



Clinical applications and prospects of PET imaging in patients with IDH-mutant gliomas

Michael M. Wollring^{1,2} · Jan-Michael Werner² · Garry Ceccon² · Philipp Lohmann¹ · Christian P. Filss^{1,3} · Gereon R. Fink^{1,2} · Karl-Josef Langen^{1,3} · Norbert Galldiks^{1,2,4}

Received: 15 November 2022 / Accepted: 14 December 2022 / Published online: 29 December 2022
© The Author(s) 2022

Abstract

PET imaging using radiolabeled amino acids in addition to MRI has become a valuable diagnostic tool in the clinical management of patients with brain tumors. This review provides a comprehensive overview of PET studies in glioma patients with a mutation in the isocitrate dehydrogenase gene (IDH). A considerable fraction of these tumors typically show no contrast enhancement on MRI, especially when classified as grade 2 according to the World Health Organization classification of Central Nervous System tumors. Major diagnostic challenges in this situation are differential diagnosis, target definition for diagnostic biopsies, delineation of glioma extent for treatment planning, differentiation of treatment-related changes from tumor progression, and the evaluation of response to alkylating agents. The main focus of this review is the role of amino acid PET in this setting. Furthermore, in light of clinical trials using IDH inhibitors targeting the mutated IDH enzyme for treating patients with IDH-mutant gliomas, we also aim to give an outlook on PET probes specifically targeting the IDH mutation, which appear potentially helpful for response assessment.

Keywords Amino acid PET · Astrocytoma · Oligodendroglioma · Non-enhancing gliomas · IDH inhibitors

Introduction

With the fifth edition of the World Health Organization (WHO) classification of tumors of the Central Nervous System (CNS) from 2021, gliomas with isocitrate dehydrogenase (IDH) mutations are classified either as diffuse astrocytoma of the WHO CNS grade 2–4, or as oligodendroglioma

of the WHO CNS grade 2–3, if a 1p/19q co-deletion is additionally present [1].

At initial diagnosis, during treatment, and at follow-up of patients with IDH-mutant gliomas, anatomical MRI is the imaging modality of choice given its widespread availability and excellent spatial resolution [2]. In contrast, the specificity of this technique for neoplastic tissue is low, and a disruption of the blood–brain barrier indicated by contrast enhancement is not limited to neoplastic tissue [3–9]. Besides, both contrast enhancement and signal changes in fluid-attenuated inversion recovery (FLAIR) and T2-weighted images may be induced by inflammation, ischemia, or reactive changes following neurooncological therapy [8, 10–12]. Of note, IDH-mutant gliomas of the WHO CNS grade 2 are mostly non-enhancing on MRI and usually evaluated solely based on a hyperintense signal extension of FLAIR- or T2-weighted MRI sequences. Furthermore, even a subgroup of IDH-mutant gliomas of the WHO CNS grades 3 or 4 may also be non-enhancing [13].

These limitations of anatomical MRI may negatively affect a reliable delineation of the spatial extent of non-enhancing glial tumors for planning of a diagnostic biopsy, tumor resection, and other local treatment options such as

Michael M. Wollring and Jan-Michael Werner have contributed equally to this work.

✉ Michael M. Wollring
m.wollring@fz-juelich.de; michael.wollring@uk-koeln.de

¹ Institute of Neuroscience and Medicine (INM-3, -4), Research Center Juelich, Leo-Brandt-St., 52425 Juelich, Germany

² Present Address: Department of Neurology, Faculty of Medicine and University Hospital Cologne, University of Cologne, Kerpener St. 62, 50937 Cologne, Germany

³ Department of Nuclear Medicine, University Hospital Aachen, Aachen, Germany

⁴ Center of Integrated Oncology (CIO), Universities of Aachen, Bonn, Cologne, and Duesseldorf, Cologne, Germany

radiotherapy including the assessment of response [9, 14]. In particular, overlooking the most malignant tumor parts may lead to both inaccurate diagnosis and grading of tumors characterized according to the WHO CNS classification.

To overcome these diagnostic challenges, PET imaging using radiolabeled amino acid tracers has become increasingly important over the past decades [9, 14–16]. Furthermore, the clinical value of this group of tracers, besides of anatomical MRI, has been recommended by the PET task force of the Response Assessment in Neuro-Oncology (RANO) Working Group for the diagnostic management of patients with either gliomas or brain metastases [17, 18].

This review summarizes the value of amino acid PET for the most relevant clinical indications in patients with IDH-mutant gliomas, with particular emphasis on non-enhancing tumors.

Methods

A PubMed search using the search terms “PET”, “glioma”, “amino acid”, “MET”, “FET”, “FDOPA”, “isocitrate dehydrogenase”, “IDH”, “non-enhancing glioma”, “radiotherapy”, “radiation necrosis”, “pseudoprogression”, “tumor extent”, “response assessment”, “treatment-related changes”, “immunotherapy”, and combinations thereof was performed until July 2022.

Amino acid PET

Different PET tracers have been evaluated in patients with gliomas and other brain tumors to visualize and quantify multiple metabolic properties such as glucose consumption, amino acid transport, proliferation, hypoxia, blood flow, and angiogenesis [19]. This section focuses on PET imaging using radiolabeled amino acids, which are increasingly used in the diagnostic management of patients with glioma, especially in Europe.

The most used amino acid tracers for PET imaging to date are O-(2-[^{18}F]-fluoroethyl)-L-tyrosine (FET), [^{11}C]-methyl-L-methionine (MET), and 3,4-dihydroxy-6-[^{18}F]-fluoro-L-phenylalanine (FDOPA). Their uptake is facilitated by large neutral amino acid transporters of the L-type (LAT) in gliomas and brain metastases (i.e., subtypes LAT1 and LAT2), which are regularly overexpressed in both brain tumor types [9, 20–23]. Most early amino acid PET studies were performed using MET, but the short half-life of 20 min imposes logistical challenges, necessitating an onsite cyclotron [24, 25]. The advent of ^{18}F -labeled tracers with a considerably longer half-life (110 min) such as FET and FDOPA allows transport to other facilities and has led to the replacement of MET predominantly by FET, especially in

Europe [9]. Notably, the physiological uptake of FDOPA in the striatum may hamper its use in evaluating tumor extent [9, 26].

In recent years, the synthetic amino acid analog anti-1-amino-3-[^{18}F]fluorocyclobutane-1-carboxylic acid (Fluciclovine) has gained clinical interest particularly for imaging of primary and secondary brain tumors. Intratumoral transport of Fluciclovine seems to be mediated by LAT1, but predominantly by the neutral alanine, serine, cysteine transporter 2 (ASCT2), another neutral amino acid transporter [27].

Most important clinical applications

Characterization of newly diagnosed non-enhancing brain lesions for differential diagnosis using amino acid PET

Brain lesions presenting hyperintense FLAIR signal alterations on anatomical MRI without concomitant contrast enhancement may frequently suggest an IDH-mutant non-enhancing glioma. On the other hand, these lesions constitute a heterogeneous group of diseases including non-neoplastic lesions such as cerebral hematoma, ischemic lesions, inflammatory or infectious processes, and even malignant gliomas without disrupted blood–brain barrier. False interpretation of these findings may result in necessary treatment being deferred or unnecessarily indicated.

Amino acid PET has a sensitivity of more than 90% to detect gliomas [28–30], but grade 2 gliomas characterized according to older WHO classifications of tumors of the CNS [31, 32] exhibit increased tracer uptake only in 70–80% [28–30]. The remaining gliomas are not avid on amino acid PET (i.e., no increased uptake compared to the unaffected brain tissue). Of note, a subgroup of patients without FET uptake in brain lesions with MRI findings suspicious for low-grade gliomas (i.e., hyperintense T2/FLAIR lesion without contrast enhancement) may even show photopenic defects on FET PET (i.e., FET uptake visually lower than the healthy background uptake) and harbor malignant gliomas [33]. This phenomenon has also been described for the radiolabeled amino acids MET and FDOPA [34].

Regarding the diagnostic performance of FET PET for differential diagnosis, a meta-analysis evaluating 13 studies with a total of 462 patients reported a specificity of 76% and a sensitivity of 82% for differentiating primary brain tumors from non-tumoral lesions [35]. A subsequent FET PET study including 174 patients ($n = 73$ patients with WHO CNS grade 2 gliomas, 75% of these without contrast enhancement) at initial diagnosis of cerebral lesions suggestive of glioma yielded a higher specificity

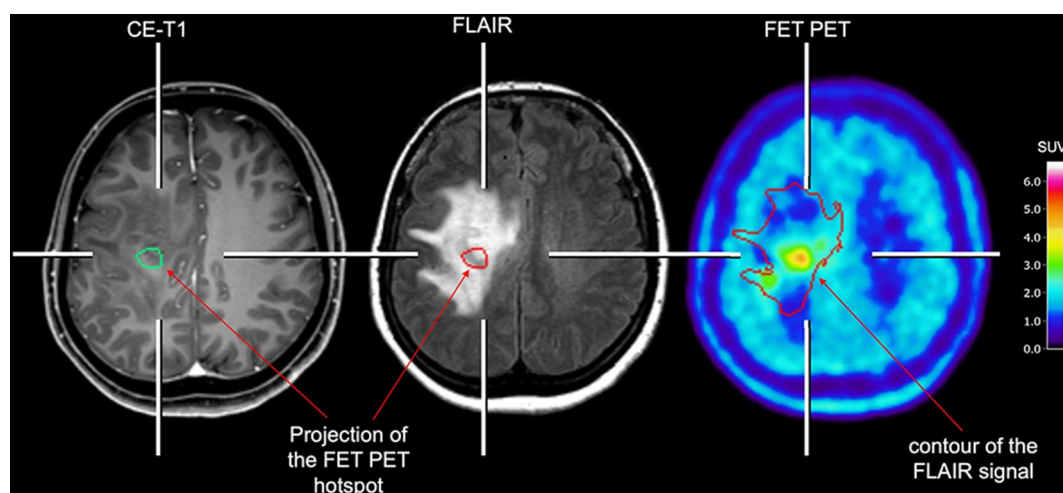


Fig. 1 Contrast-enhanced MRI and FET PET of a 49-year-old female patient with a large FLAIR-hyperintense lesion without contrast enhancement in the right central cortex suggesting a glioma. Due to the widespread FLAIR signal, a definite biopsy target is lacking. The additional FET PET scan shows a localized area with pathologically increased metabolic activity (maximum tumor-to-brain ratio,

3.3), considerably smaller than the hyperintense FLAIR signal (red contour), and thereby offers a target for diagnostic biopsy. After FET PET-guided stereotactic biopsy of the metabolically active lesion, neuropathological evaluation of the obtained tissue revealed an IDH-mutant astrocytoma, WHO CNS grade 2

(92%) and a positive predictive value for glioma tissue of 98% using a maximum tumor-to-brain ratio of 2.5 [36].

For differential diagnosis within the subgroup of gliomas, patients with oligodendroglioma characterized by an IDH mutation and a 1p/19q co-deletion [1] frequently exhibit a considerably higher amino acid uptake than patients with IDH-mutant astrocytomas [37]. For example, a more recent MET PET study by Kim and co-workers reported that in 74 patients with IDH-mutant gliomas of the WHO CNS grade 2 or 3, oligodendrogliomas had significantly higher average median tumor-to-brain ratios than astrocytomas (2.90 vs. 1.40; $P < 0.001$), but not higher than in patients with IDH-wildtype glioma ($n = 70$) (averaged median tumor-to-brain ratio, 3.35) [38].

In summary, amino acid PET may add differential diagnostic information in patients with brain lesions suggestive of non-enhancing glioma. Of note, available studies do not primarily focus on patients with IDH-mutant gliomas. Thus, the impact of this genetic alteration on the diagnostic performance of amino acid PET warrants further investigation.

Target definition for diagnostic biopsy and delineation of glioma extent for treatment planning using amino acid PET

Particularly in patients with non-enhancing gliomas, it is difficult to identify the biopsy target including the most

malignant tumor parts using anatomical MRI [39–41], especially when a widespread T2/FLAIR signal on MRI is present [42]. For PET, several studies have correlated histomolecular findings obtained from tissue specimens with imaging findings on amino acid PET, predominantly using the tracers FET and MET, and provided evidence that this technique detects the solid mass of gliomas including most malignant parts and the metabolically active tumor more reliably than conventional MRI [43–49] Fig. 1. Therefore, amino acid PET appears to be a highly valuable tool for target definition. In addition, combining amino acid PET with advanced MRI techniques such as diffusion- or perfusion-weighted MR imaging may further improve target definition for diagnostic biopsy planning [47, 50]. Initial studies suggest that also the synthetic amino acid analog Fluciclovine accumulates in non-enhancing gliomas and identifies infiltrating tumor areas without contrast enhancement on MRI [51, 52]. Considering the significantly higher tumor-to-brain contrast of Fluciclovine compared to other amino acid tracers [53], this tracer may be of additional value for the delineation of tumor extent and target definition in this group of patients.

Using dynamic FET PET acquisition, additional imaging parameters derived from time-activity curves such as time-to-peak values (i.e., time from tracer injection to maximum tracer uptake) or quantitative approaches to characterize time-activity curve patterns (e.g., calculation of slope) can be obtained [54]. For instance, the

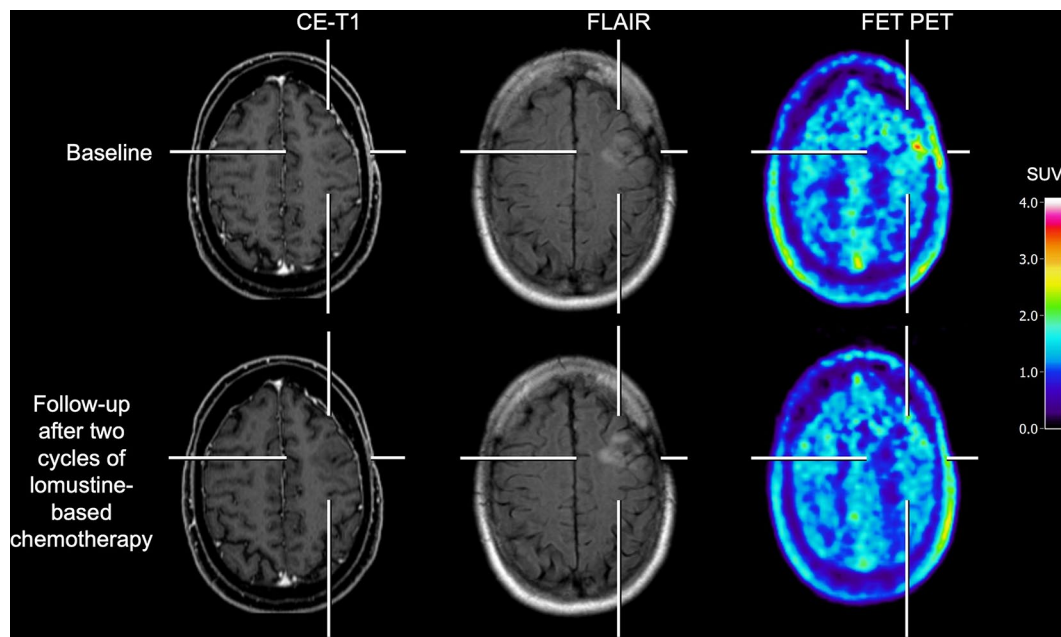


Fig. 2 Contrast-enhanced MRI and FET PET of a 52-year-old male patient with a left frontal IDH-mutant, 1p/19q-codeleted oligodendroglioma of the WHO CNS grade 2 after resection and radiotherapy (top row). The corresponding FET PET scan shows residual metabolic activity (maximum tumor-to-brain ratio, 2.1). After two cycles of adjuvant chemotherapy with procarbazine and lomustine, the fol-

low-up MRI (bottom row) shows an increasing FLAIR signal alteration without contrast enhancement, suggesting tumor progression. In contrast, the follow-up FET PET shows no increased metabolic activity, indicating a metabolic response. During adjuvant chemotherapy, the hyperintense FLAIR signal regressed partially, and the patient was free of tumor progression for more than two years

information derived from dynamic FET PET parameters may also help identify most malignant tumor parts in suspected gliomas, thus offering an additional tool to define the biopsy target [39–41].

Furthermore, accurate delineation of tumor extent to ensure a maximal resection is also of particular interest in patients with IDH-mutant gliomas, as even minimal tumor remnants after surgery may negatively impact overall survival [55]. A recent study by Ninatti et al. compared the additional value of preoperative MET PET for delineating tumor extent in patients with IDH-mutant gliomas (i.e., oligodendrogliomas and astrocytomas of the WHO CNS grade 2 or 3) with anatomical MRI alone [56]. In that study, MET PET improved the target volume for surgical resection in 28 of 153 patients (25%). Moreover, in patients with IDH-mutant astrocytomas, higher maximum tumor-to-brain ratios on preoperative MET PET were independent predictors of shorter progression-free survival [56].

Differentiation of treatment-related changes from glioma progression using amino acid PET

The differentiation of treatment-related changes such as pseudoprogression or radiation necrosis from tumor progression is of utmost importance in clinical routine. Especially

amino acid PET using FET [57–65] or FDOPA [66–68] achieved a high diagnostic accuracy for differentiating treatment-related changes from tumor progression in glioma patients Fig. 2. Of note, these studies have been performed primarily on IDH-wildtype glioblastoma patients. In more recent studies, the diagnostic performance of amino acid PET for this indication has also been evaluated in patients with IDH-mutant gliomas. In a study including 127 patients (48% of patients had IDH-mutant gliomas), Maurer and colleagues reported that the combined analysis of static and dynamic FET PET parameters achieved an overall accuracy of 81% for the differentiation of treatment-related changes from tumor progression [69]. Interestingly, a subgroup analysis suggested that the diagnostic accuracy was only 67% in patients with IDH-mutant gliomas compared to 91% in IDH-wildtype glioma patients [69]. On the other hand, in subsequent studies the latter finding could be confirmed only partially [70–72], indicating that further studies in this subgroup of patients with IDH-mutant gliomas are warranted.

Assessment of response to alkylating chemotherapy using amino acid PET

Two studies evaluated the response to temozolomide chemotherapy in patients with previously untreated WHO grade 2 gliomas classified according to older WHO classifications

[31, 32] using serial MET PET or FET PET compared to anatomical MRI [73, 74]. In one of these studies, the IDH mutational status was partially available (67% of patients had a confirmed IDH mutation) [74]. A metabolic response, defined as a decrease of metabolically active tumor volume of more than 10% was observed in 8 of 11 patients (73%), and in 25 of 33 patients (76%), respectively [73, 74]. Notably, MET and FET PET detected treatment response significantly earlier than FLAIR signal changes on MRI. Moreover, a reduced metabolic tumor volume was associated with a significantly longer progression-free survival and an improved seizure control [74].

In a more recent study by Suchorska et al. [75] FET PET and MRI were used to assess response to temozolomide or lomustine-based regimens in 61 patients with non-enhancing gliomas, including 19 patients with IDH-mutant, 1p/19q-non-codeleted gliomas (31%), and 30 IDH-mutant and 1p/19q-codeleted gliomas (49%). Metabolic response, defined as any decrease of the metabolic tumor volume, was observed in 34 patients (range of decrease relative to baseline, 10–25%). These patients had a significantly longer time-to-treatment failure than patients with stable or increasing metabolic tumor volumes (median time, 78.5 vs. 24.1 months; $P=0.001$). On the other hand, signal changes on T2-weighted MRI did not correlate with the patients' survival.

Future prospects

PET imaging of the IDH mutation

In patients with IDH-mutant gliomas, the efficacy of novel therapies targeting the mutant IDH enzyme using oral IDH1-inhibitors (e.g., ivosidenib), IDH2-inhibitors such as endasidenib, inhibitors of both IDH1 and IDH2 mutations (e.g., vorasidenib), and vaccines targeting the IDH1 (R132H) neoepitope are currently under investigation. Initial phase I clinical trials suggested promising antitumoral activity [76–78]. At initial diagnosis, neuropathological diagnostics, including genomic sequencing, is currently the method of choice for detecting an IDH mutation. During follow-up, quantification of changes of IDH expression levels in patients undergoing these treatment options using neuropathological techniques always requires the invasive tissue removal. Alternatively, the use of proton MR spectroscopy for the non-invasive evaluation of signal changes of the oncometabolite 2-hydroxyglutarate related to IDH mutations is an option for response assessment, but this technique is highly prone to susceptibility artefacts due to bone, hemorrhage, calcifications, or surgical material and may even be false-positive in 20% of patients with newly diagnosed IDH-wildtype glioblastoma [79, 80].

Regarding PET imaging of IDH mutations, novel PET tracers such as radiolabeled triazinediamine or butyl-phenyl sulfonamide analogs, and the radiolabeled IDH1 inhibitor AGI-5198 may be valuable candidates [81–83]. Furthermore, in an animal study, Koyaso and colleagues observed that the uptake of ^{14}C -labeled acetate is significantly higher in IDH-mutant cells than in IDH-wildtype cells [84]. An initial clinical study in 28 glioma patients reported similar results for the differentiation between the IDH-mutant and IDH-wildtype genotype using the radiotracer ^{18}F -fluoroethylcholine [85]. Overall, further development of these tracers is warranted, primarily when IDH inhibitors are used in clinical routine.

Conclusions

Amino acid PET has become increasingly relevant in the clinical care of patients with gliomas. While most amino acid PET studies focused mainly on IDH-wildtype glioma patients, the available literature regarding its use in patients with IDH-mutant gliomas suggests that this technique also adds valuable clinical information for decision-making. In particular, this has been demonstrated for detecting the most malignant tumor parts, delineating glioma extent, diagnosing treatment-related changes, and assessing treatment response in IDH-mutant glioma patients.

Nevertheless, it has to be pointed out that in a considerable fraction of available studies, the neuropathological characterization of the patients' gliomas is either based on older WHO classifications for CNS tumors (i.e., solely on histology) or is only partially histomolecularly characterized according to current classifications (i.e., according to the WHO classification for CNS tumors from 2016 or 2021). Thus, further studies in more homogenous patient groups with well-defined glioma characteristics in line with the latest 2021 WHO classification for CNS tumors, preferably in a prospective setting, are warranted.

One prospect is the specific PET imaging of IDH mutations for response assessment in patients undergoing IDH-targeted therapies. The added clinical value is related to the considerably increased specificity of these PET probes for IDH mutations. Therefore, it offers a more reliable response evaluation since FLAIR signal alterations may be unspecific for neoplastic tissue, and 2-hydroxyglutarate MR spectroscopy is highly susceptible to artifacts.

Author contributions MMW, NG: Conceptualization, visualization, writing original draft. JMW: Visualization, writing original draft. PL, GC, CPF, GRF, KJL: editing the original draft. All authors reviewed the manuscript.

Funding Open Access funding enabled and organized by Projekt DEAL. The Cologne Clinician Scientist Program (CCSP) of the Deutsche Forschungsgemeinschaft (DFG, FI 773/15-1), Germany, supported this work.

Declarations

Conflict of interest Norbert Galldiks and Philipp Lohmann received honoraria for lectures from Blue Earth Diagnostics. The other authors disclosed no potential conflicts of interest related to the present work.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

References

- Louis DN, Perry A, Wesseling P et al (2021) The 2021 WHO classification of tumors of the central nervous system: a summary. *Neuro Oncol* 23(8):1231–1251
- Weller M, van den Bent M, Preusser M et al (2021) EANO guidelines on the diagnosis and treatment of diffuse gliomas of adulthood. *Nat Rev Clin Oncol* 18(3):170–186
- Lohmann P, Stavrinou P, Lipke K et al (2019) FET PET reveals considerable spatial differences in tumour burden compared to conventional MRI in newly diagnosed glioblastoma. *Eur J Nucl Med Mol Imaging* 46(3):591–602
- Dhermain FG, Hau P, Lanfermann H, Jacobs AH, van den Bent MJ (2010) Advanced MRI and PET imaging for assessment of treatment response in patients with gliomas. *Lancet Neurol* 9(9):906–920
- Brandsma D, Stalpers L, Taal W, Sminia P, van den Bent MJ (2008) Clinical features, mechanisms, and management of pseudoprogression in malignant gliomas. *Lancet Oncol* 9(5):453–461
- Hygino da Cruz LC, Rodriguez I, Domingues RC, Gasparetto EL, Sorensen AG (2011) Pseudoprogression and pseudoresponse: imaging challenges in the assessment of posttreatment glioma. *Am J Neuroradiol* 32:1978–1985
- Yang I, Aghi MK (2009) New advances that enable identification of glioblastoma recurrence. *Nat Rev Clin Oncol* 6:648–657
- Kumar AJ, Leeds NE, Fuller GN et al (2000) Malignant gliomas: MR imaging spectrum of radiation therapy- and chemotherapy-induced necrosis of the brain after treatment. *Radiology* 217(2):377–384
- Langen KJ, Galldiks N, Hattingen E, Shah NJ (2017) Advances in neuro-oncology imaging. *Nat Rev Neurol* 13(5):279–289
- Wen PY, Macdonald DR, Reardon DA et al (2010) Updated response assessment criteria for high-grade gliomas: response assessment in neuro-oncology working group. *J Clin Oncol* 28(11):1963–1972
- Dhermain FG, Hau P, Lanfermann H, Jacobs AH, van den Bent MJ (2010) Advanced MRI and PET imaging for assessment of treatment response in patients with gliomas. *Lancet Neurol* 9(9):906–920
- Ahluwalia MS, Wen PY (2011) Antiangiogenic therapy for patients with glioblastoma: current challenges in imaging and future directions. *Expert Rev Anticancer Ther* 11(5):653–656
- Eichberg DG, Di L, Morell AA et al (2020) Incidence of high grade gliomas presenting as radiographically non-enhancing lesions: experience in 111 surgically treated non-enhancing gliomas with tissue diagnosis. *J Neurooncol* 147(3):671–679
- Werner JM, Stoffels G, Lichtenstein T et al (2019) Differentiation of treatment-related changes from tumour progression: a direct comparison between dynamic FET PET and ADC values obtained from DWI MRI. *Eur J Nucl Med Mol Imaging* 46(9):1889–1901
- Nandu H, Wen PY, Huang RY (2018) Imaging in neuro-oncology. *Ther Adv Neurol Disord* 11:1756286418759865
- Galldiks N, Kocher M, Ceccon G et al (2020) Imaging challenges of immunotherapy and targeted therapy in patients with brain metastases: response, progression, and pseudoprogression. *Neuro Oncol* 22(1):17–30
- Galldiks N, Langen KJ, Albert NL et al (2019) PET imaging in patients with brain metastasis-report of the RANO/PET group. *Neuro Oncol* 21(5):585–595
- Albert NL, Weller M, Suchorska B et al (2016) Response Assessment in Neuro-Oncology working group and European Association for Neuro-Oncology recommendations for the clinical use of PET imaging in gliomas. *Neuro Oncol* 18(9):1199–1208
- Werner JM, Lohmann P, Fink GR, Langen KJ, Galldiks N (2020) Current landscape and emerging fields of pet imaging in patients with brain tumors. *Molecules* 25(6):1471
- Youland RS, Kitange GJ, Peterson TE et al (2013) The role of LAT1 in (18)F-DOPA uptake in malignant gliomas. *J Neurooncol* 111(1):11–18
- Papin-Michault C, Bonnetaud C, Dufour M et al (2016) Study of LAT1 expression in brain metastases: towards a better understanding of the results of positron emission tomography using amino acid tracers. *PLoS ONE* 11(6):e0157139
- Wiriyaerkmul P, Nagamori S, Tominaga H et al (2012) Transport of 3-fluoro-L-alpha-methyl-tyrosine by tumor-upregulated L-type amino acid transporter 1: a cause of the tumor uptake in PET. *J Nucl Med* 53(8):1253–1261
- Okubo S, Zhen HN, Kawai N, Nishiyama Y, Haba R, Tamiya T (2010) Correlation of L-methyl-11C-methionine (MET) uptake with L-type amino acid transporter 1 in human gliomas. *J Neurooncol* 99(2):217–225
- Langen KJ, Hamacher K, Weckesser M et al (2006) O-(2-[18F] fluoroethyl)-L-tyrosine: uptake mechanisms and clinical applications. *Nucl Med Biol* 33(3):287–294
- Galldiks N, Law I, Pope WB, Arbizu J, Langen KJ (2017) The use of amino acid PET and conventional MRI for monitoring of brain tumor therapy. *Neuroimage Clin* 13:386–394
- Cicone F, Filss CP, Minniti G et al (2015) Volumetric assessment of recurrent or progressive gliomas: comparison between F-DOPA PET and perfusion-weighted MRI. *Eur J Nucl Med Mol Imaging* 42(6):905–915
- Ono M, Oka S, Okudaira H et al (2013) Comparative evaluation of transport mechanisms of trans-1-amino-3-[(1)(8)F]fluorocyclobutanecarboxylic acid and L-[methyl-(1)(1)C]methionine in human glioma cell lines. *Brain Res* 1535:24–37
- Hutterer M, Nowosielski M, Putzer D et al (2013) [18F]-fluoroethyl-L-tyrosine PET: a valuable diagnostic tool in neuro-oncology, but not all that glitters is glioma. *Neuro Oncol* 15(3):341–351
- Jansen NL, Graute V, Armbruster L et al (2012) MRI-suspected low-grade glioma: is there a need to perform dynamic FET PET? *Eur J Nucl Med Mol Imaging* 39(6):1021–1029
- Pichler R, Dunzinger A, Wurm G et al (2010) Is there a place for FET PET in the initial evaluation of brain lesions with unknown significance? *Eur J Nucl Med Mol Imaging* 37:1521–1528

31. Louis DN, Ohgaki H, Wiestler OD et al (2007) The 2007 WHO classification of tumours of the central nervous system. *Acta Neuropathol* 114(2):97–109
32. Louis DN, Perry A, Reifenberger G et al (2016) The 2016 World Health Organization classification of tumors of the central nervous system: a summary. *Acta Neuropathol* 131(6):803–820
33. Galldiks N, Unterrainer M, Judov N et al (2019) Photopenic defects on O-(2-[18F]-fluoroethyl)-L-tyrosine PET: clinical relevance in glioma patients. *Neuro Oncol* 21(10):1331–1338
34. Zaragori T, Castello A, Guedj E et al (2021) Photopenic Defects in Gliomas With Amino-Acid PET and Relative Prognostic Value: A Multicentric 11C-Methionine and 18F-FDOPA PET Experience. *Clin Nucl Med* 46(1):e36–e37
35. Dunet V, Rossier C, Buck A, Stupp R, Prior JO (2012) Performance of 18F-fluoro-ethyl-tyrosine (18F-FET) PET for the differential diagnosis of primary brain tumor: a systematic review and Metaanalysis. *J Nucl Med* 53(2):207–214
36. Rapp M, Heinzel A, Galldiks N et al (2013) Diagnostic performance of 18F-FET PET in newly diagnosed cerebral lesions suggestive of glioma. *J Nucl Med* 54(2):229–235
37. Takei H, Shinoda J, Ikuta S, Usefulness of positron emission tomography for differentiating gliomas according to the, et al (2016) World Health Organization classification of tumors of the central nervous system. *J Neurosurg* 2019:1–10
38. Kim D, Chun JH, Kim SH et al (2019) Re-evaluation of the diagnostic performance of (11)C-methionine PET/CT according to the 2016 WHO classification of cerebral gliomas. *Eur J Nucl Med Mol Imaging* 46(8):1678–1684
39. Kunz M, Thon N, Eigenbrod S et al (2011) Hot spots in dynamic (18)FET-PET delineate malignant tumor parts within suspected WHO grade II gliomas. *Neuro Oncol* 13:307–316
40. Thon N, Kunz M, Lemke L et al (2015) Dynamic 18F-FET PET in suspected WHO grade II gliomas defines distinct biological subgroups with different clinical courses. *Int J Cancer* 136(9):2132–2145
41. Kunz M, Albert NL, Unterrainer M et al (2019) Dynamic 18F-FET PET is a powerful imaging biomarker in gadolinium-negative gliomas. *Neuro Oncol* 21(2):274–284
42. Galldiks N, Brunn A, Fink GR, Langen KJ (2019) Dynamic FET PET Imaging of a “Butterfly” IDH-Wildtype Anaplastic Astrocytoma. *Clin Nucl Med* 44(10):e581–e582
43. Kracht LW, Miletic H, Busch S et al (2004) Delineation of brain tumor extent with [11C]L-methionine positron emission tomography: local comparison with stereotactic histopathology. *Clin Cancer Res* 10(21):7163–7170
44. Pauleit D, Floeth F, Hamacher K et al (2005) O-(2-[18F] fluoroethyl)-L-tyrosine PET combined with MRI improves the diagnostic assessment of cerebral gliomas. *Brain* 128(3):678–687
45. Pafundi DH, Laack NN, Youland RS et al (2013) Biopsy validation of 18F-DOPA PET and biodistribution in gliomas for neurosurgical planning and radiotherapy target delineation: results of a prospective pilot study. *Neuro Oncol* 15(8):1058–1067
46. Roodakker KR, Alhuseinalkhudhur A, Al-Jaff M et al (2019) Region-by-region analysis of PET, MRI, and histology in en bloc-resected oligodendrogliomas reveals intra-tumoral heterogeneity. *Eur J Nucl Med Mol Imaging* 46(3):569–579
47. Verbarg N, Koopman T, Yaqub MM et al (2020) Improved detection of diffuse glioma infiltration with imaging combinations: a diagnostic accuracy study. *Neuro Oncol* 22(3):412–422
48. Schön S, Cabello J, Liesche-Starnecker F et al (2020) Imaging glioma biology: spatial comparison of amino acid PET, amide proton transfer, and perfusion-weighted MRI in newly diagnosed gliomas. *Eur J Nucl Med Mol Imaging* 47(6):1468–1475
49. Pauleit D, Floeth F, Hamacher K et al (2005) O-(2-[18F] fluoroethyl)-L-tyrosine PET combined with MRI improves the diagnostic assessment of cerebral gliomas. *Brain* 128(Pt 3):678–687
50. Girard A, Le Reste PJ, Metais A et al (2021) Combining (18) F-DOPA PET and MRI with perfusion-weighted imaging improves delineation of high-grade subregions in enhancing and non-enhancing gliomas prior treatment: a biopsy-controlled study. *J Neurooncol* 155(3):287–295
51. Tsuyuguchi N, Terakawa Y, Uda T, Nakajo K, Kanemura Y (2017) Diagnosis of Brain Tumors Using Amino Acid Transport PET Imaging with (18)F-fluciclovine: A Comparative Study with L-methyl-(11)C-methionine PET Imaging. *Asia Ocean J Nucl Med Biol* 5(2):85–94
52. Wakabayashi T, Iuchi T, Tsuyuguchi N et al (2017) Diagnostic Performance and Safety of Positron Emission Tomography Using (18)F-Fluciclovine in Patients with Clinically Suspected High- or Low-grade Gliomas: A Multicenter Phase IIb Trial. *Asia Ocean J Nucl Med Biol* 5(1):10–21
53. Albano D, Tomasini D, Bonu M, Giubbini R, Bertagna F (2020) (18)F-Fluciclovine ((18)F-FACBC) PET/CT or PET/MRI in gliomas/glioblastomas. *Ann Nucl Med* 34(2):81–86
54. Cecon G, Lohmann P, Stoffels G et al (2017) Dynamic O-(2-[18F]-fluoroethyl)-L-tyrosine positron emission tomography differentiates brain metastasis recurrence from radiation injury after radiotherapy. *Neuro Oncol* 19(2):281–288
55. Wijnenga MMJ, French PJ, Dubbink HJ et al (2018) The impact of surgery in molecularly defined low-grade glioma: an integrated clinical, radiological, and molecular analysis. *Neuro Oncol* 20(1):103–112
56. Ninatti G, Sollini M, Bono B et al (2022) Preoperative [11C] methionine PET to personalize treatment decisions in patients with lower-grade gliomas. *Neuro Oncol* 24(9):1546–1556
57. Pyka T, Hiob D, Preibisch C et al (2018) Diagnosis of glioma recurrence using multiparametric dynamic 18F-fluoroethyl-tyrosine PET-MRI. *Eur J Radiol* 103:32–37
58. Bashir A, Mathilde Jacobsen S, Molby Henriksen O et al (2019) Recurrent glioblastoma versus late posttreatment changes: diagnostic accuracy of O-(2-[18F]fluoroethyl)-L-tyrosine positron emission tomography (18F-FET PET). *Neuro Oncol* 21(12):1595–1606
59. Kebir S, Fimmers R, Galldiks N et al (2016) Late pseudoprogression in glioblastoma: diagnostic value of dynamic O-(2-[18F] fluoroethyl)-L-tyrosine PET. *Clin Cancer Res* 22(9):2190–2196
60. Werner JM, Weller J, Cecon G et al (2021) Diagnosis of Pseudoprogression following lomustine-temozolomide chemoradiation in newly diagnosed glioblastoma patients using FET-PET. *Clin Cancer Res* 27(13):3704–3713
61. Galldiks N, Dunkl V, Stoffels G et al (2015) Diagnosis of pseudoprogression in patients with glioblastoma using O-(2-[18F] fluoroethyl)-L-tyrosine PET. *Eur J Nucl Med Mol Imaging* 42(5):685–695
62. Mihovilovic MI, Kertels O, Hanscheid H et al (2019) O-(2-((18) F)fluoroethyl)-L-tyrosine PET for the differentiation of tumour recurrence from late pseudoprogression in glioblastoma. *J Neurol Neurosurg Psychiatry* 90(2):238–239
63. Galldiks N, Stoffels G, Filss C et al (2015) The use of dynamic O-(2-[18F]-fluoroethyl)-L-tyrosine PET in the diagnosis of patients with progressive and recurrent glioma. *Neuro Oncol* 17(9):1293–1300
64. Kim SJ, Ryul SS (2019) Diagnostic value of radiolabeled amino acid PET for detection of pseudoprogression of brain tumor after treatment: a meta-analysis. *Nucl Med Commun* 40(9):965–972
65. Lohmann P, Elahmadawy MA, Gutsche R et al (2020) FET PET Radiomics for differentiating pseudoprogression from early tumor progression in glioma patients post-chemoradiation. *Cancers (Basel)*. <https://doi.org/10.3390/cancers12123835>

66. Karunanithi S, Sharma P, Kumar A et al (2013) 18F-FDOPA PET/CT for detection of recurrence in patients with glioma: prospective comparison with 18F-FDG PET/CT. *Eur J Nucl Med Mol Imaging* 40(7):1025–1035
67. Karunanithi S, Sharma P, Kumar A et al (2013) Comparative diagnostic accuracy of contrast-enhanced MRI and (18)F-FDOPA PET-CT in recurrent glioma. *Eur Radiol* 23(9):2628–2635
68. Herrmann K, Czernin J, Cloughesy T et al (2014) Comparison of visual and semiquantitative analysis of 18F-FDOPA-PET/CT for recurrence detection in glioblastoma patients. *Neuro Oncol* 16(4):603–609
69. Maurer GD, Brucker DP, Stoffels G et al (2020) (18)F-FET PET imaging in differentiating glioma progression from treatment-related changes: a single-center experience. *J Nucl Med* 61(4):505–511
70. Celli M, Caroli P, Amadori E et al (2021) Diagnostic and Prognostic potential of (18)F-FET PET in the differential diagnosis of glioma recurrence and treatment-induced changes after chemoradiation therapy. *Front Oncol* 11:721821
71. Skoblar Vidmar M, Doma A, Smrdel U, Zevnik K, Studen A (2022) The Value of FET PET/CT in recurrent glioma with a different IDH mutation status: the relationship between imaging and molecular biomarkers. *Int J Mol Sci* 23(12):6787
72. Steidl E, Langen KJ, Hmeidan SA et al (2021) Sequential implementation of DSC-MR perfusion and dynamic [(18)F] FET PET allows efficient differentiation of glioma progression from treatment-related changes. *Eur J Nucl Med Mol Imaging* 48(6):1956–1965
73. Wyss M, Hofer S, Bruehlmeier M et al (2009) Early metabolic responses in temozolomide treated low-grade glioma patients. *J Neurooncol* 95(1):87–93
74. Roelcke U, Wyss MT, Nowosielski M et al (2016) Amino acid positron emission tomography to monitor chemotherapy response and predict seizure control and progression-free survival in WHO grade II gliomas. *Neuro Oncol* 18(5):744–751
75. Suchorska B, Unterrainer M, Biczok A et al (2018) (18) F-FET-PET as a biomarker for therapy response in non-contrast enhancing glioma following chemotherapy. *J Neurooncol* 139(3):721–730
76. Mellinghoff IK, Penas-Prado M, Peters KB et al (2021) Vorasidenib, a dual inhibitor of mutant IDH1/2, in Recurrent or progressive glioma; results of a first-in-human phase I trial. *Clin Cancer Res* 27(16):4491–4499
77. Mellinghoff IK, Ellingson BM, Touat M et al (2020) Ivosidenib in isocitrate dehydrogenase 1-mutated advanced glioma. *J Clin Oncol* 38(29):3398–3406
78. Platten M, Bunse L, Wick A et al (2021) A vaccine targeting mutant IDH1 in newly diagnosed glioma. *Nature* 592(7854):463–468
79. Suh CH, Kim HS, Paik W et al (2019) False-positive measurement at 2-hydroxyglutarate MR spectroscopy in isocitrate dehydrogenase wild-type glioblastoma: a multifactorial analysis. *Radiology* 291(3):752–762
80. Choi C, Ganji SK, DeBerardinis RJ et al (2012) 2-hydroxyglutarate detection by magnetic resonance spectroscopy in IDH-mutated patients with gliomas. *Nat Med* 18(4):624–629
81. Chitneni SK, Reitman ZJ, Gooden DM, Yan H, Zalutsky MR (2016) Radiolabeled inhibitors as probes for imaging mutant IDH1 expression in gliomas: Synthesis and preliminary evaluation of labeled butyl-phenyl sulfonamide analogs. *Eur J Med Chem* 119:218–230
82. Chitneni SK, Yan H, Zalutsky MR (2018) Synthesis and evaluation of a (18)F-labeled triazinediamine analogue for imaging mutant IDH1 expression in gliomas by PET. *ACS Med Chem Lett* 9(7):606–611
83. Chitneni SK, Reitman ZJ, Spicehandler R, Gooden DM, Yan H, Zalutsky MR (2018) Synthesis and evaluation of radiolabeled AGI-5198 analogues as candidate radiotracers for imaging mutant IDH1 expression in tumors. *Bioorg Med Chem Lett* 28(4):694–699
84. Koyasu S, Shimizu Y, Morinibu A et al (2019) Increased (14) C-acetate accumulation in IDH-mutated human glioblastoma: implications for detecting IDH-mutated glioblastoma with (11) C-acetate PET imaging. *J Neurooncol* 145(3):441–447
85. Kong Z, Zhang Y, Liu D et al (2021) Role of traditional CHO PET parameters in distinguishing IDH, TERT and MGMT alterations in primary diffuse gliomas. *Ann Nucl Med* 35(4):493–503

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.