol m}^{-2} \text{ s}^{-1}$, bean: $0.22 (\pm 0.02) \text{ mol m}^{-2} \text{ s}^{-1}$), some water loss is also very likely, which results in an even stronger decrease of τ .

To evaluate the impact of τ on T_L , and thus the thermal responsiveness on T_L , we mapped both, τ and $T_L - T_a$ spatially (Figures 3 and 4). Correlations between $T_L - T_a$ and τ in our experiments indicate a strong relationship between T_L and thermal responsiveness. The highest τ -values were associated with leaf vascular tissues, because leaf veins have a higher $C A^{-1}_{\text{leaf}}$ compared to the leaf lamina (McKown et al., 2010; Sack and Scoffoni, 2013). Particularly visible at bean leaves, those areas, which were associated with higher τ -values, appeared also warmer compared to the rest of the leaf. Near the veins the density of stomata is low and with it the leaf conductance to water vapor and thus the overall leaf heat transfer. In contrast, areas in between the larger order veins, where the stomata density is higher and with it the leaf conductance to water vapor, showed lower τ -values and appeared cooler. Additionally, in these regions leaf thickness is lower and less water is present. We conclude that local variations in the leaf heat transfer and water within the leaves ($\approx C A^{-1}_{\text{leaf}}$) will result in local differences of thermal responsiveness of the leaf and thus T_L . Using the computed spatial maps of τ , it is possible to separate areas in which thermal responsiveness mainly depends on $C A^{-1}_{\text{leaf}}$ from those in which thermal responsiveness mainly depends on hleaf. Finally, those τ -maps may contribute to studies aiming at detect heterogenous leaf conductance to water vapor (e.g., Nardini et al., 2008) or at detection of stomata heterogeneity (e.g., Fanourakis et al., 2015).

Our results indicate that h_H is essential to assess the overall h_{leaf} . At all experimental conditions h_H strongly contributed to h_{leaf} (Figure 6). Only at zero wind speed hleaf was dominated by $h_{\lambda E}$. At all remaining conditions, h_H was the dominant heat transfer coefficient. Convective heat and transpiration are inevitably coupled because heat associated with water vapor must penetrate the boundary layer depending on the prevailing convective conditions. It follows that modeling of transpiration requires consideration of convective processes. Because τ is a good measurement of h_{leaf} as demonstrated here and in agreement with previous reports (Pearman, 1972; Parlange and Waggoner, 1971; Saldin and Barthakur, 1971; Kumar and Barthakur, 1971) and we could also map τ spatially using active thermography, this approach should contribute to more precise modeling of leaf heat transfer processes in the future.

Plant water relations and h_{leaf} , both related to τ , are of importance for modeling plant-environment interactions (Foley et al., 2003), as well as for plant phenotyping, especially considering the necessary gain in plant productivity and water-use-efficiency under water-limited agriculture (Passioura and Angus, 2010; Sampoux et al., 2011; Farrar et al., 2011). We provide experimental evidence that τ measured with the active

thermography approach is an important parameter describing leaf heat transfer processes.

However, we also want to point out the novelty of our approach, as active thermography is not well established in plant sciences yet. Unidentified sources of error may still persist, as for instance uncertainties in model parameterization. While the values for the numeric constant b for free and mixed convection (cf. Eqs. 7 and 8, Table S1) were in the range of values presented by Dixon and Grace (1983), values for the constant a were considerably higher than reported previously for artificial leaves in a laminar air flow (Bailey & Meneses, 1993). Additionally, we found different values for a under conditions of free and mixed convection, whereas Bailey and Meneses (1993) reported this not to be the case. The question arises if the differences in experimental setups (e.g., artificial leaves vs. plant leaves, laminar air flow vs. turbulent air flow) allow a direct cross-comparison of the numeric constants. The application of active thermography requires constant ambient conditions during the induced T_L -shift and the subsequent cooling process, and consequently this approach is only applicable in (semi)-controlled conditions. As mentioned above, we measured a small increase of background temperature after application of the heat pulse (0.3–0.4 K). However, background temperature remained stable during the subsequent cooling phase. Thus, we conclude that ambient conditions were kept sufficiently stable to allow reliable and repeatable measurements. Still, we recommend the use of an alternative heat source for future experiments. Especially shorter response times of the IR heating units would be desirable. Newly developed IR-LED panels may provide an improvement here.

In this study, we deliberately chose a realistic setup resembling natural conditions, relating active thermography to independent parameters, as for instance leaf transpiration. For further validation of our parameterization, we aimed to test our setup under more standardized conditions, including artificial reference materials, also helping us to calibrate our experimental setup. Measurements using differently wetted sheets of filter paper (Figure S10) confirmed the strong dependency of leaf cooling curves on the water content per area, as presented in Figure 1. However, there is no reference material available to mimic the highly variable structural and physiological properties of plant leaves entirely.

We conclude that active thermography provides a powerful tool in studying plant water relations and plant heat transfer processes when ambient conditions are monitored simultaneously and in high detail. The adaptability of the method towards semi-controlled conditions opens the way to new applications of active thermography in plant sciences, as for example in large-scale green-house phenotyping facilities.

AUTHOR CONTRIBUTIONS

HA: designed and performed the experiments and analyzed the data, conceived the project and wrote the article with contributions of all authors. FF: supervised the experiments, supervised and complemented the writing. RP: supervised the experiments, supervised and complemented the writing. MM-L:

analyzed the data and complemented the writing. CJ: complemented the writing, designed and performed further experiments and analyzed the data. LS: supervised and complemented the writing. US: supervised and complemented the writing. UR: supervised the experiments, supervised and complemented the writing.

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SUPPLEMENTARY MATERIALS

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fpls.2019.01684/full#supplementary-material>

REFERENCES

- Asner, G. (1998). Biophysical and biochemical sources of variability in canopy reflectance. *Remote Sens. Environ.* 253, 234–253. doi: 10.1016/S0034-4257(98)00014-5
- Bailey, B. J., and Meneses, J. F. (1993). Modelling leaf convective heat transfer. *Acta Horticult.* 399, 191–198. doi: 10.17660/ActaHortic.1995.399.22
- Blum, A. (2009). Effective use of water (EUW) and not water-use-efficiency (WUE) is the target of crop yield improvement under drought stress. *Field Crop Res.* 112, 119–123. doi: 10.1016/j.fcr.2009.03.009
- Bunce, J. A. (1985). Effect of boundary layer conductance on the response of stomata to humidity. *Plant Cell Environ.* 8, 55–57. doi: 10.1111/j.1365-3040.1985.tb01209.x
- Cannon, J. N., Krantz, W. B., and Kreith, F. (1979). A study of transpiration from porous flat plates simulating plant leaves. *Int. J. Heat Mass Transfer* 22, 469–483. doi: 10.1016/0017-9310(79)90013-9
- Dannenhoffer, J. M., Ebert, W. J., and Evert, R. F. (1990). Leaf vasculature in barley, *Hordeum vulgare* (Poaceae). *Am. J. Bot.* 77, 636–652. doi: 10.1002/j.1537-2197.1990.tb14449.x
- Defraeye, T., Verboven, P., Ho, Q. T., and Nicolai, B. (2013). Convective heat and mass exchange predictions at leaf surfaces: applications, methods and perspectives. *Comput. Electron. In Agric.* 96, 180–201. doi: 10.1016/j.compag.2013.05.008
- Dixon, M., and Grace, J. (1983). Natural convection from leaves at realistic Grashof numbers. *Plant Cell Environ.* 6, 665–670. doi: 10.1111/1365-3040.ep11589240
- Fanourakis, D., Heuvelink, E., and Carvalho, S. M. P. (2015). Spatial heterogeneity in stomatal features during leaf elongation: an analysis using *Rosa hybrida*. *Funtcional Plan Biol.* 42, 737–745. doi: 10.1071/FP15008
- Farrar, K., Bryant, D. N., Turner, L., Gallagher, J. A., Thomas, A., Farrell, M., et al. (2011). Breeding for bio-ethanol production in *Lolium perenne* L.: association of allelic variation with high water-soluble carbohydrate content. *Bioenergy Res.* 5, 149–157. doi: 10.1007/s12155-011-9156-0
- Foley, J. A., Costa, M. H., Delire, C., Ramankutty, N., Costaz, M. H., and Snyder, P. (2003). Green surprise? How terrestrial ecosystems could affect earth's climate. *Front. Ecol. Environ.* 1, 38–44. doi: 10.2307/3867963
- Gates, D. M. (1965). Energy, plants, and ecology. *Ecology* 46, 1–13. doi: 10.2307/1935252
- Gates, D. M. (1968). Transpiration and leaf temperature. *Annu. Rev. Plant Physiol.* 19, 211–238. doi: 10.1146/annurev.pp.19.060168.001235
- Gausman, H. W., and Allen, W. A. (1973). Optical parameters of leaves of 30 plant species. *Plant Physiol.* 52, 57–62. doi: 10.1104/pp.52.1.57
- Grace, J. (1974). The effect of wind on grasses 1. *Cuticular Stomatal Transpiration J. Exp. Bot.* 25, 542–551. doi: 10.1093/jxb/25.3.542
- Jayalakshmy, M. S., and Philip, J. (2010). Thermophysical properties of plant leaves and their influence on the environment temperature. *Int. J. Thermophysics* 31, 2295–2304. doi: 10.1007/s10765-010-0877-7
- Jones, H. G. (1992). *Plants and microclimate* (Cambridge: Cambridge University Press).
- Kümmerlen, B., Dauwe, S., Schmundt, D., and Schurr, U. (1999). “Thermography to Measure Water Relations of Plant Leaves,” in *Handbook of Computer Vision and Applications, Volume 3: Systems and Applications*, (London: Academic Press), 763–7881.
- Kitano, M., and Eguchi, H. (1990). Buoyancy effect on forced convection in the leaf boundary layer. *Plant Cell Environ.* 13, 965–970. doi: 10.1111/j.1365-3040.1990.tb01987.x
- Kumar, A., and Barthakur, N. (1971). Convective heat transfer measurements of plants in a wind tunnel. *Boundary-Layer Meteorol.* 2, 218–227. doi: 10.1007/BF00192131
- Leigh, A., Sevanto, S., Ball, M. C., Close, J. D., Ellsworth, D. S., Knight, C. A., et al. (2012). Do thick leaves avoid thermal damage in critically low wind speeds? *New Phytol.* 194, 477–487. doi: 10.1111/j.1469-8137.2012.04058.x
- Linacre, E. (1972). Leaf temperatures, diffusion resistances, and transpiration. *Agric. Meteorol.* 10, 365–382. doi: 10.1016/0002-1571(72)90038-6
- McKown, A. D., Cochard, H., and Sack, L. (2010). Decoding leaf hydraulics with a spatially explicit model: Principles of venation architecture and implications for its evolution. *Am. Nat.* 175, 447–460. doi: 10.1086/650721
- Monteith, J. L., and Unsworth, M. H. (2008). *Principles of environmental physics* (Burlington, USA: Elsevier Academic Press).
- Mott, K. A., and Peak, D. (2011). Alternative perspective on the control of transpiration by radiation. *Proc. Natl. Acad. Sci. U. S. A. (PNAS)* 108, 19820–19823. doi: 10.1073/pnas.1113878108
- Mott, K. A., Denne, F., and Powell, J. (1997). Interactions among stomata in response to perturbations in humidity. *Plant Cell Environ.* 20, 1098–1107. doi: 10.1046/j.1365-3040.1997.d01-138.x
- Munns, R., James, R. A., Sirault, X. R. R., Furbank, R. T., and Jones, H. G. (2010). New phenotyping methods for screening wheat and barley for beneficial responses to water deficit. *J. Exp. Bot.* 61, 3499–3507. doi: 10.1093/jxb/erq199
- Nardini, A., Gortan, E., Ramani, M., and Salleo, S. (2008). Heterogeneity of gas exchange rates over the leaf surface in tobacco: an effect of hydraulic architecture? *Plant Cell Environ.* 3, 804–812. doi: 10.1111/j.1365-3040.2008.01798.x
- Nelder, J., and Mead, R. (1965). A simplex method for function minimization. *Comput. J.* 7, 308–313. doi: 10.1093/comjnl/7.4.308

- Nobel, P. S. (2009). *Physicochemical and environmental plant physiology* (Oxford, UK: Elsevier Academic Press).
- Parkhurst, D. F. (1976). Effects of verbascum thapsus leaf hairs on heat and mass transfer: a reassessment. *New Phytol.* 76, 453–457. doi: 10.1111/j.1469-8137.1976.tb01481.x
- Parlange, J. Y., and Waggoner, P. E. (1971). Boundary layer resistance and temperature distribution on still and flapping leaves: I. Theory and laboratory experiments. *Plant Physiol.* 48, 437–442. doi: 10.1104/pp.50.1.60
- Passioura, J. B., and Angus, J. F. (2010). Improving productivity of crops in water-limited environments. *Adv. Agron.* 106, 37–67. doi: 10.1016/S0065-2113(10)06002-5
- Pearman, G. I., Weaver, H. L., and Tanner, C. B. (1972). Boundary layer heat transfer coefficients under field conditions. *Agric. Meteorol.* 10, 83–92. doi: 10.1016/0002-1571(72)90009-X
- Penman, H. L. (1948). Natural evaporation from open water, bare soil and grass. *Proc. R. Soc. A: Math. Phys. Eng. Sci.* 193, 120–145. doi: 10.1098/rspa.1948.0037
- Pieruschka, R., Huber, G., and Berry, J. A. (2010). Control of transpiration by radiation. *Proc. Natl. Acad. Sci. U. S. A. (PNAS)* 107, 13372–13377. doi: 10.1073/pnas.0913177107
- Raschke, K. (1960). Heat transfer between the plant and the environment. *Annu. Rev. Plant Physiol.* 11, 111–126. doi: 10.1146/annurev.pp.11.060160.000551
- Roelfsema, M. R. G., and Hedrich, R. (2005). In the light of stomatal opening: new insights into 'the Watergate'. *New Phytol.* 167, 665–691. doi: 10.1111/j.1469-8137.2005.01460.x
- Roth-Nebelsick, A. (2001). Computer-based analysis of steady-state and transient heat transfer of small-sized leaves by free and mixed convection. *Plant Cell Environ.* 24, 631–640. doi: 10.1046/j.1365-3040.2001.00712.x
- Sack, L., and Scoffoni, C. (2013). Leaf venation: structure, function, development, evolution, ecology and applications in the past, present and future. *New Phytol.* 198, 983–1000. doi: 10.1111/nph.12253
- Saldin, T. F., and Barthakur, N. (1971). Heat transfer between Phaseolus vulagris and the environment. *Can. J. Bot.* 49, 833–838. doi: 10.1139/b71-123
- Sampoux, J. P., Baudouin, P., Bayle, B., Béguier, V., Buordon, P., Chosson, J. F., et al. (2011). Breeding perennial grasses for forage usage: an experimental assessment of trait changes in diploid perennial ryegrass (Lolium perenne L.) cultivars released in the last four decades. *Field Crops Res.* 123, 117–129. doi: 10.1016/j.fcr.2011.05.007
- Schreuder, M. D. J., Brewer, C. A., and Heine, C. (2001). Modelled influences of non-exchanging trichomes on leaf boundary layers and gas exchange. *J. Theor. Biol.* 210, 23–32. doi: 10.1006/jtbi.2001.2285
- Schuepp, P. H. (1993). Leaf boundary layers. *New Phytol.* 59, 477–507. doi: 10.1111/j.1469-8137.1993.tb03898.x
- Schymanski, S. J., and Or, D. (2015). Wind increases leaf water use efficiency. *Plant Cell Environ.* 39, 1448–1459. doi: 10.1111/pce.12700
- Seelig, H. D., Hoehn, A., Stodieck, L. S., Klaus, D. M., Adams, III W.W., and Emery, W. J. (2008). The assessment of leaf water content using leaf reflectance ratios in the visible, near- and short-wave-infrared. *Int. J. Remote Sens.* 29, 3701–3713. doi: 10.1080/01431160701772500
- Seelig, H. D., Hoehn, A., Stodieck, L. S., Klaus, D. M., Adams, III W.W., and Emery, W. J. (2009). Plant water parameters and the remote sensing R1300/R1450 leaf water index: controlled conditions dynamics during the development of water deficit stress. *Irrig. Sci.* 27, 357–365. doi: 10.1007/s00271-009-0152-5
- Shibuya, T., Tsuruyama, J., Kitaya, Y., and Kiyota, M. (2006). Enhancement of photosynthesis and growth of tomato seedlings by forced ventilation within the canopy. *Sci. Hortic.* 109, 218–222. doi: 10.1016/j.scienta.2006.04.009
- Shimazaki, K., Doi, M., Assmann, S. M., and Kinoshita, T. (2007). Light regulation of stomatal movement. *Annu. Rev. Plant Biol.* 58, 219–247. doi: 10.1146/annurev.arplant.57.032905.105434
- Sinclair, R. (1970). Convective heat transfer from narrow leaves. *Aust. J. Biol. Sci.* 23, 309–322. doi: 10.1071/BI9700309
- Ueno, O., Kawano, Y., Wakayama, M., and Takeda, T. (2006). Leaf vascular systems in C3 and C4 grasses: a two-dimensional analysis. *Ann. Bot.* 97, 611–621. doi: 10.1093/aob/mcl010
- Vogel, S. (2009). Leaves in the lowest and highest winds: temperature, force and shape. *New Phytol.* 183, 13–26. doi: 10.1111/j.1469-8137.2009.02854.x

Conflict of Interest: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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