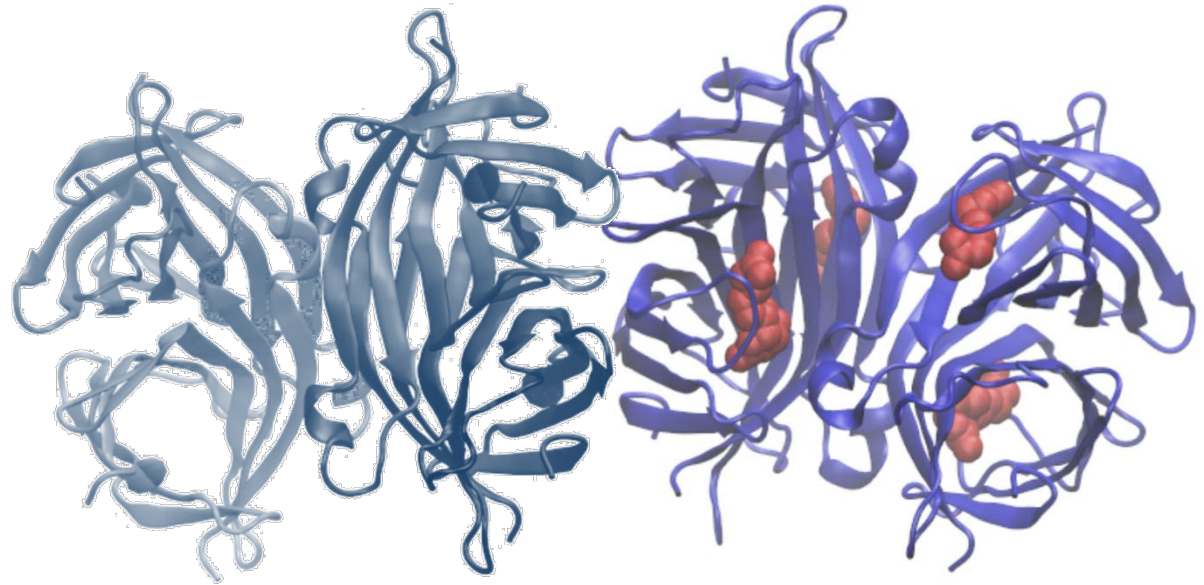
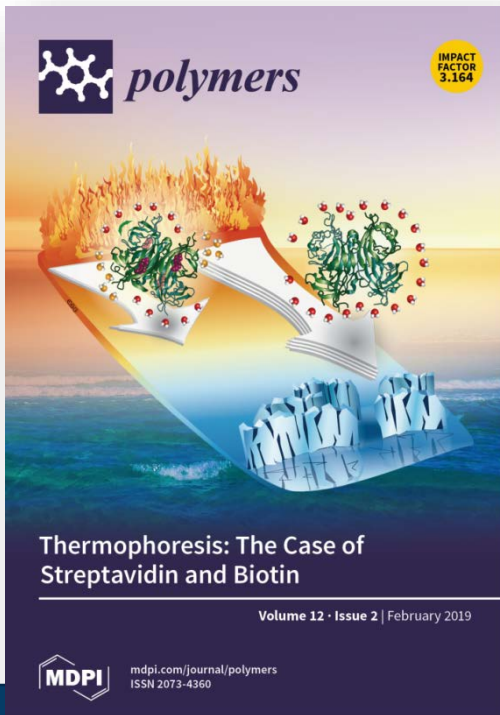




THERMOPHORESIS: THE CASE OF STREPTAVIDIN AND BIOTIN

19.11.2020 | DOREEN NIETHER, MONA SARTER, BERND W. KÖNIG, JÖRG FITTER, ANDREAS M. STADLER, SIMONE WIEGAND

2020 EUSMI User Meeting

THERMOPHORESIS

Movement of particles driven by a temperature gradient

$$\vec{j} = -\rho D \vec{\nabla} w - w(1-w)\rho D_T \vec{\nabla} T$$

steady state $\vec{j} = 0$

$$\frac{D_T}{D} = - \frac{\vec{\nabla} w}{w(1-w)\vec{\nabla} T}$$

$$S_T = \frac{D_T}{D} \propto \frac{\Delta w}{\Delta T}$$

D

diffusion coefficient

w

weight fraction

ρ

mass density

D_T

thermodiffusion coefficient

$\vec{j}$

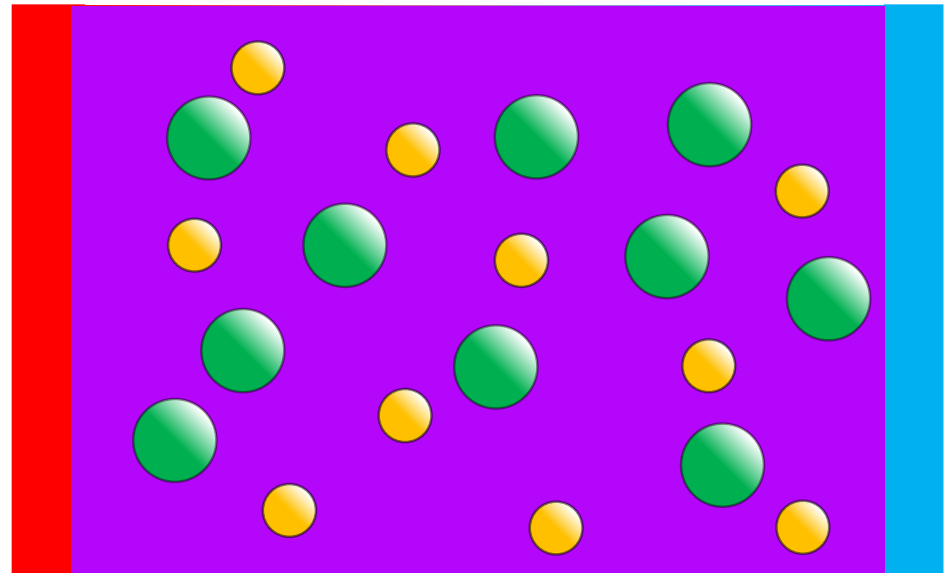
flux

T

temperature

S_T

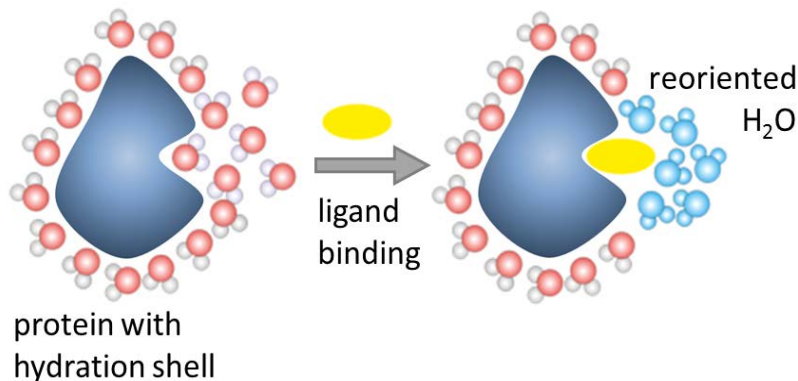
Soret coefficient



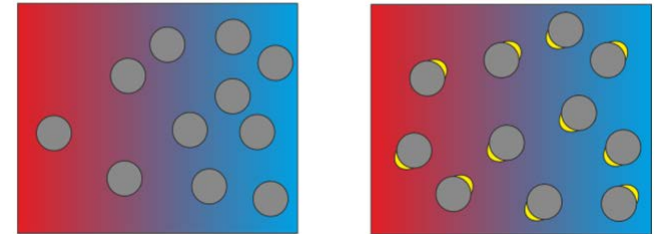
MOTIVATION

Microscale thermophoresis (MST)

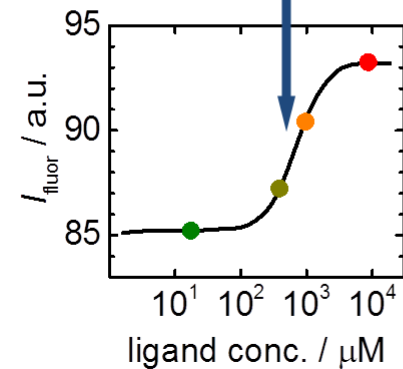
- method to determine kinetic constant of binding reactions
- protein's response to thermal gradient changes when ligand binds
- detected through change in fluorescence intensity during titration
- change due to modification of hydration shell



thermodiffusion changes



equilibration constants of biochemical reactions

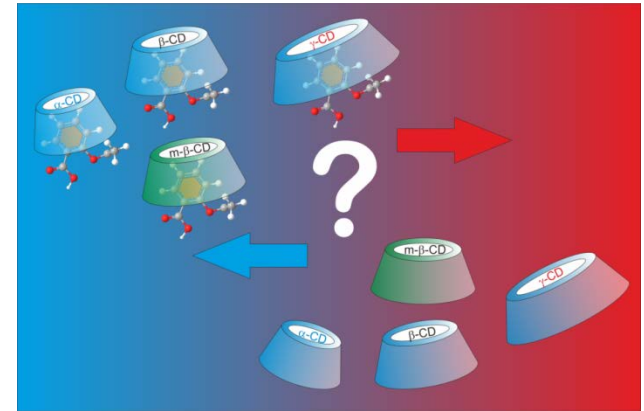


quantitative information about hydration shell?

titration curve:
 $\Delta I_{\text{fluor}} \propto \Delta S_T$

Hydration has strong influence on thermophoretic response.

- Can this connection be quantified?
- Can it be used to gain information about change in hydration shell upon complex formation?



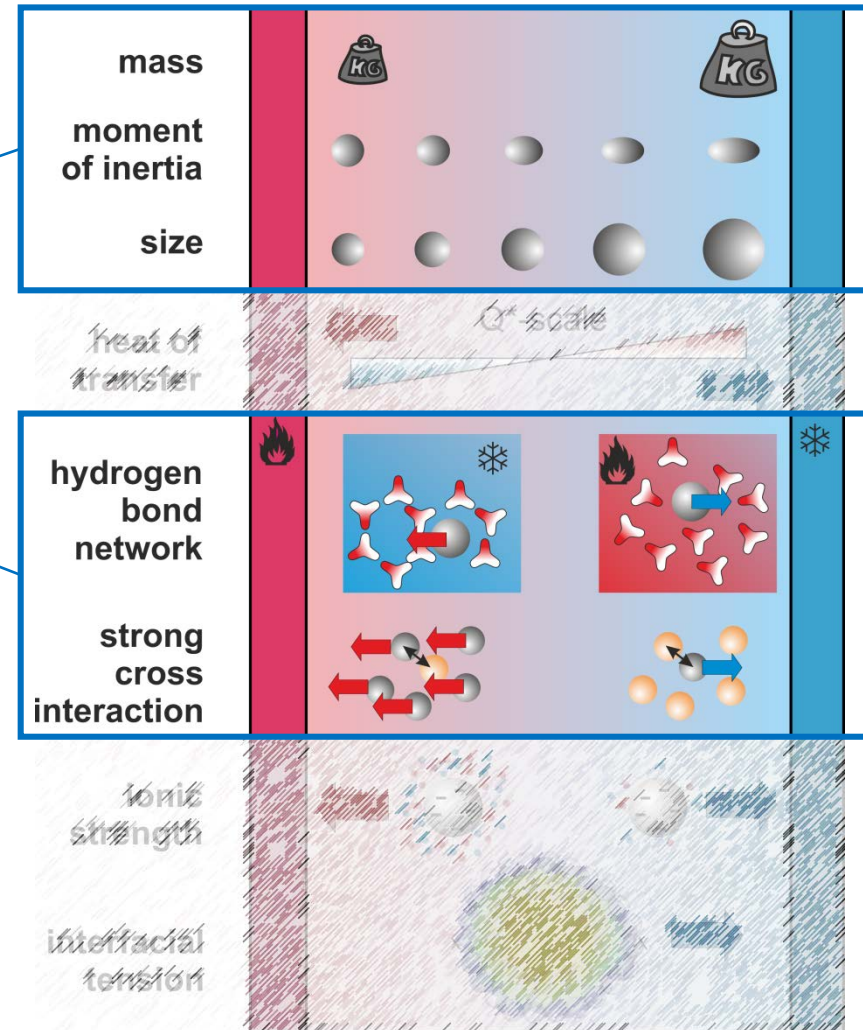
OPEN QUESTIONS

CONTRIBUTIONS

Aqueous systems

$$S_T \approx S_T^i + S_T^{chem}$$

- aqueous systems
- S_T influenced by hydrogen bonds
 - HB network of water
 - HB between solute and water
- T -dependence of HB $\Leftrightarrow$ thermodiffusion

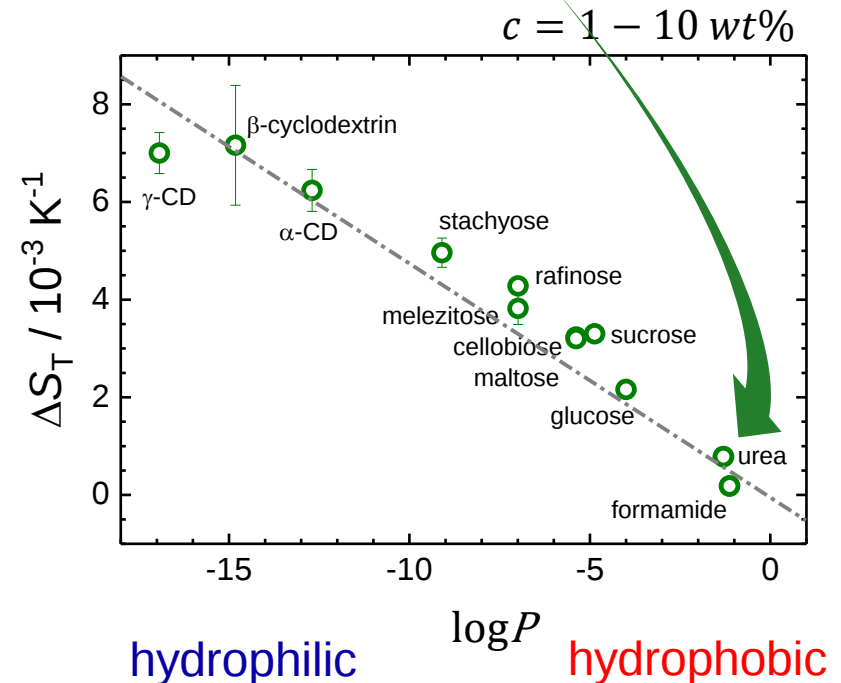
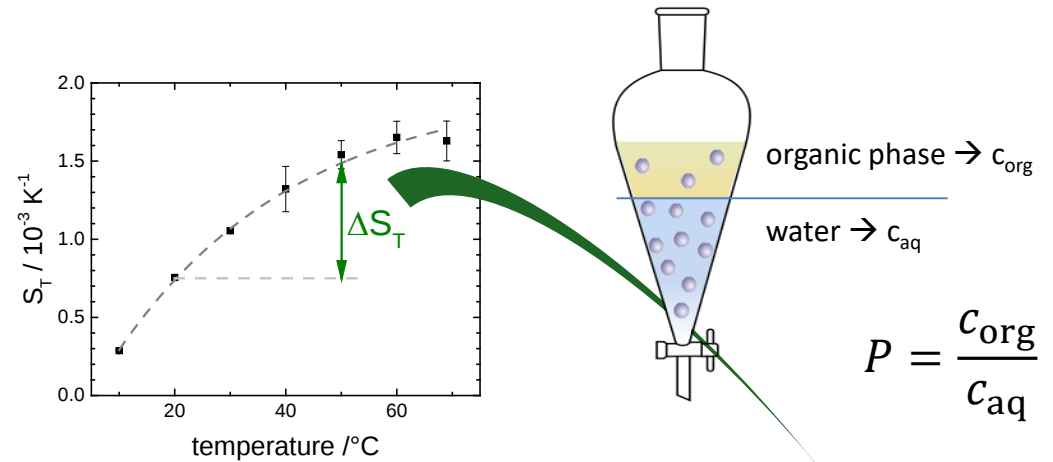


RESULTS

Correlation with $\log P$

$$\Delta S_T = S_T(50^\circ\text{C}) - S_T(20^\circ\text{C})$$

- ΔS_T is measure for temperature dependence $\rightarrow$ proportional to chemical contribution S_T^{chem}
- ΔS_T correlates with hydrophobicity ($\log P$)
- connection between hydration and thermodiffusion

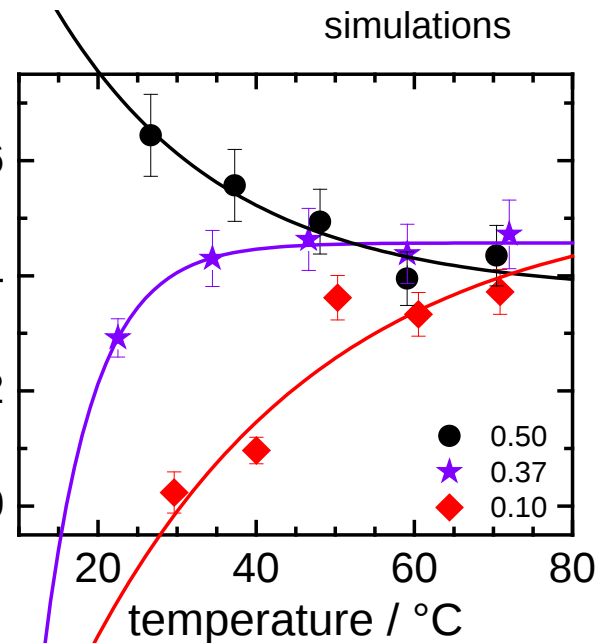
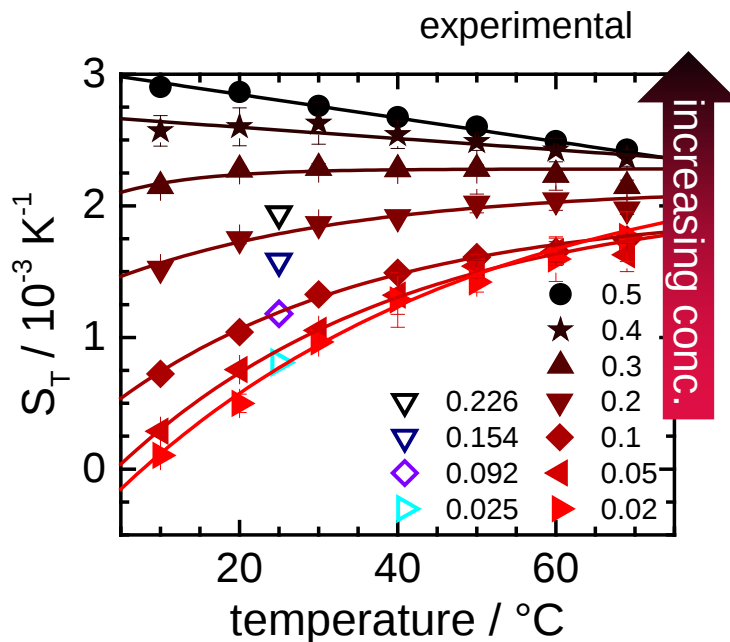
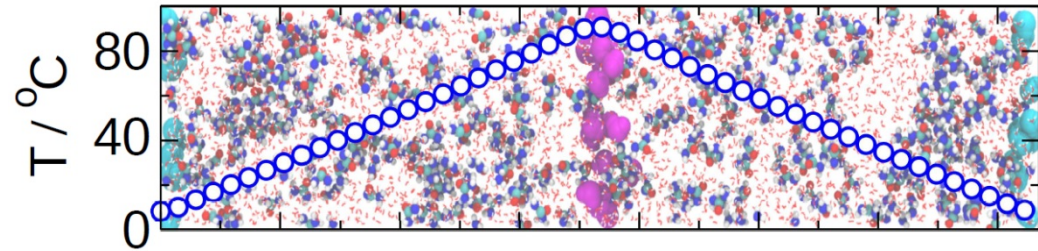


saccharides: P. Blanco et al., J. Phys. Chem. B (2010)
 cyclodextrins: K. Eguchi et al., Eur. Phys. J. E (2016)
 urea: D. Niether et al., PCCP. (2018)
 formamide: D. Niether et al., PNAS (2016)

UREA + WATER

NEMD-simulations – microscopic understanding

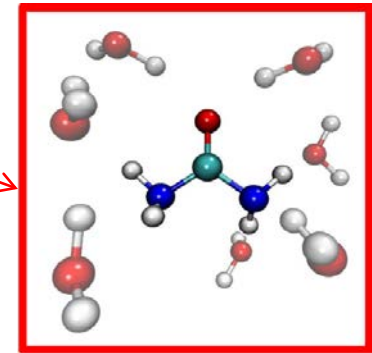
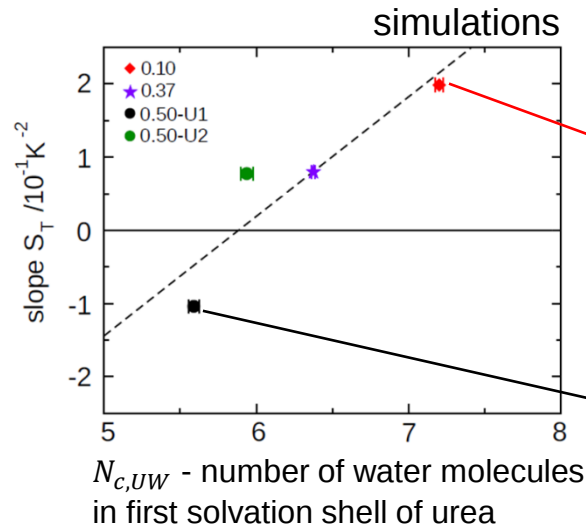
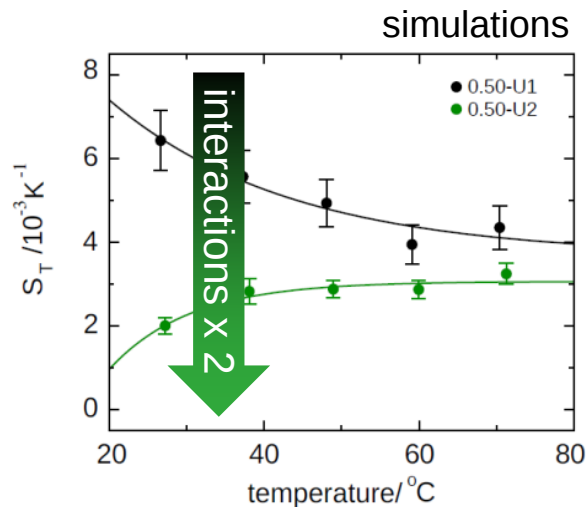
- non-equilibrium molecular dynamics simulations
- T -dependence of S_T decreases with rising concentration



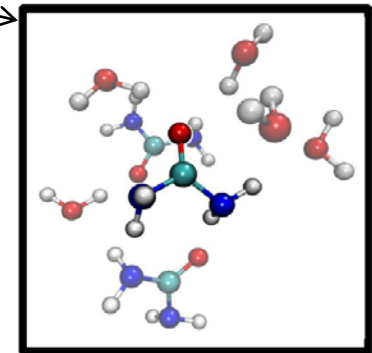
UREA + WATER

NEMD-simulations – microscopic understanding

NEMD simulations show
that interactions
between **urea** and
water decrease with
rising urea conc.



w.f. = 0.10



w.f. = 0.50

increasing U-W
interactions: slope goes
from negative to positive

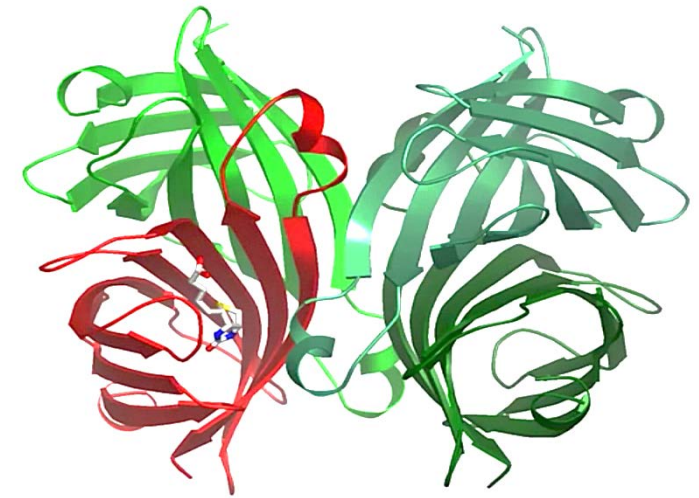
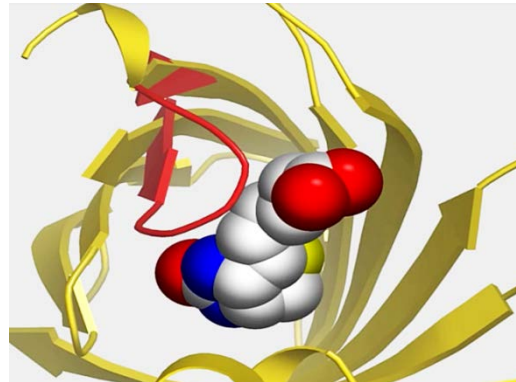
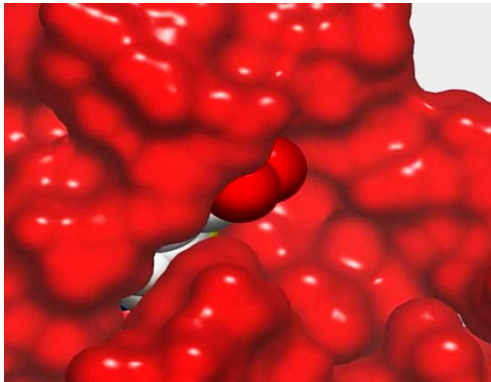
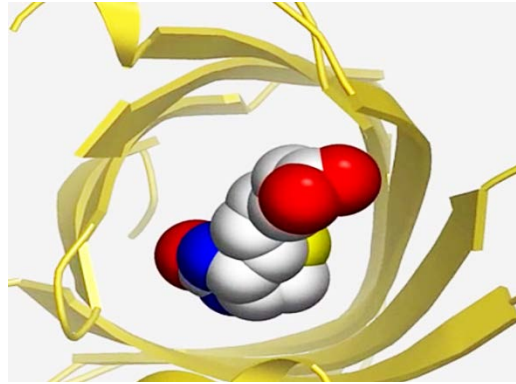
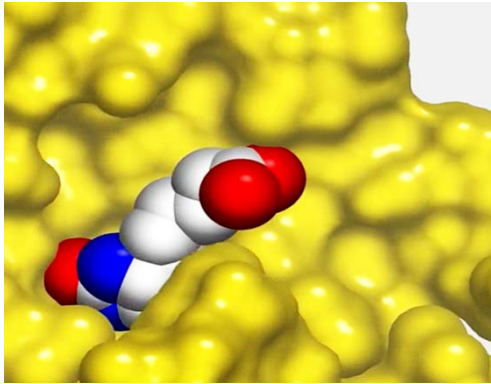
HYDROPHILICITY AND S_T

$$S_T \approx S_T^i + S_T^{chem}$$


- dominated by hydrogen bonds in aqueous systems
→ proportional to ΔS_T (T-dependence of S_T)
- Correlation with logP
 - Hydrophilic substances → high ΔS_T
 - Hydrophobic substances → low/negative ΔS_T

STREPTAVIDIN + BIOTIN

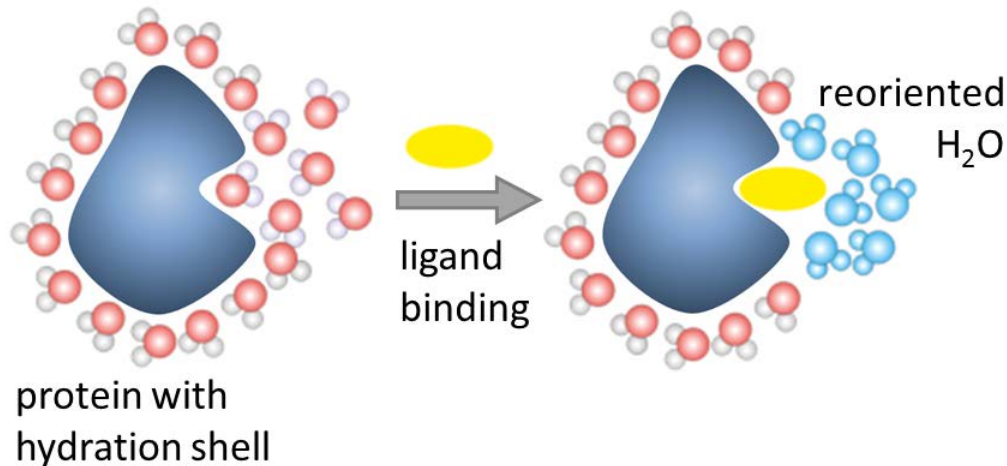
Protein-ligand system



streptavidin tetramer

PROTEIN LIGAND INTERACTIONS

Influence of hydration layer



ΔH enthalpy: forces in the protein

- VdW interactions
- H-bonding
- screened charges

ΔS entropy: number of accessible states

$$\Delta S = \Delta S_{\text{protein}} + \Delta S_{\text{hydration}}$$

< 0 ?

ligand binding determined by change of free energy

$$\Delta G = G^{\text{bound}} - G^{\text{free}} = \Delta H - T\Delta S$$

COMBINING THREE DIFFERENT METHODS TO UNDERSTAND THE BINDING

ITC

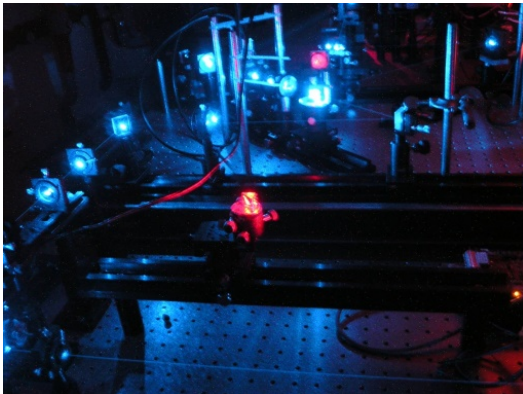


measures the entropy and free enthalpy changes due to the binding

QENS at SPHERES [Garching]



neutron back scattering probes dynamic processes



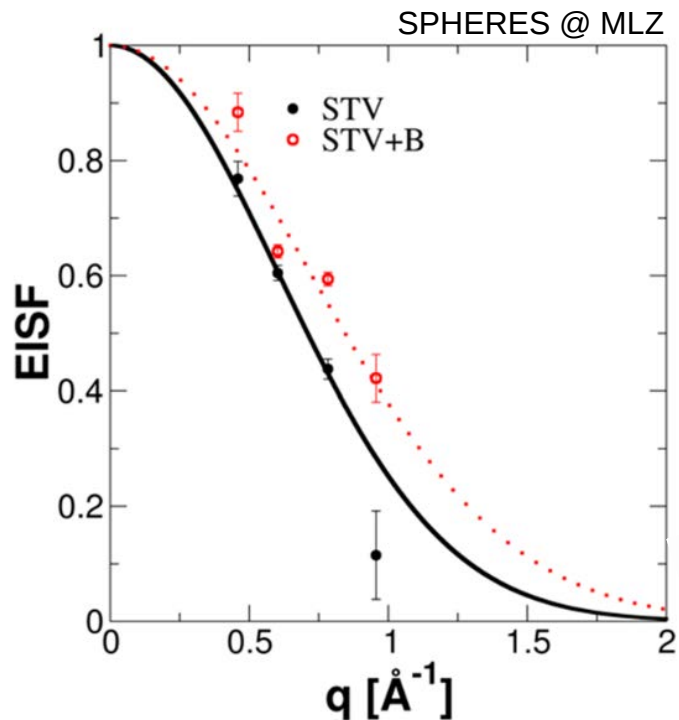
TDFRS

measures the diffusion of solute molecules in a temperature gradient

QUASI-ELASTIC NEUTRON SCATTERING

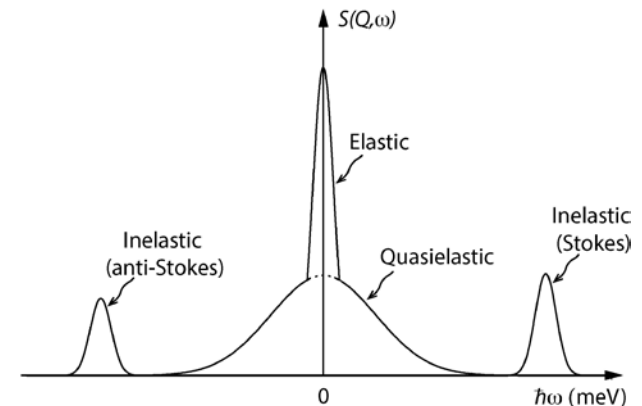
Determination of protein dynamics in solution

Andreas Stadler and Mona Sarter,
ICS-1, JCNS and RWTH Aachen



EISF –
elastic
incoherent
structure
factor →
amplitudes
of motion

measured in D₂O: only dynamic of **protein**



Quasi-elastic scattering:

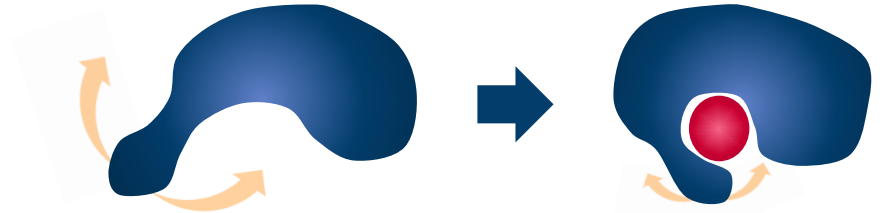
- small energy exchange between neutron and particle
- processes with distribution of energies (translations, rotations, ...)

$$A_0(q) = \exp(-\langle x^2 \rangle * q^2)$$

CONFORMATIONAL ENTROPY

Reduction of protein conformational entropy due to biotin binding

$$\Delta S_{conf} = 3R \cdot \ln \left[\sqrt{\frac{\langle x^2 \rangle_{bound}}{\langle x^2 \rangle_{free}}} \right]$$



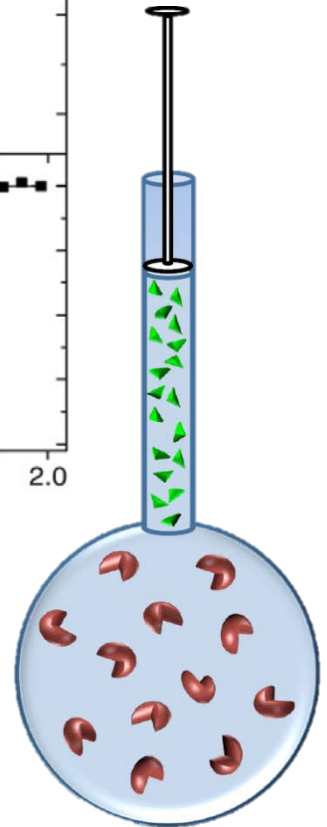
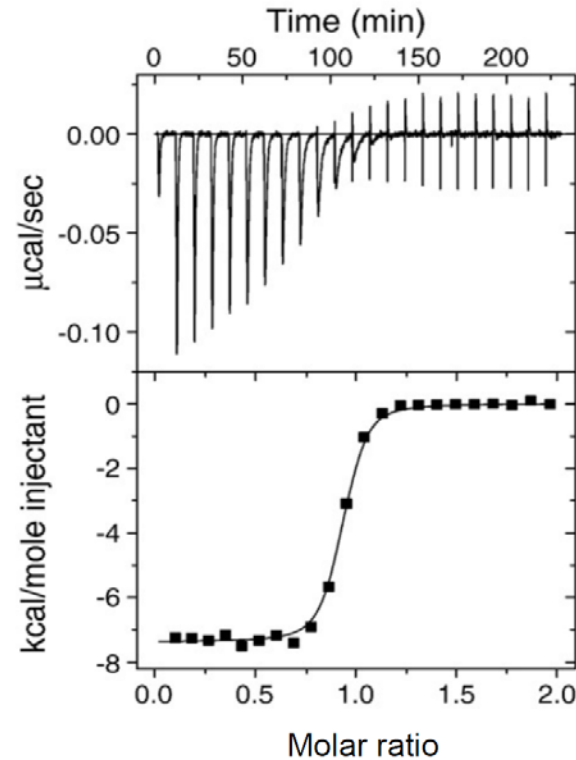
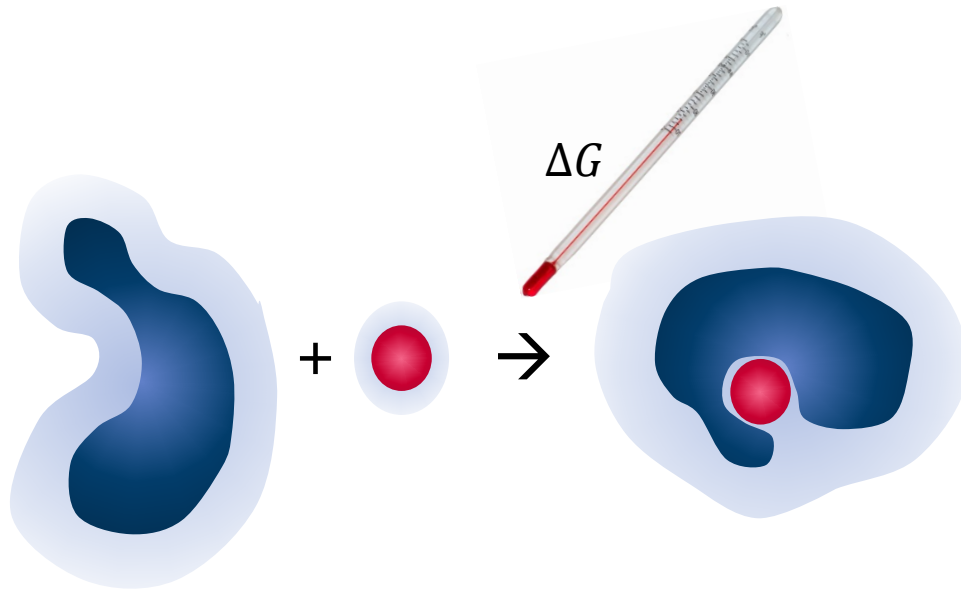
- streptavidin is more mobile than streptavidin+biotin complex
- fast motions attributed to side-chains
- conformational entropy negative on ps to ns time-scale
- entropy change free protein → complex:

$$\Delta S^{QENS} = -2.2 \pm 0.3 \text{ kJ mol}^{-1} \text{ K}^{-1}$$

Time-scale measured	$\Delta S_{conf} \left[\frac{\text{kJ}}{\text{mol K}} \right]$
ns	-2.2 ± 0.3
ps	-1.3 ± 0.2

ITC

Isothermal titration calorimetry

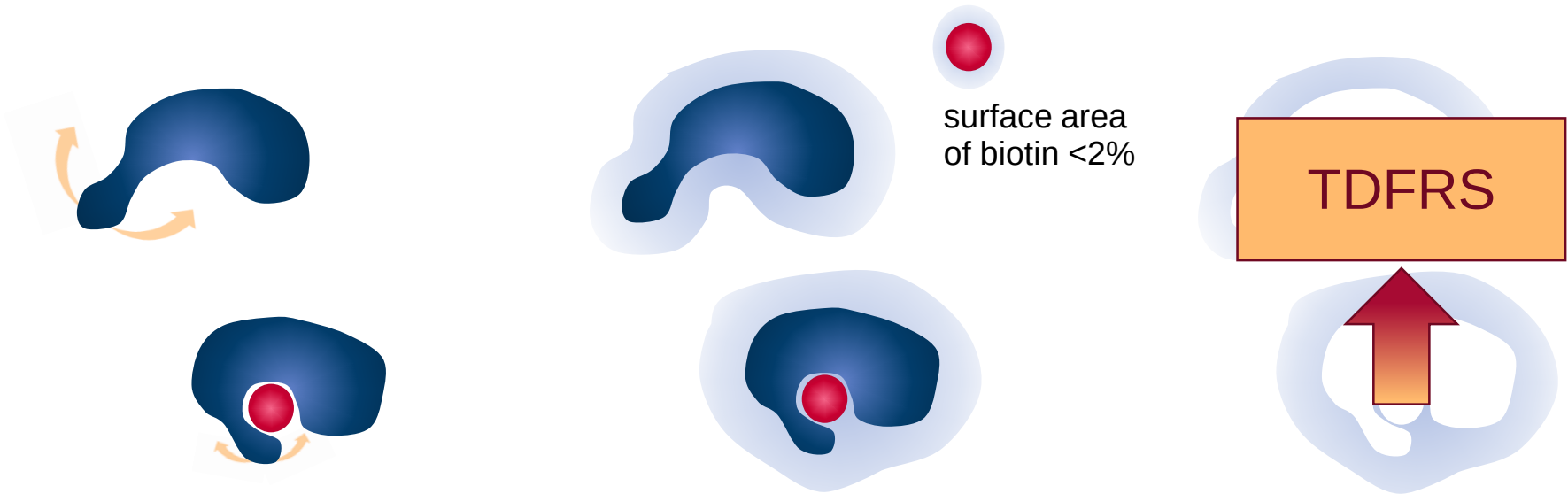


measures free energy change upon complex formation (**protein + hydration shell**)

$$\Delta S^{ITC} = -0.35 \pm 0.13 \text{ kJ mol}^{-1} \text{ K}^{-1}$$

RESULTS

Streptavidin + biotin

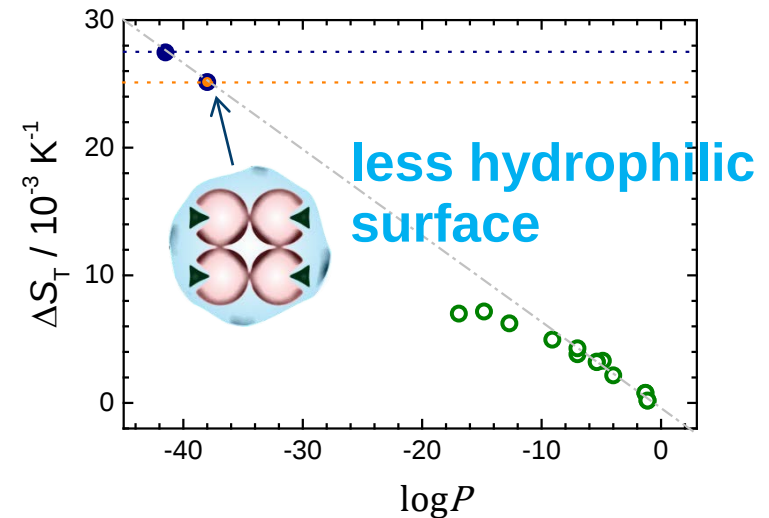
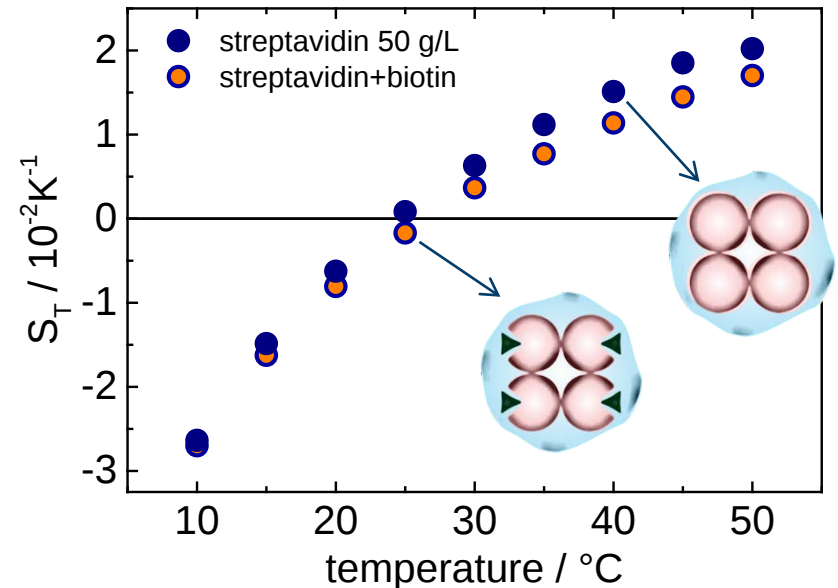
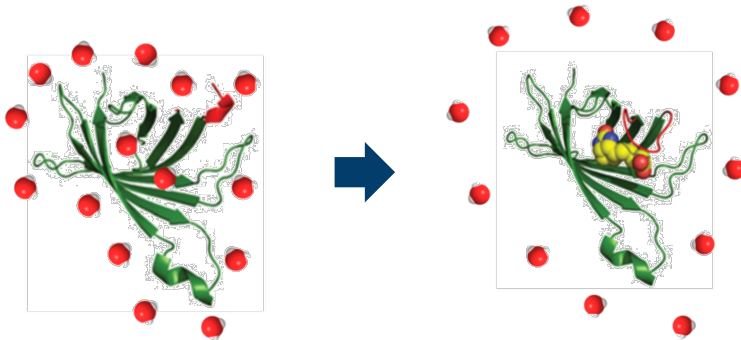


QENS	ITC	Difference
$\Delta S^{QENS} =$ $-2.2 \pm 0.3 \text{ kJ mol}^{-1} \text{ K}^{-1}$	$\Delta S^{ITC} =$ $-0.4 \pm 0.1 \text{ kJ mol}^{-1} \text{ K}^{-1}$	$\Delta S^{hs} = \Delta S^{ITC} - \Delta S^{QENS} =$ $1.8 \pm 0.4 \text{ kJ mol}^{-1} \text{ K}^{-1}$
entropy change of protein	entropy change of protein + hydration shell	→ entropy of hydration shell increases when biotin binds

RESULTS

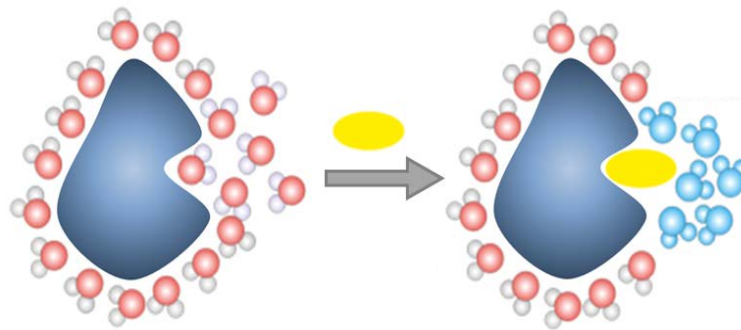
Streptavidin + biotin

- reduced ΔS_T for strep + biotin
→ surface of complex less hydrophilic
- indicates breaking of HB
- results from neutron scattering data:
 - protein-complex more rigid
 - entropy increase in hydration shell



SUMMARY

- Influence of hydration on S_T was investigated
- Conclusion on complex formation of streptavidin: disordered hydration layer compensates less flexible protein complex
- Clear change in ΔS_T when biotin binds on streptavidin, connection to entropy change in hydration shell



ACKNOWLEDGEMENT

Simone Wiegand, Jan K.G. Dhont, and the IBI-4

collaborators

Silvia Di Lecce and Fernando Bresme,
Imperial College London, UK – NEMD simulations

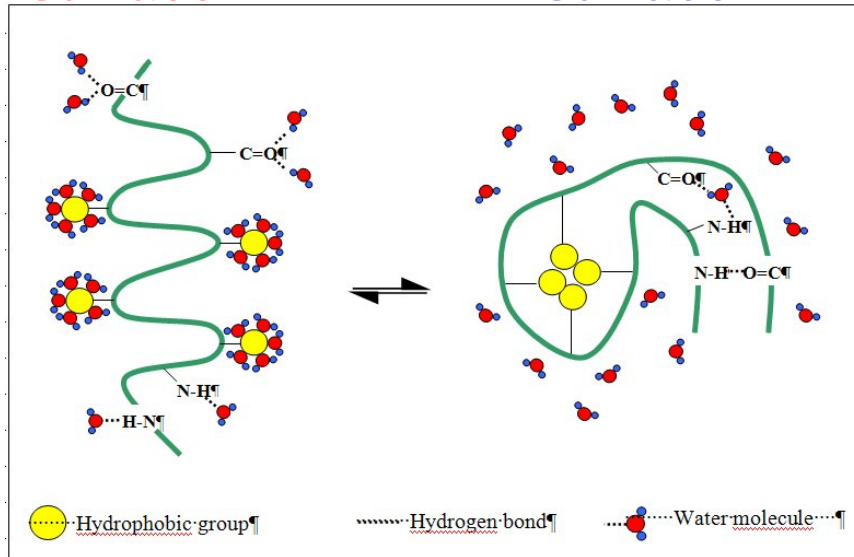
Mona Sarter, Andreas Stadler, Bernd König, and Jörg Fitter,
FZJ and RWTH Aachen – streptavidin QENS

Thank you for your attention!

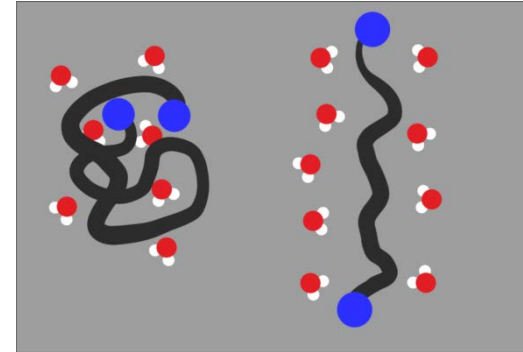
MICROSCOPIC PICTURE

hydrophobic
surface

hydrophilic
surface



- protein flexible
- hydration water less mobile
- protein less flexible
- hydration water mobile



flexible coil

stretched chain

lower entropic
contribution
from the
hydration shell

larger entropic
contribution from
the hydration
shell

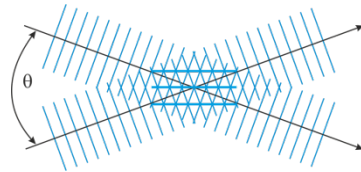
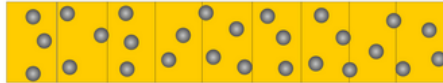
Gallat et al. Biophys J. 2012

Liese, S. et al., ACS Nano, **11** (2017) 702-712 22

SETUP

Infra-red Thermal Diffusion Forced Rayleigh Scattering (IR-TDFRS)

homogeneous
temperature
and particle
distribution

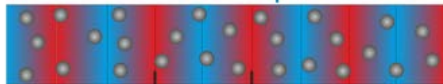


laser grating

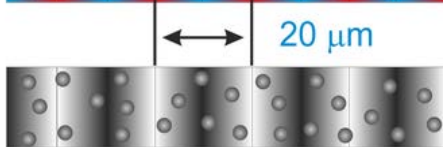


$$\Delta T = 20 - 100 \mu\text{K}$$

temperature
grating



↓
refractive index
grating

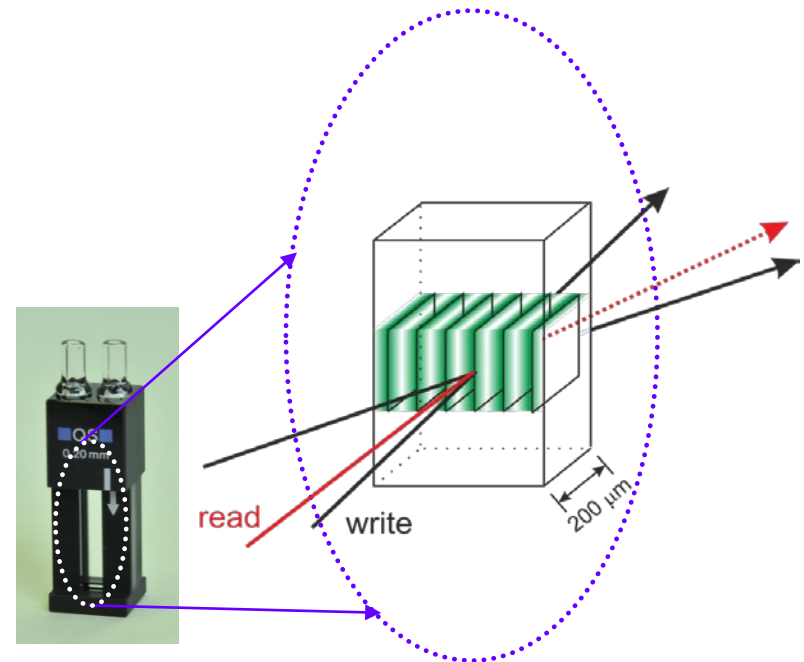


thermal diffusion

↓
concentration
grating



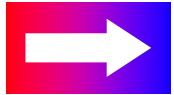
Measured quantity:
Intensity of the diffracted beam



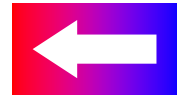
SETUP

Signal

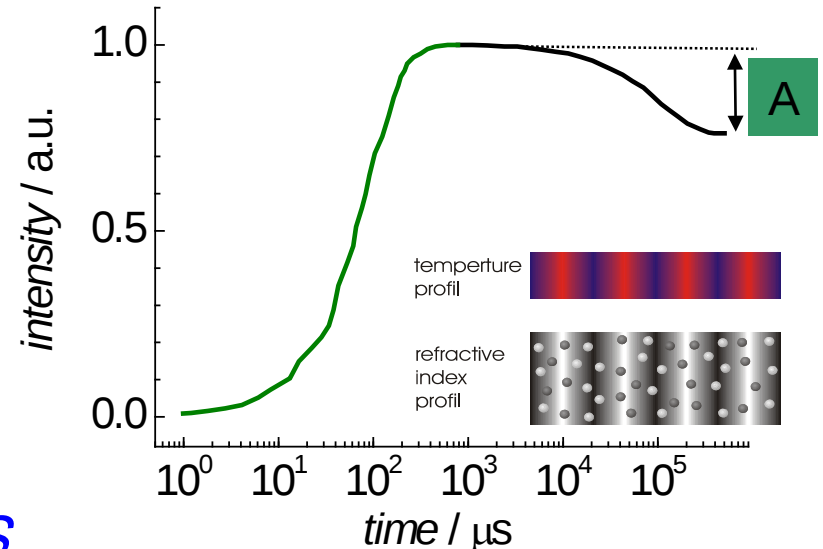
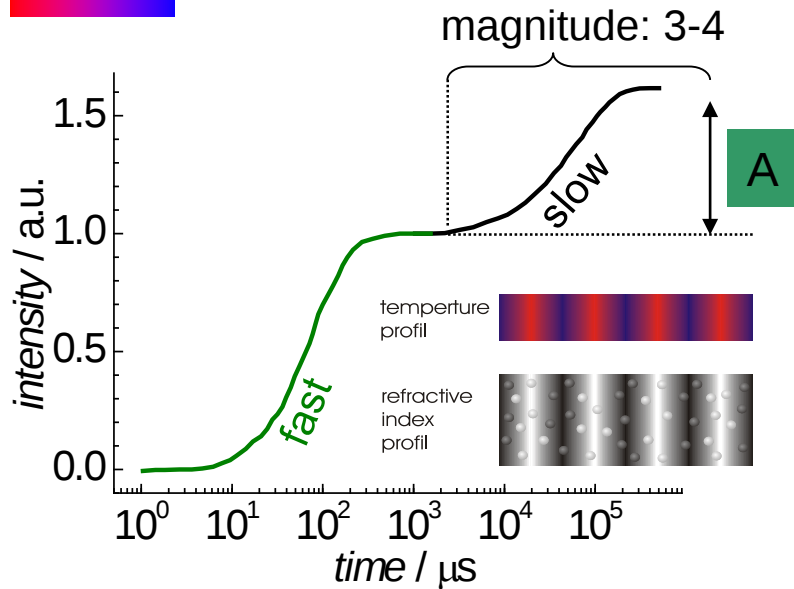
Measured quantity:
Intensity of the diffracted beam



Molecules/colloides with higher refractive index moves to...
...cold side



...warm side

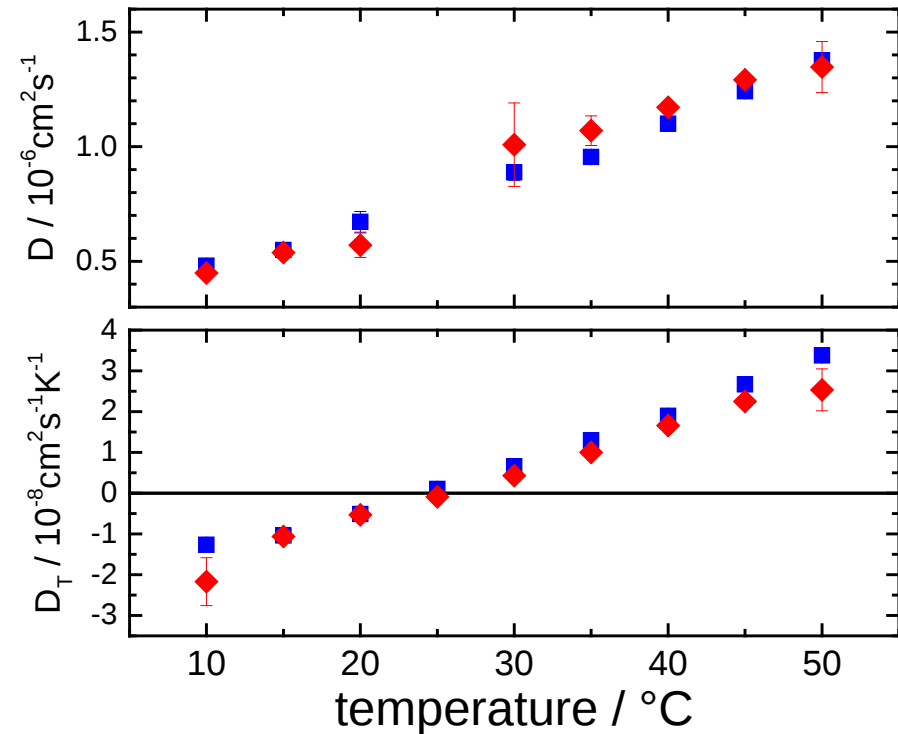
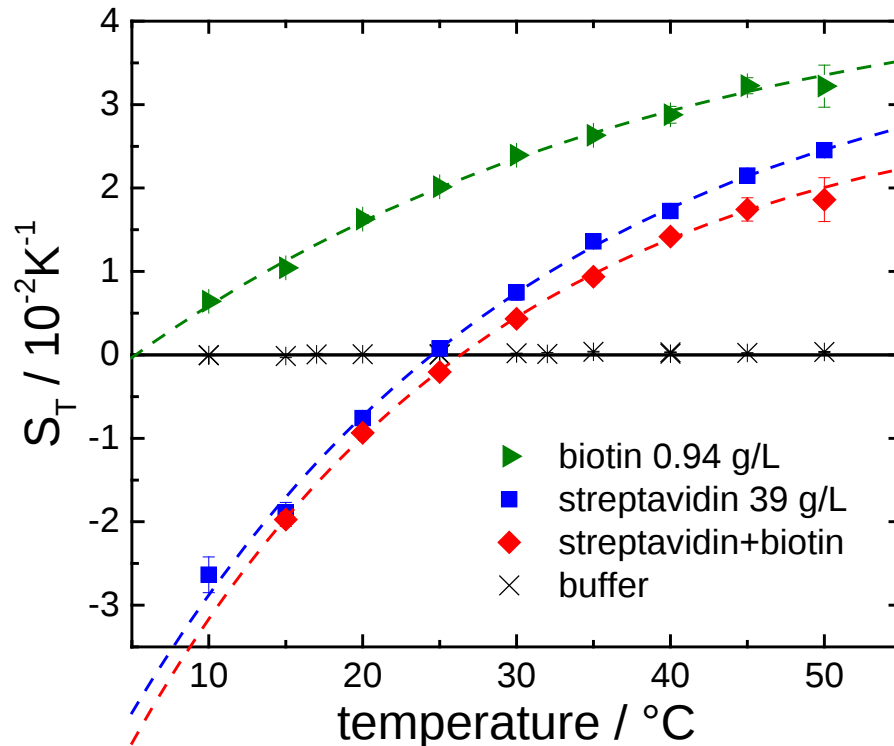
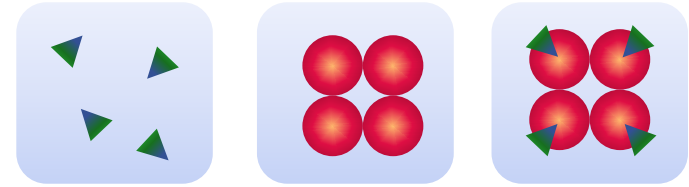


$$\zeta_{\text{het}} = \underbrace{\left(1 - e^{-t/\tau_{\text{th}}}\right)}_{\text{temperature}} - \underbrace{\frac{(\partial n / \partial c)_{p,T}}{(\partial n / \partial T)_{p,c}} \frac{D_T}{D} c(1-c)}_A \frac{1}{\tau - \tau_{\text{th}}} \left[\tau \left(1 - e^{-t/\tau}\right) - \tau_{\text{th}} \left(1 - e^{-t/\tau_{\text{th}}}\right) \right]$$

A

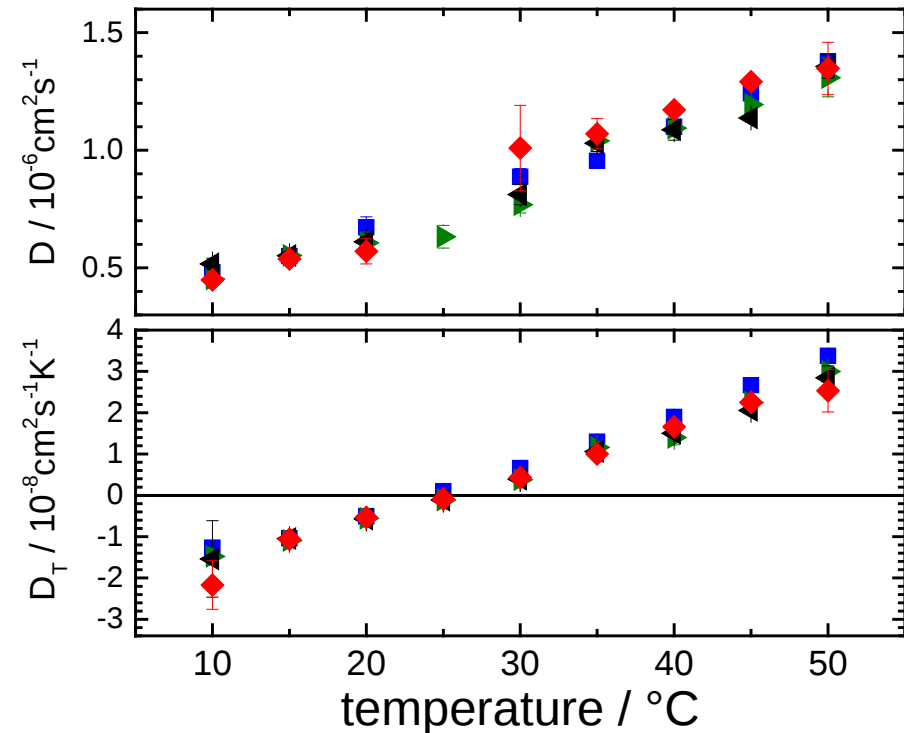
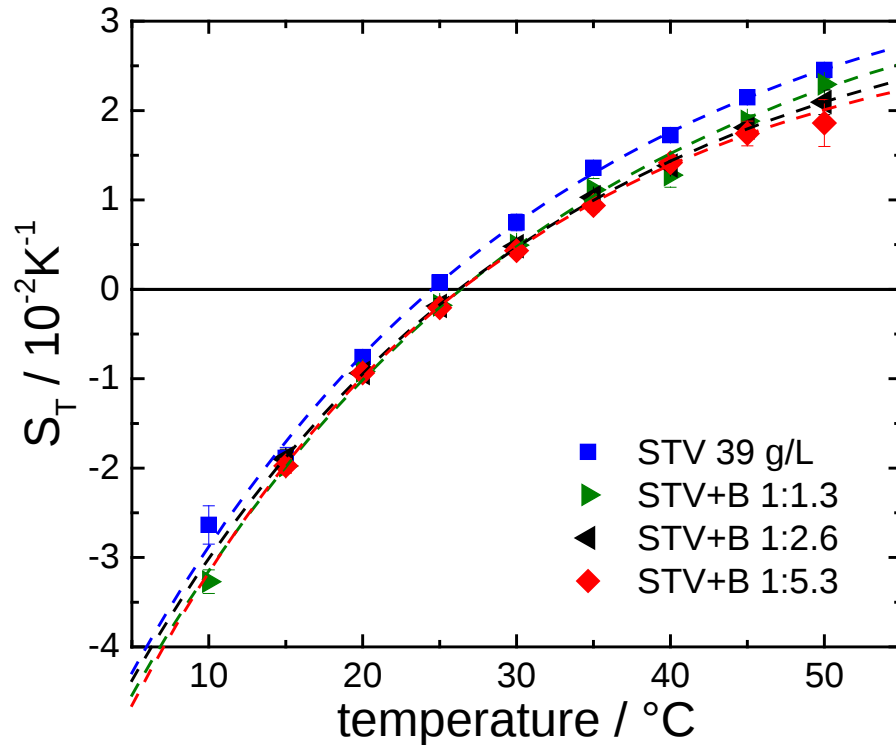
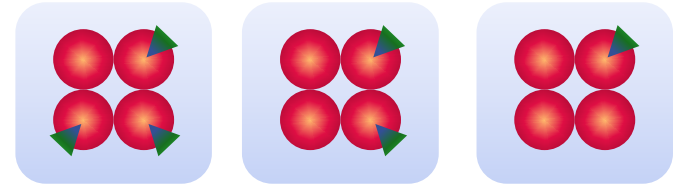
RESULTS

Streptavidin + biotin in H₂O-buffer



RESULTS

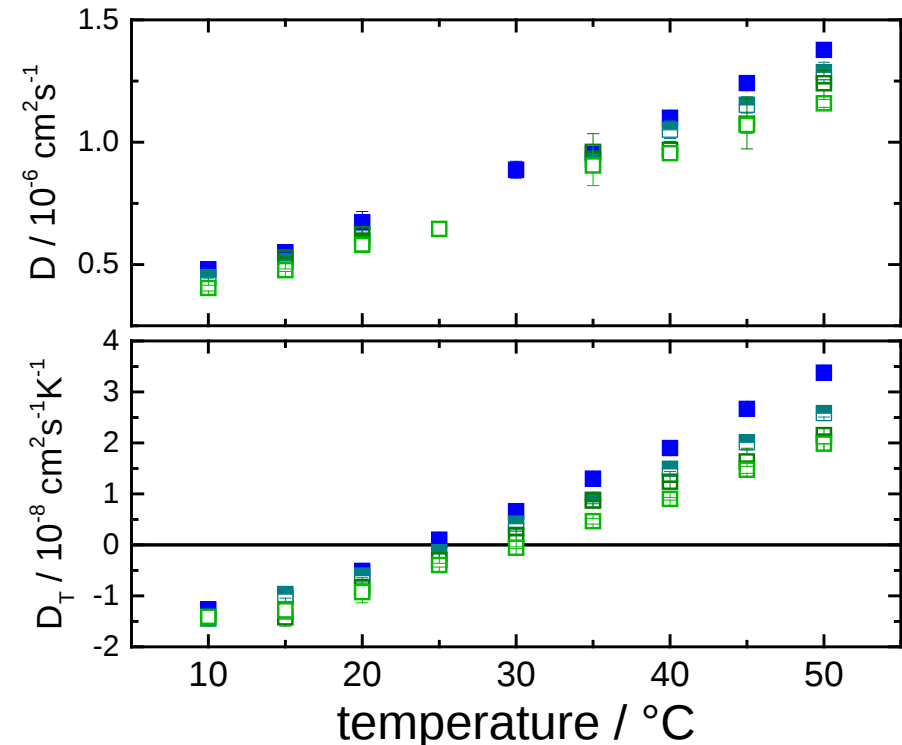
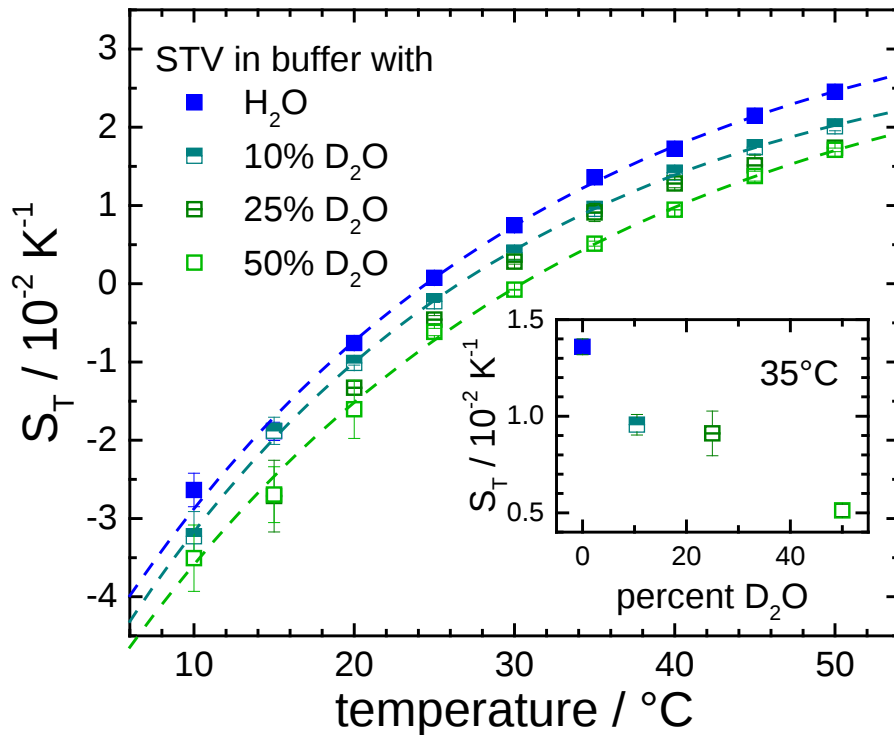
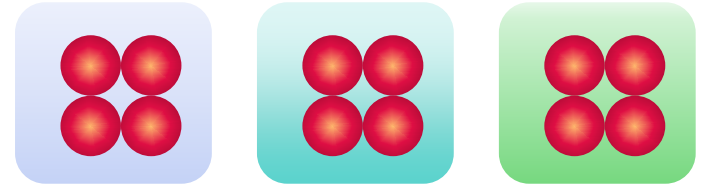
Streptavidin + biotin - stoichiometry



non-linear dependence on biotin concentration might point to cooperative binding

RESULTS

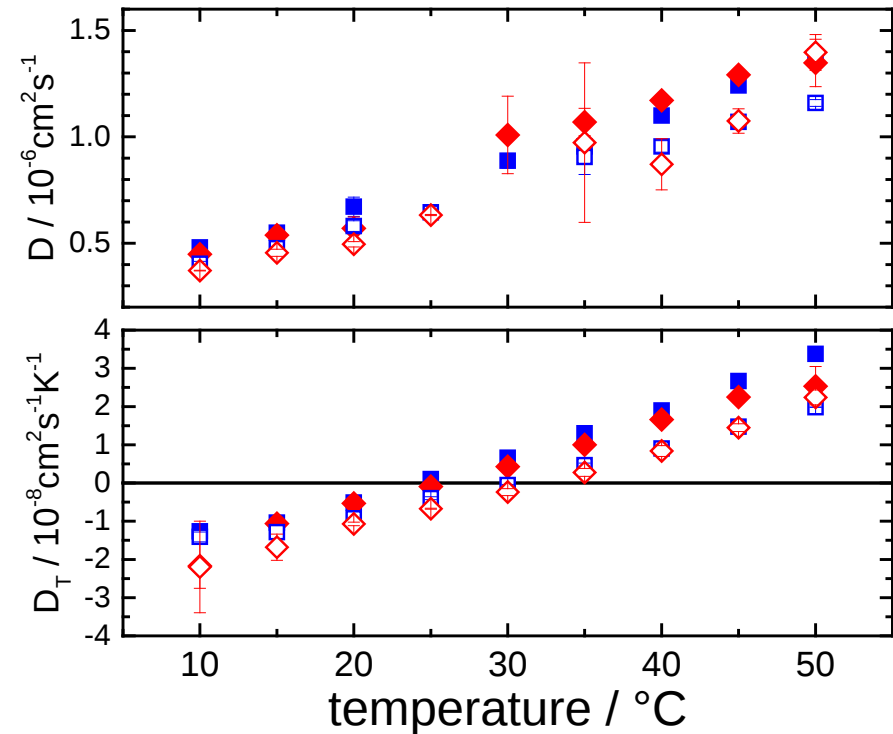
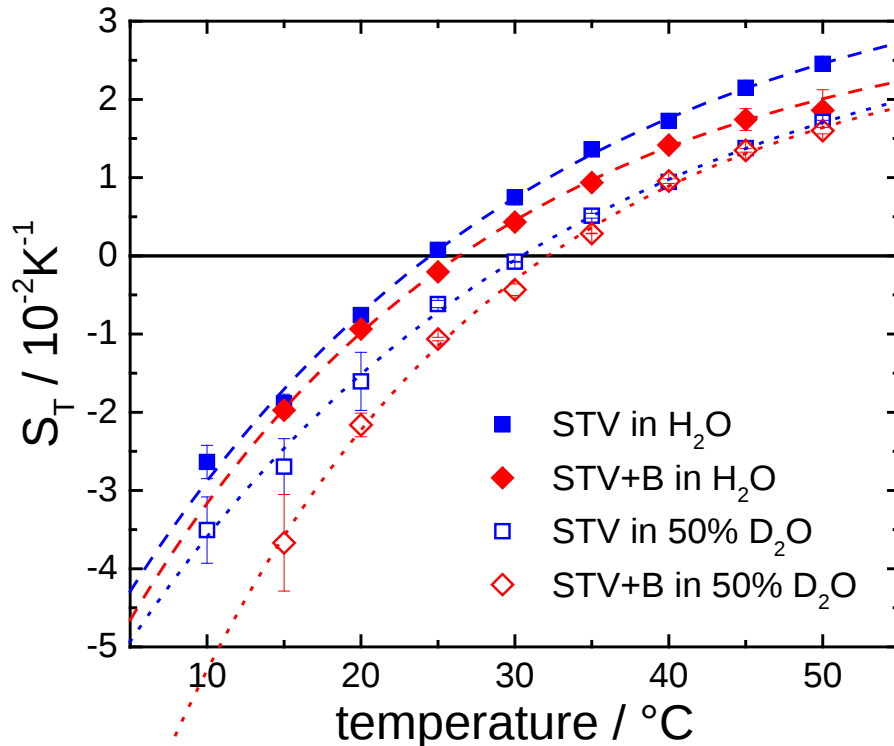
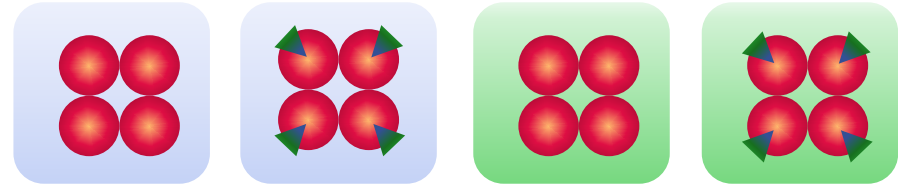
Streptavidin in buffer with D₂O



Measurement above ~50% D₂O not possible due to reduced IR-absorption of solvent

RESULTS

Streptavidin + biotin – H₂O and D₂O



It's likely that the behaviour of hydration shell differs between H₂O and D₂O