



Article

Flexible Sample Environments for the Investigation of Soft Matter at the European Spallation Source: Part I—The In Situ SANS/DLS Setup

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Abstract: As part of the development of the new European Spallation Source (ESS) in Lund (Sweden), which will provide the most brilliant neutron beams worldwide, it is necessary to provide different sample environments with which the potential of the new source can be exploited as soon as possible from the start of operation. The overarching goal of the project is to reduce the downtimes of the instruments related to changing the sample environment by developing plug and play sample environments for different soft matter samples using the same general carrier platform and also providing full software integration and control by just using unified connectors. In the present article, as a part of this endeavor, the sample environment for in situ SANS and dynamic light scattering measurements is introduced.

Keywords: dynamic light scattering; small angle neutron scattering; instrumentation; microgels

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1. Introduction

The new European Spallation Source [1] (ESS) in Lund (Sweden) will provide the most brilliant neutron beams worldwide. To make efficient use of this source, it is necessary to provide different sample environments with which the potential of the ESS can be exploited as soon as possible from the start of operation. Accordingly, in our opinion, now is the right time to construct and build the sample environments with the construction of "Day 1" instruments [2]. For this purpose, the expertise of three university-based groups from the field of polymer and colloid research was bundled with construction teams of the SKADI [3] and the LOKI [4] instrument and also with the sample environment group of the ESS. Three different "Day 1" sample environments were developed that use a common universal carrier system. This system was intended to enable maximum flexibility with

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minimal downtime caused by changing the sample environment. In addition, it should be possible to partially automate the conversion, at least in perspective, since higher radiation is to be expected at the sample location. The following three sample environments were developed as part of the joint project:

- Environment for in situ dynamic light scattering (DLS) combined with small angle neutron scattering;
- Environment for small-angle neutron scattering under grazing incidence (GISANS) [5];
- Environment for neutron scattering on foams [6].

In the first expansion step, these sample environments are to be used on ESS instruments that are optimized for elastic small angle neutron scattering (SANS) for volume samples and GISANS and reflectometry on interface samples. The necessary tests during the development phase were carried out on instruments from the MLZ (Heinz Maier–Leibnitz Zentrum) such as KWS-1 [7] and REFSANS [8]. An innovative concept for a modular sample geometry for the ESS was developed and technically implemented. In particular, the specific demands at the ESS are included in the planning.

In the project, the common model systems used to benchmark all sample environments are so-called smart microgels which are particles with tunable interaction potential. They are called "smart" due to their ability to respond to external stimuli leading to a steadily increasing number of studies dealing with these fascinating materials [9]. The major scientific question of this collaborative research endeavor addresses the interactions of these soft deformable particles in the volume phase and at interfaces. This is extremely important both with regard to the fundamental understanding of the interactions between the particles [10,11] and for the transfer to the life sciences, as well as for technical applications in new technologies [12–15].

The present work focuses on the sample environment for in situ dynamic light scattering (DLS) measurements in combination with small angle neutron scattering. SANS is a valuable method for the investigation of soft matter. Due to the high sensitivity of SANS for organic samples compared to X-ray scattering and the possibility to selectively set the contrast of the molecular species by selecting the isotopic composition of samples and solvent (so-called contrast matching), it is an extremely valuable method in many soft matter fields like studies of self-assembly [16,17], polymer melts [18,19], colloidal polymers such as microgels [20–27], micelles [28,29] and bio-macromolecules like proteins [30,31] or vesicles from lipids [17,32]. Due to the relatively low intensity of neutron beams, often rather long measurements, e.g. compared to light scattering are required to gain data of high quality.

Due to the combination of long measurements and irradiation with neutrons, a stable sample is not always guaranteed. Biological samples might be especially prone to degradation. Therefore, monitoring the sample stability during extended measurements with in situ DLS has recently gained popularity [33–37]. In this work, the components of the sample environment designed for the simultaneous measurement of SANS and DLS at the ESS with a focus on portability and the accommodation of many samples are presented (Section 2). A detailed description of the complete setup, the control software layout and the DLS data analysis can be found in the Section 3.

2. Sample Environment Components

In light of the high-intensity neutron source ESS, our setup features a sample changer for up to 39 samples with three individually temperature controlled compartments.

2.1. Sample Rack/Sample Magazines

The sample rack (Figure 1) holds three magazines for up to 13 standard 1 mm cuvettes (external dimensions: $52 \, \text{mm} \cdot 12.5 \, \text{mm} \cdot 3.5 \, \text{mm}$) each. The cuvettes can be flushed with dried air to prevent condensation when measuring at low temperatures. Each of the magazines has a separate connector (CBI, Stäubli, Pfäffikon, Switzerland) for temperature control fluid and is insulated with PEEK, so that three individual temperatures can be

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used simultaneously, provided enough thermostats are present. Within each magazine, three positions can be equipped with in-cell Pt-100 thermometers (P0K1.161.2K.Y.5000-4.S, Innovative Sensor Technology IST AG, Ebnat-Kappel, Schweiz). The remaining cuvette temperatures can then be interpolated. The Pt-100 thermometers are read out with a temperature monitor (Model 224, Lake Shore Cryotronics, Westerville, OH, USA). The sample holder temperature is controlled with fluid pumped by a thermostat (FP50-HL, Julabo, Seelbach, Germany).



Figure 1. Photograph of the sample changer. The sample changer consists of a rack for 3 magazines each holding up to 13 cuvettes. Each magazine's temperature can be controlled individually. The open sides can be flushed with dried air to prevent condensation, when measuring at low temperatures. This part of the setup can also be used for standard SANS experiments without DLS. The red arrows indicate the cuvette positions, that were used for the temperature measurements in Figure 2.

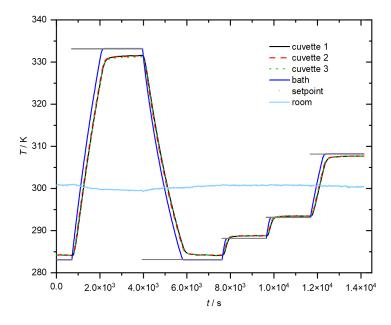


Figure 2. Comparison of set-point temperature, bath temperature and the temperature in three different cuvettes inside the sample holder plotted against time.

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The sample aperture is 9 mm \cdot 20 mm for the incident beam and 10 mm \cdot 25 mm on the back. The largest scattering angle possible with this sample environment is 42.5° in the vertical direction. However, it should be noted that the possible scattering angle also depends on the actual configuration of the SANS instruments, e.g., the neutron spot size. However, the sample-to-aperture distance will be a compromise to allow for the incident light to reach the sample cells. The sample environment can be moved along the Y- and Z-axis (in the instrument coordinate system) to change the sample in the neutron beam and laser focus. This is accomplished with two linear translation stages (LS-180, Physik Instrumente, Karlsruhe, Germany).

Temperature Control

Precise and reproducible temperature control is an important issue when working with soft matter samples, due to the influence on molecular interactions and the sample structure. Therefore, the sample holder constructed for this instrument is characterized regarding the temperature control capabilities. The sample environment's temperature can be controlled with various fluids, allowing a temperature range from 253.15 K to 393.15 K, depending on the chosen fluid.

An example of the temperature control characteristics is shown in Figure 2. Here, the set-point temperature, the bath temperature and the temperatures in three different cuvettes filled with water in one magazine (see Figure 1) are plotted against the time. It can be seen that the cuvette temperatures follow the bath temperature rapidly. The cuvette temperatures reach 1 K of their equilibrium temperatures after 1 min and 0.1 K of the equilibrium temperature within 3 min after the bath reached the setpoint temperature even for large temperature steps (50 K). The thermostat takes ca. 30 min to reach the setpoint for cooling 50 K from 333.1 K to 283.1 K. Heating from 283.1 K to 333.1 K takes 23 min. However, smaller temperature steps, from 283.1 K to 288.1 K only take 3 min. These times are dependent on the thermostat used and are only valid for the thermostat (FP50-HL, Julabo, Seelbach, Germany) applied here.

The sample temperature can be measured using a PT-100 thermometer inside three cuvettes for each magazine. Figure 2 shows that all three cuvette temperatures are nearly identical. This indicates a homogeneous temperature distribution for all samples in one magazine. However, to correct even for these slight deviations, the sample temperature for all other cuvettes can be interpolated from the three measured temperatures in each magazine.

2.2. Light Scattering

Dynamic light scattering is a well established method for the characterization of particles in suspension by probing their diffusive behavior [38].

For the dynamic light scattering setup inspired by [36,37], the light scattering plane is inclined by 20° relative to the SANS plane, to prevent problems arising from reflections on the cuvettes. A scheme of the optical setup can be found in Figure 3.

The laser light (633 nm, 21 mW, HNL210L-EC, Thorlabs, Newton, NJ, USA) passes a shutter (SH05/M, Thorlabs, Newton, NJ, USA) and a filter wheel (FW212C, Thorlabs, Newton, NJ, USA) equipped with neutral density filters with optical densities of 0.1, 0.2, 0.3, 0.4, 0.5, 0.6, 1.0, 1.3, 2.0 and 3.0 and is coupled into a polarization-maintaining single mode fiber (P3-630PM-FC-1, Thorlabs, Newton, NJ, USA) with a fiber port (PAF-X-5-A, Thorlabs, Newton, NJ, USA). The laser light is emitted from a fiber collimator (60FC-4-M5-33, Schäfter and Kirchhoff, Hamburg, Germany) and focused onto the sample through a Glan–Thompson polarizer (GTH10M-A, Thorlabs, Newton, NJ, USA). After passing another Glan–Thompson polarizer and a laser line filter (FL632.8-1, Thorlabs, Newton, NJ, USA) the light scattered by the sample is collected by the same collimator type and coupled into a single mode optical fiber (630HP with FC/PC and FC/APC connectors, Thorlabs, Newton, NJ, USA). For the detection in pseudo-cross configuration, a fiber optic beam splitter (ALV, Langen, Germany) is used to split the scattered light in a ratio of about 50:50.

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Afterwards, the light is detected into two single photon counting modules (SPCM-AQRH-14-FC, Excelitas Technologies, Waltham, MA, USA). For correlation, a field programmable gate array board (Spartan-6 FPGA, Xilinx, San José, CA, USA) is programmed analogously to a previous implementation for fluorescence correlation spectroscopy [39]. For the alignment of the setup, at first the incident laser beam is focused on the sample position. Then, an auxiliary laser is coupled into the fiber couplers of the detection side, to enable a prealignment by eye. After that, the detection can be easily adjusted to maximum scattered intensity and the quality of the correlation function.

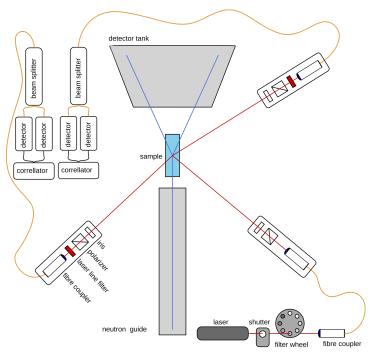


Figure 3. Scheme of the instrument components required for light scattering and the resulting beampath.

The sample environment for in situ dynamic light scattering and SANS measurements is equipped with two parallel setups for measuring dynamic light scattering at two different scattering angles θ of approximately 120° and 71°. Due to refraction on the cuvette–air and cuvette–solvent interfaces, the precise scattering angle is calculated using the refractive index n of the solvent according to Snell's law for each temperature.

To validate the correct operation of the dynamic light scattering setup, polystyrene standard spheres with a radius of 25 nm and 200 nm were measured. Figure 4 shows a autocorrelation function and the respective cumulant fit for each angle of measurement as an example. Here, sound autocorrelation functions with an intercept of about 1 indicate a well-adjusted light scattering setup. The measurement of an autocorrelation function is completed within 30 s. This enables multiple DLS experiments within the duration of a SANS experiment (some 10 min) and therefore the in situ monitoring of the sample stability.

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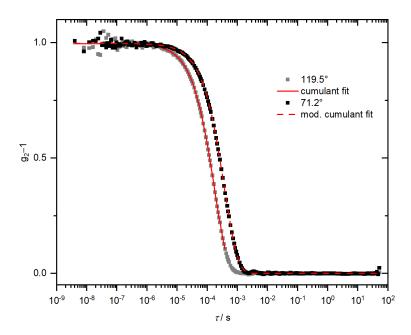


Figure 4. Two correlation functions measured simultaneously at both detection angles. The sample used here was a diluted 50 nm (diameter) polystyrene-sphere standard in H_2O at a temperature of 10.5 °C.

The relaxations rates Γ received from the fit can be converted into the translational diffusion coefficient D_T with the magnitude of the scattering vector $q = \frac{4\pi n}{\lambda} \sin(\frac{\Theta}{2})$. Here, λ is the wavelength, Θ is the scattering angle and n is the refractive index of the solvent.

$$D_T = \frac{\Gamma}{q^2} \tag{1}$$

Then, the hydrodynamic radius R_H is calculated with the Boltzmann constant k_b , the solvent viscosity [40] η and the temperature T using the Stokes–Einstein equation:

$$R_H = \frac{k_b T}{6\pi \eta D_T}. (2)$$

The hydrodynamic radii calculated this way for the particles are shown in Table 1. The measured hydrodynamic radii are consistently slightly bigger than the nominal particle size as indicated by the producer of the standard. This is a common effect in dynamic light scattering and can be attributed to the interaction of the particles with the solvent H_2O and maybe attractive interactions. These values are well in accordance with the measurements performed on these particles with other light scattering setups in our lab.

Table 1. Results of the dynamic light scattering measurements using two different polystyrene latex standards diluted in water with nominal radii of 25 nm and 200 nm, respectively.

	25 nm	200 nm
71°	$29.9\pm0.1~\mathrm{nm}$	$209 \pm 6\mathrm{nm}$
120°	$28.5\pm0.2~\mathrm{nm}$	$215\pm3~\mathrm{nm}$

To test the dynamic light scattering setup at different temperatures, microgels are a suitable and interesting model system, due to the temperature-dependent swelling behavior. Figure 5 shows the swelling behavior of a poly N-isopropylmethacrylamide (NIPMAM) microgel crosslinked with the molecule N,N'-methylenebisacrylamide. The data were

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measured at both angles using the SANS/DLS sample environment. The fully swollen microgel at 12 °C exhibits a hydrodynamic radius of 150 nm. With increasing temperature, the hydrodynamic radius decreases—at first continuously, then drastically from 132 nm at 39 °C to 83 nm at 50 °C with the typical volume phase transition temperature of 45 °C for NIPMAM microgels. Within the experimental precision, the results agree perfectly with previous measurements on commercial DLS machines [26,27]. To further illustrate this agreement, Figure 5, right, shows the correlation functions of the NIPMAM microgel measured at 56 °C with the SANS/DLS setup compared to correlation functions obtained at the same q-values with a commercial setup (ALV GmbH, Langen, Germany) equipped with a Nd:YAG-laser ($\lambda = 532\,\text{nm}$). The data are in excellent agreement. Moreover, no compressed exponential decays were observed for the microgel samples measured here, pointing to the absence of convection [41].

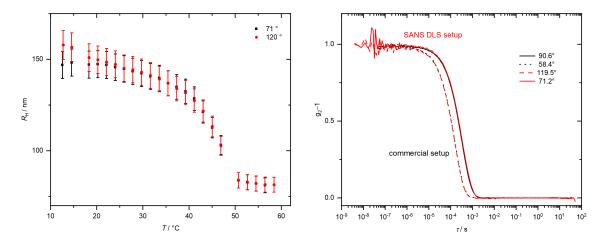


Figure 5. (**left**): Swelling behavior of a NIPMAM microgel measured with the SANS/DLS setup at both scattering angles. (**right**): comparison of autocorrelation functions of the NIPMAM microgel at 56 °C for both angles (red) compared to ACFs measured with a commercial ALV setup (black curves, measured with a different laser wavelength and therefore at different angles to achieve the same q value) at the same q-values of 0.23 nm⁻¹ and 0.15 nm⁻¹.

2.3. SANS Measurements

To validate the sample environment design for SANS measurements, tests were performed at the MLZ. A sample was measured in the newly constructed sample environment and compared to neutron data previously recorded using the D11 SANS machine at the Institute Laue Langevin (ILL). Figure 6 shows the results of a measurement using the sample holder introduced in this work at the KWS-1 [7] SANS instrument at the MLZ. Figure 6, top, shows a comparison of measurements of the same NIPMAM microgel as investigated in Figure 5, performed as the reference at the ILL using the instrument D11 and with the new sample environment at MLZ. The scattering curves show a satisfactory agreement. The slight deviation at low q can be attributed to resolution effects.

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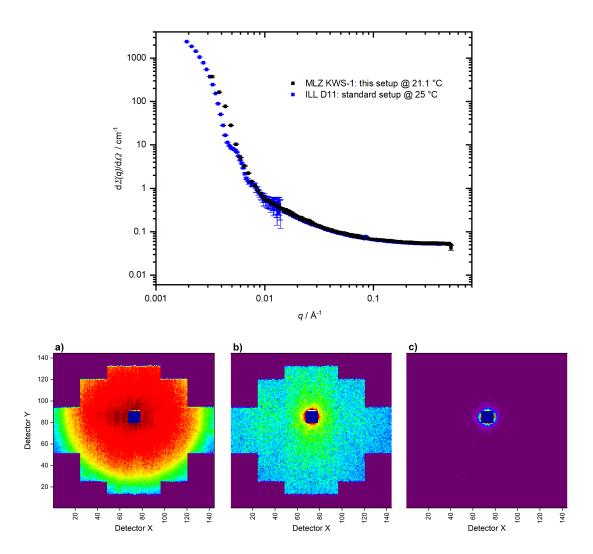


Figure 6. (**top**): SANS data measured employing the new sample environment from this work using the KWS1 machine at MLZ compared to SANS measurements made with the ILL standard setup at the D11 instrument. For both measurements, the same NIPMAM microgel sample as for the DLS measurements was used. (**bottom**): detector images for an empty cell measurement at 2 m (**a**), 5 m (**b**) and 20 m (**c**). The reflections seen here, are caused by the large distance between the nose of the collimation system and the sample cells.

Figure 6, bottom, shows detector images of empty-cell measurements at 2 m, 5 m and 20 m sample–detector distances. Especially for the detector image of the 5 m measurement, an inhomogeneous scattering can be observed. This behavior is attributed to reflections on the sample holder and the walls of the cuvettes which is caused by the rather large distance between the final window of the collimation system and the sample cells and the not fully adapted collimation. This large distance is due to spatial constraints of the sample environment which was not optimized for the KWS-1 machine.

Furthermore, a rather strong background scattering can be observed. This is also due to the relatively long air path which we had to take in the test measurements. This causes the increased background scattering. However, this will not be a problem for the instruments SKADI and LOKI at ESS, due to the optimization of the sample environment for these SANS instruments.

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3. The Fully Assembled Sample Environment

3.1. General Remarks

The in situ SANS/DLS instrument setup was designed with portability and modularity in mind, to guarantee the compatibility with various SANS instruments at the ESS and to enable a fast change of sample environments with only minimal work necessary. For the portability of the setup, all equipment and electronics needed for the light scattering measurements were mounted in 19 inch racks inside a box. This box is constructed from aluminum profiles (Rose und Krieger, Minden, Germany) and fits into the footprint of a EUR-pallet. A photograph of the complete setup can be seen in Figure 7. As versatile interface a 120 cm · 90 cm breadboard (M-SG-34-2, Newport, CA, USA) is mounted on top of the aluminum box. The breadboard is screwed down on the aluminum box to prevent the shifting of the optical setup and to guarantee a stable transport of the complete setup e.g., by forklift or crane. For transport using a crane, a crane eye is installed on each edge of the box. This base construction is identical for all FlexiProb sample environments and can be easily moved to the instrument cave using a pallet jack.

The setup is constructed, so that it can be put into operation with only a few connections made. Power is supplied by a single 32 A IEC 60309 plug. The thermostat cycle needs to be connected and the RS232-cable needs to be connected to the thermostat. Further pressurized dried air, the laser interlock and an Ethernet connection are required for operation. The patch panel layout of the setup is suitable for connectors available at ESS instruments such as SKADI and LOKI.

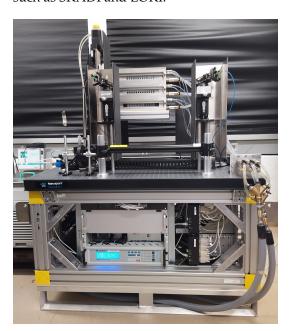


Figure 7. Photograph of the complete SANS/DLS setup with carrier structure.

The general architecture of the interacting systems is described in the scheme in Figure 8. The user interacts with the setup either via the networked instrument control system (NICOS) [42,43] client or the DLS-GUI program, for direct control of the data fits. The measurement is controlled by the NICOS server, where the single NICOS devices can communicate with the input output controllers (IOCs) in the experimental physics and industrial control system (EPICS) [44] server. From the EPICS server, the hardware devices are controlled via their respective protocols, namely USB, Ethernet, an Ethernet-to-RS232-converter (EX-6034, EXSYS, Steinbach, Germany) or by the direct control of the power supply with switchable power outlets (Multibox, ANTRAX Datentechnik, Herford, Germany).

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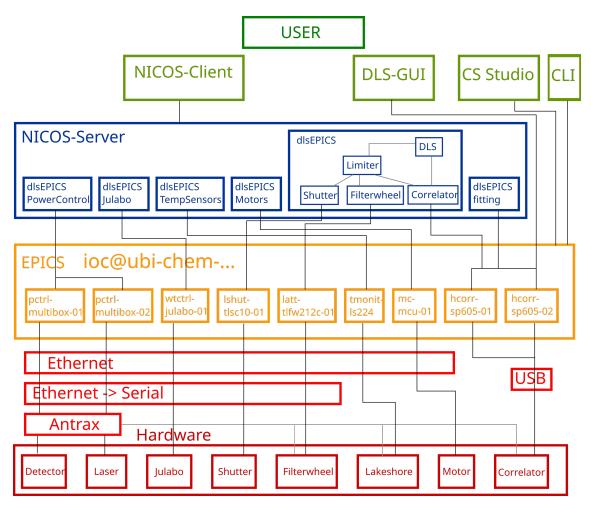


Figure 8. Schematic representation of the architecture of the setup. The user can interact via the NICOS client with the NICOS server [42,43], where the measurements are controlled. Low level-control of the devices is implemented via EPICS, that is connected via different protocols to the hardware. Alternatively, the EPICS process variables can be directly manipulated via the program CS Studio or the command line interface (CLI).

3.1.1. Safety Precautions

For laser safety, a shutter is installed in the setup. The shutter interlock will be connected to the instruments' safety environments' doors, so that no laser radiation can escape during operation. To prevent laser reflexes from the sample holder and physical contact with the cadmium, the Cd-mask in the sample holder is covered by a black polymer film. To prevent the activation of the sample environment from the neutron radiation, the sample rack and magazines are constructed from aluminum, polyether ether ketone (PEEK), and titanium screws. Boron-based neutron shielding (5 mm Mirrobor, Mirrotron, Budapest, Hungary) is applied to avert activation of the sample environment from neutron reflexes originating from the sample. For this, shielding is mounted on the side of the sample holder facing the axis of the translation unit and below the sample holder. As the optical setup could not be realized using only neutron-compatible materials, neutron shielding is also applied around the three optical towers used for light scattering.

3.1.2. Automated Data Analysis

For the analysis of the autocorrelation functions, three methods were implemented in the software. The data can be evaluated during the measurement using inverse Laplace transformation by means of the program CONTIN [45]. Moreover, the standard version of the method of cumulants [46] can be used for fits of the region of short correlation times or a modified method of cumulants can be used for larger times [47,48]. This way, the relaxation

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rate Γ and accordingly, the sample size and stability, can be monitored on-line during the SANS measurement.

Figure 9 shows the GUI of a custom program developed to monitor the measurements and fits during the runtime. Shown is the auto correlation function, the intensity trace (top row), the residuals from the fit and the resulting gamma distribution function (bottom row). This program bypasses the NICOS-server and receives the data directly from the correlators. Therefore, custom fits can also be tried out locally before modifying the settings for the automated fitting. Hence, the sample can be characterized in situ and deterioration can be detected during the course of the SANS measurements.

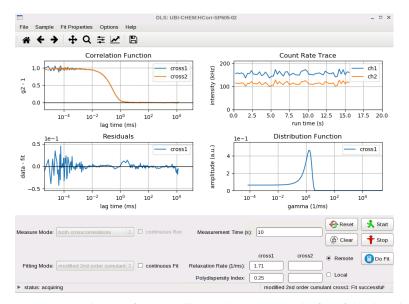


Figure 9. Application for controlling and monitoring the fits of the data during the measurements.

4. Conclusions

A new sample environment for in situ dynamic light scattering and SANS at the ESS was developed. The entire setup was transferred to MLZ and test measurements were performed. The sample environment was used to successfully measure SANS. The background problem will be solved at the ESS using a smaller distance between the individual sample cells and the end window of the collimation system. The simultaneous operation of the DLS system was proven for both configured scattering angles. The sample environments' capabilities regarding temperature control and measurement were also characterized and fulfill all specifications. The integration of all devices in both EPICS and NICOS was successfully finished, making the sample environment ready for integration and first tests at the ESS.

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